

# Anthocyanin profiles of colored wheat crosses via HPTLC

## Highlights

- Method development for anthocyanin analysis of wheat crosses → simple extraction of the wheat → challenging matrix
- Pronounced differences in the anthocyanin fingerprints of colored wheat extracts
- Fast visual comparison of combinations of blue aleurone and purple pericarp genotypes
- Further characterization via derivatizations, evaluation of anthocyanin sugar components and HPTLC-UV/Vis-MS of anthocyanins

## 2. Characterization via MS

For each genotype, major anthocyanin zones were further analyzed via HPTLC-ESI<sup>+</sup>-MS (Fig. 2). The purple-grained variety (no. 6) showed four zones ( $hR_F \geq 42$ ; A-D) and the blue-grained variety (no. 16) three zones ( $hR_F \leq 42$ ; A-C). The zone at  $hR_F$  42 was found in both varieties (no. 6D and 16A). Mass spectra of both zones showed the molecular ion of cyanidin  $[M]^+$  at  $m/z$  287, suggesting a cyanidin-backbone. With the cyanidin-3-glycoside standard this was confirmed via comparison of colors and  $hR_F$  values.

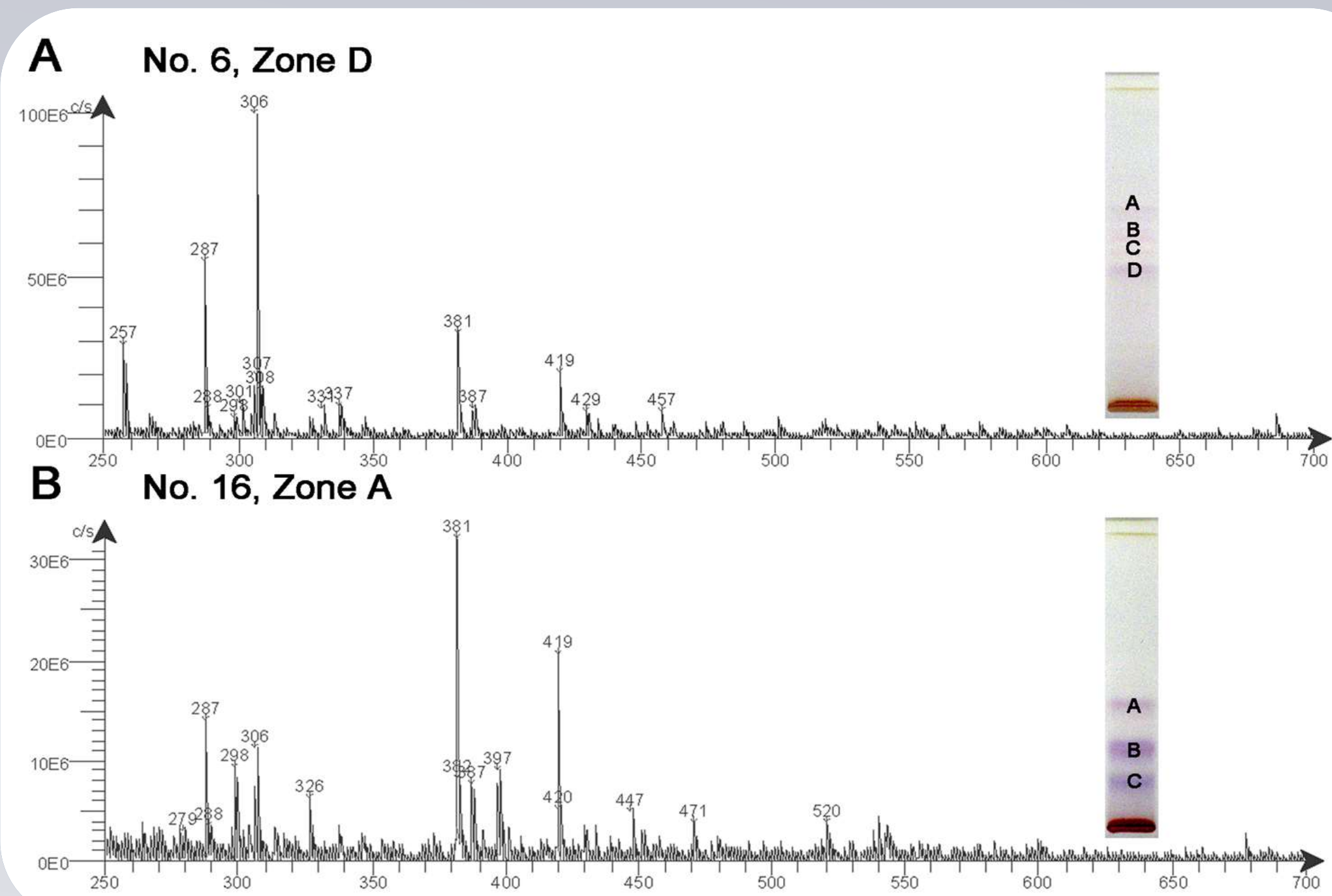


Fig. 2 HPTLC-MS via the elution head-based interface of two anthocyanin zones

## 1. Anthocyanin fingerprints

Milled colored wheat grains were suspended in methanolic hydrochloric acid, 0.5% (0.1 g/1.5 mL), mixed, ultrasonicated (0.5 h), centrifuged and membrane filtered. Anthocyanins were separated on either HPTLC silica gel 60 NH<sub>2</sub> F<sub>254</sub> with ethyl acetate - 2-butanone - water - formic acid 7:3:1.5:1 or RP-18 WF<sub>254</sub>s with water - *n*-propanol - formic acid 9:4:0.6 [1]. The profiles on the amino phases (Fig. 1) clearly differentiated purple- (no. 1-13) versus blue-grained (no. 14-19) varieties as well as combinations of the different genetic backgrounds of blue aleurone and purple pericarp genotypes. [1,2].

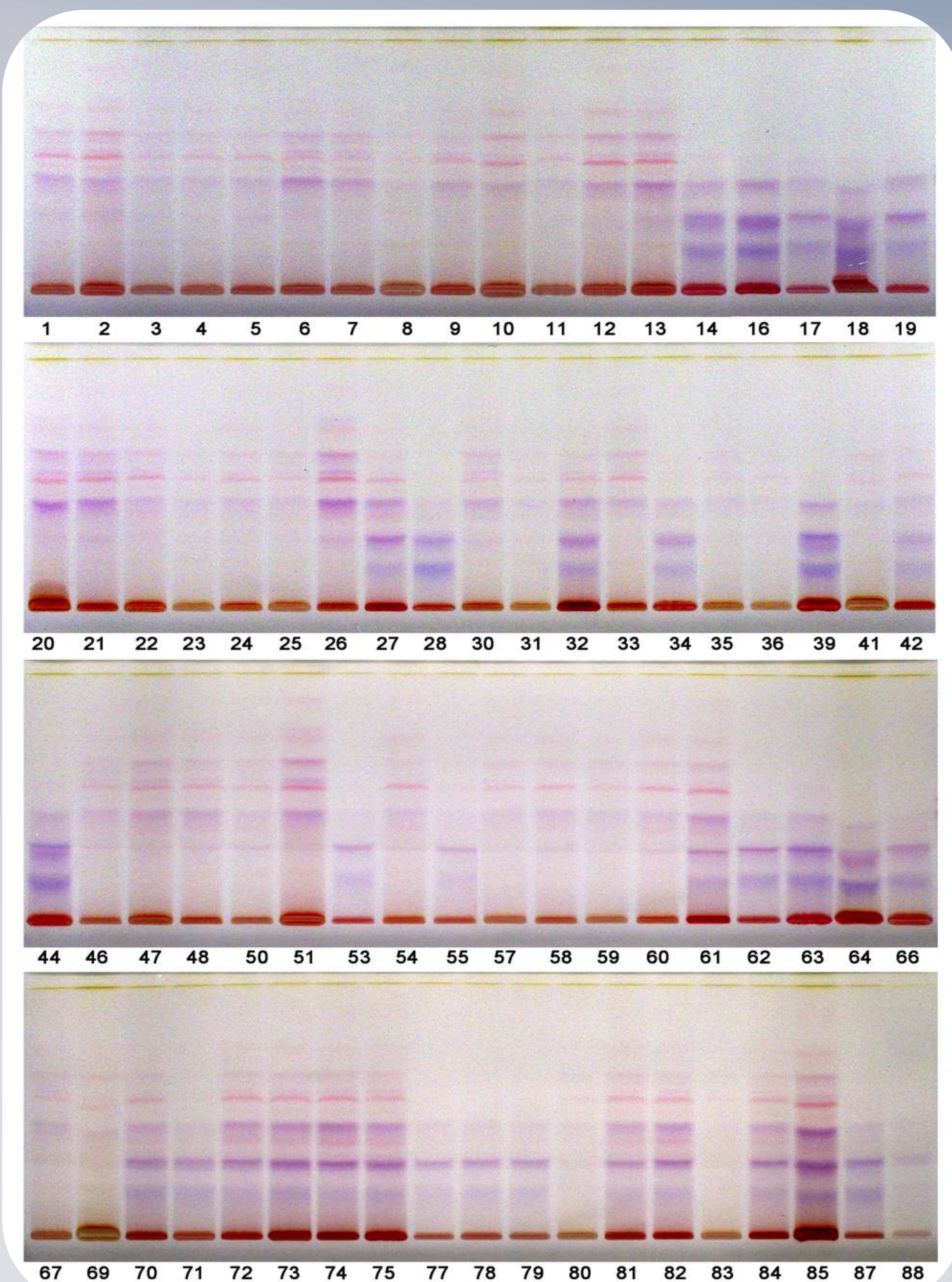


Fig. 1 HPTLC chromatograms of 88 colored wheat extracts showing purple- (no. 1-13) versus blue-grained (no. 14-19) varieties

## 3. Multi-detection → matrix

The chromatograms were detected using various derivatization reagents to obtain further information on matrix compounds (Fig. 3).

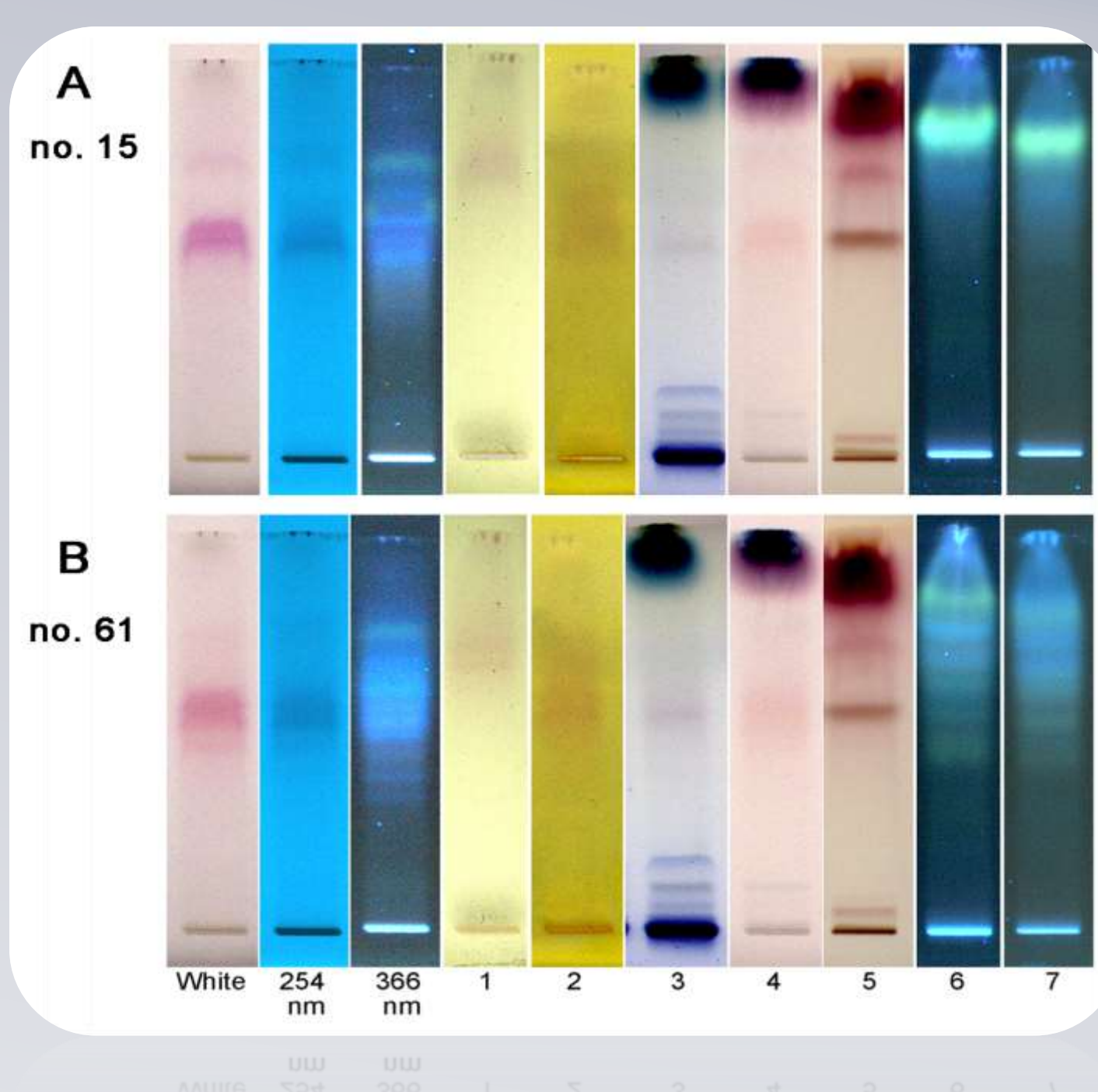


Fig. 3 Multi-detection of samples no. 15 and 61 at white light, UV 254 nm, UV 366 nm and after derivatization with fast blue salt B (1), Dragendorff (2), anisaldehyde sulfuric acid (3), diphenylamine aniline phosphoric acid (4), ninhydrine (5), all at white light, as well as after natural product reagent A (6) and aluminum chloride (7) at UV 366 nm [1]

**Literature** [1] S. Krüger, G.E. Morlock, J. Chromatogr. A, in submission. [2] S.N.S. Jaafar, J. Baron, S. Siebenhandl-Ehn, T. Rosenau, S. Böhmendorfer, H. Grausgruber, Plant Breed. 132 (2013) 546-552

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