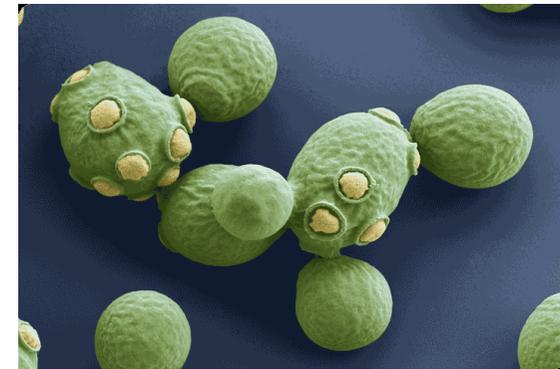


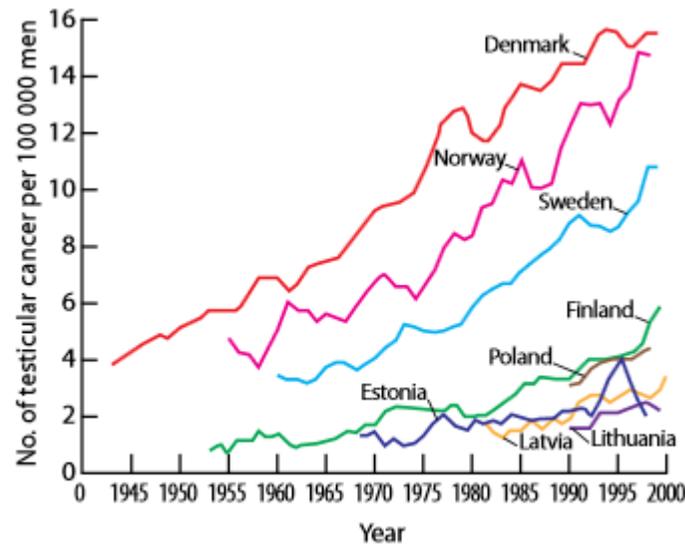
Screening untreated drinking water on estrogenic activity

using the planar-YES, a TLC-EDA tool

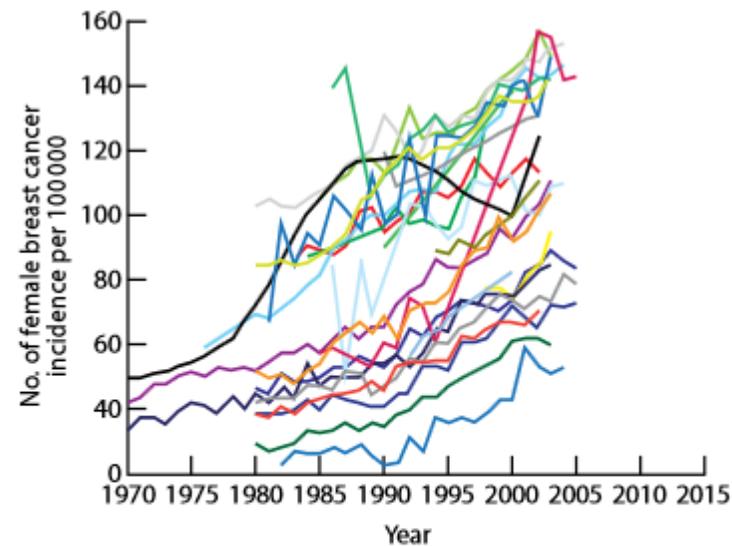
Andreas Schoenborn // July 3, 2014 // HPTLC 2014 Lyon



Testicular & breast cancer increase (WHO)



Richiardi et al., 2004



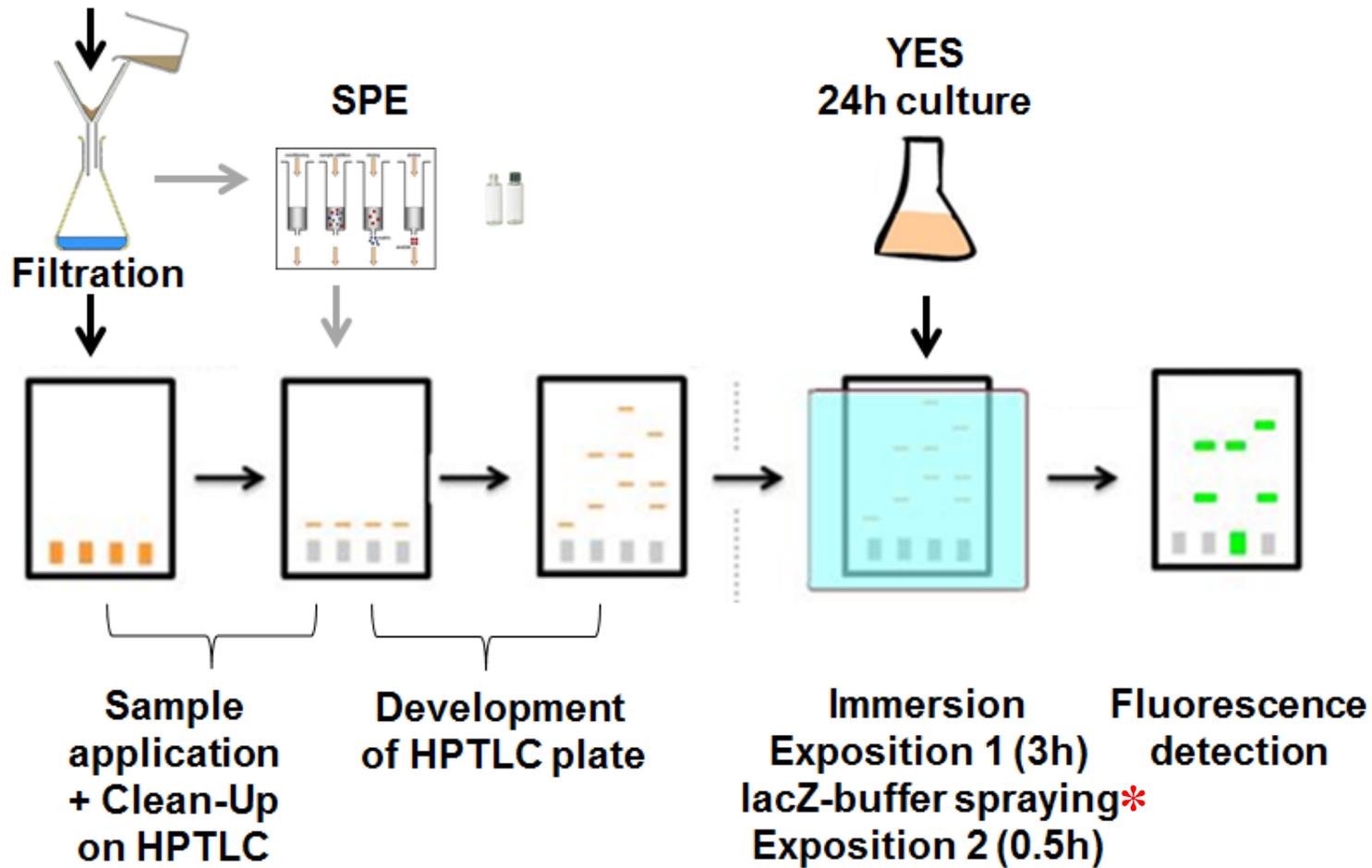
<http://data.euro.who.int/hfad/>

Four study objectives

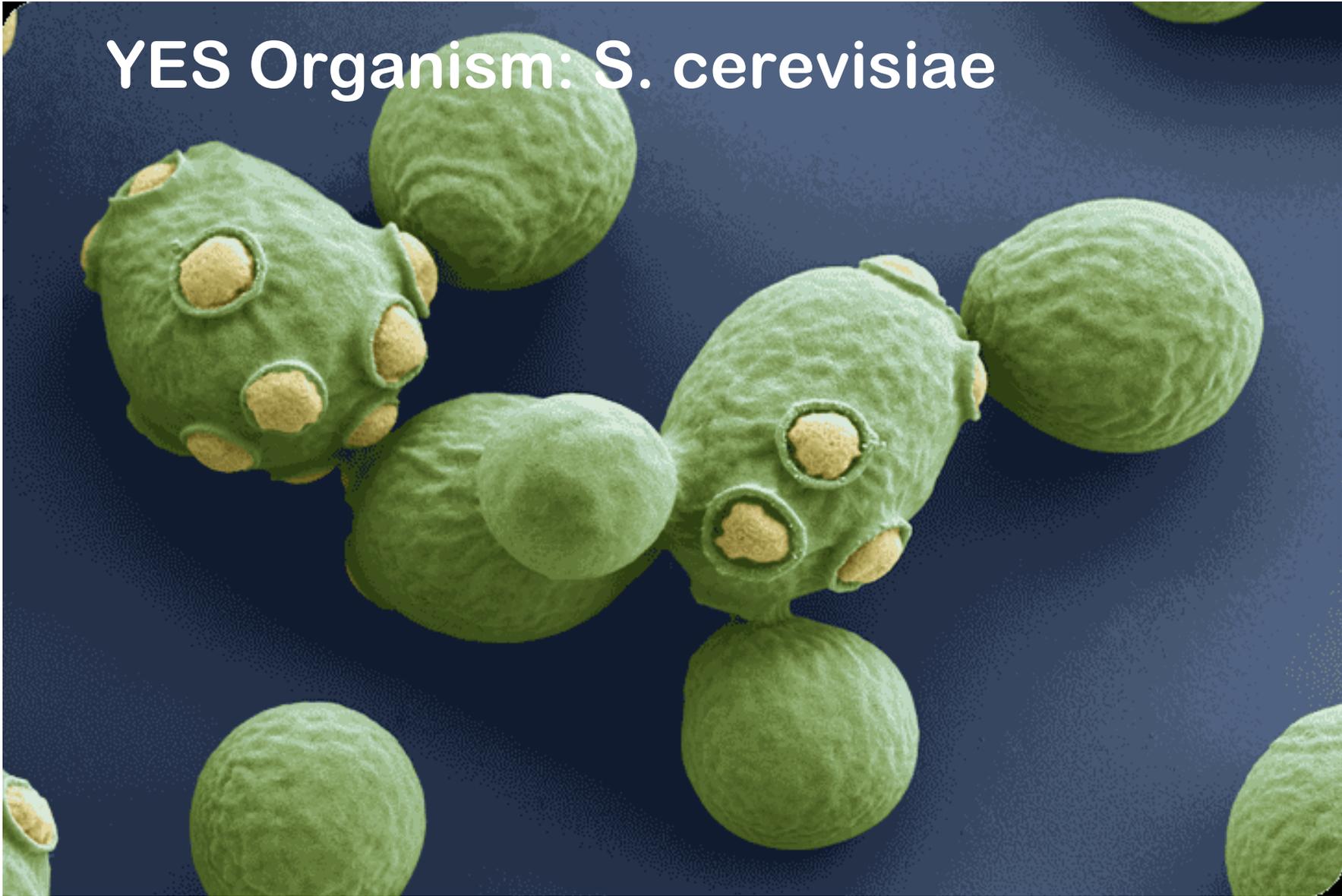
1. Estrogenic activity of selected untreated drinking water samples
2. Indications for the origin of estrogenic activity
3. Added value of undeveloped vs. developed plates
4. Proof-of-concept for direct processing of water samples on HPTLC plates (data not presented)

«planar-YES» - Workflow

Raw water sample



YES Organism: *S. cerevisiae*



http://www.microbiologyonline.org.uk/themed/sgm/img/slideshows/3.1.4_fungi_2.png

Sample provenience

1. Evian-Water (negative control)
 2. River bank infiltrate, Zurich
 3. Sewage treatment plant outlet, Zurich
 4. Limmat water, Zurich
 5. Rhine water, Basel
- *Water sample divided in 2 parts*
 - *Part 2 spiked with 7 estrogens and xenoestrogens to 10 ng/l each.*
 - *Extraction 3 times with SPE on consecutive days => 6 vials per sample*

Standard mixture for spikes

Added to sample before extraktion;
10 ng/L of each substance:

- Bisphenol A (REP = 0.00005)
- E1 Estrone (REP = 0.47)
- E2 17 β -Estradiol (REP = 1)
- Mestranol (REP = 0.073)
- Norethisterone (REP = 0.04)
- EE2, 17 α -Ethinylestradiol (REP = 0.88)
- Estriol (REP = 0.0076)

Source: Michael Koss, Wasserversorgung Zürich

ZHAW // IUNR Institute of Natural Resource Sciences

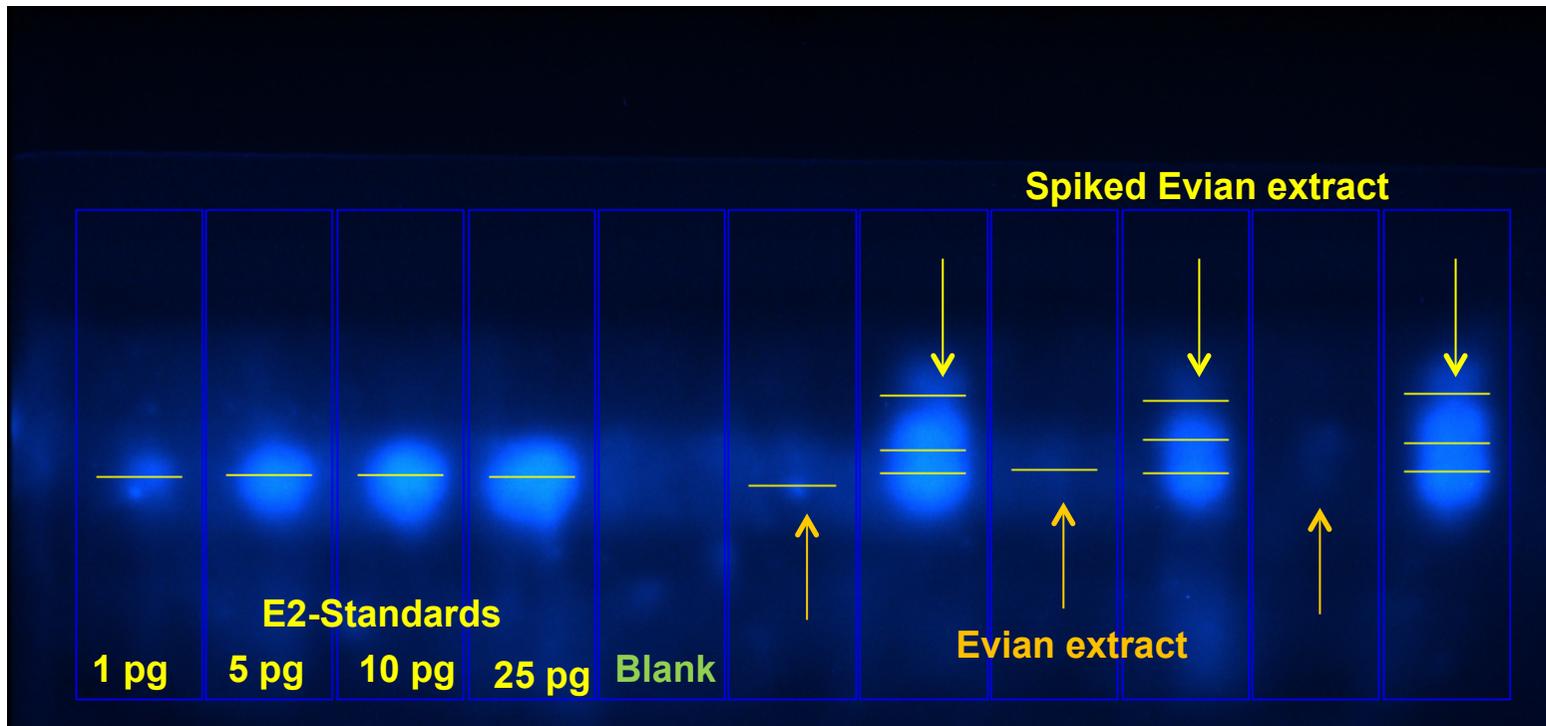
**Expected EEQ
in sample:
app. 25 ng/l**

SPE procedure (performed by WVZ)

- pH-Wert of sample adjusted to 7 (with conc. HCl)
- Pre-filtration with GF
- (50% of samples: addition of standard mixture)
- SPE with Oasis HLB 6cc on Caliper Autotrace
- Drying of solid phase with N₂, 60 min
- Elution with 4 mL acetone, 1 mL methanol
- Drying
- Take up in 1 mL ethanol

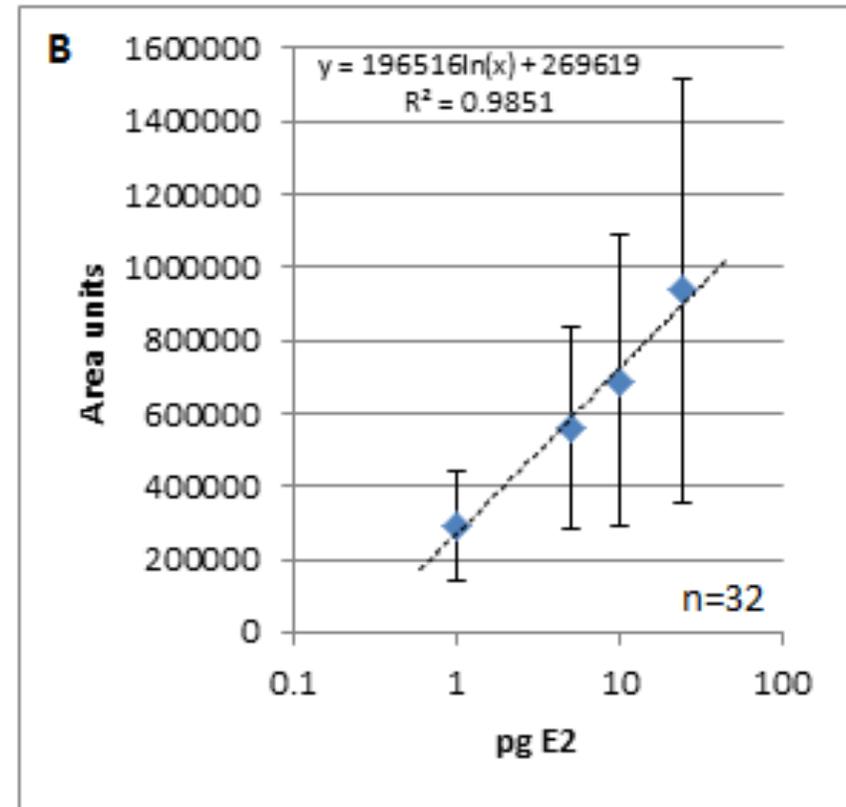
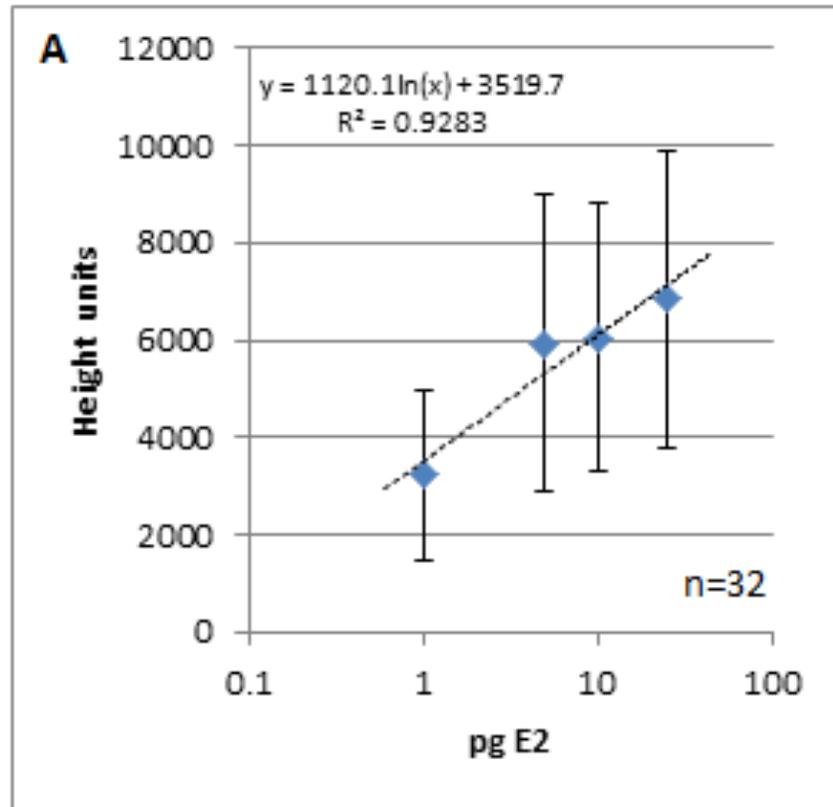
Concentration factor
931 ± 5%

Evian water (negative control)

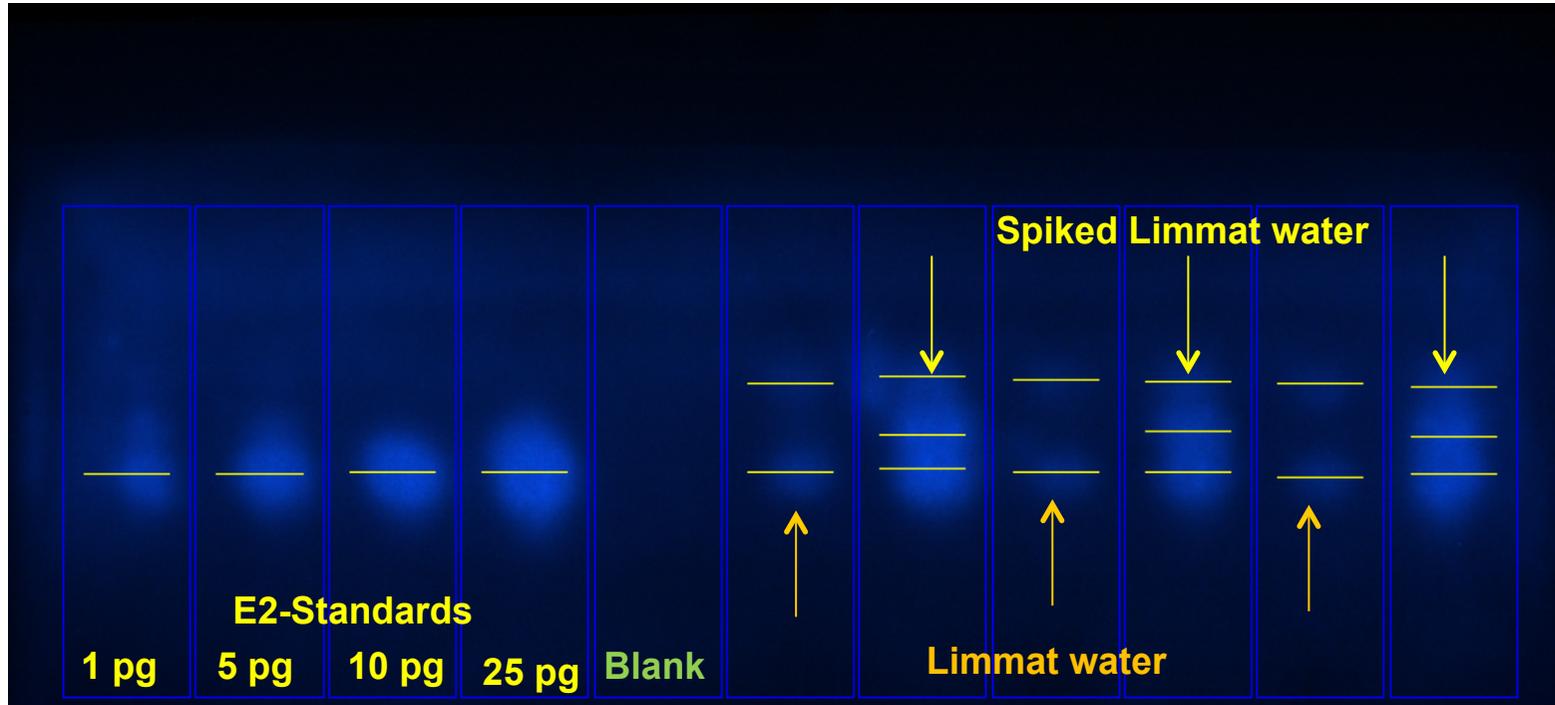


Spot-Nummer	1	2	3	4	5	6	7	8	9	10	11
Aufgetragene Proben	1 pg E2 (0.2 µg/l)	5 pg E2 (1.0 µg/l)	10 pg E2 (2.0 µg/l)	25 pg E2 (5.0 µg/l)	P5 Lösemittel- kontrolle WWZ: P5	Kontrolle: Evian WWZ: K/R1 1/1 nativ (05.08.13)	Kontrolle: Evian WWZ: K/R1 1/4 aufgestockt (05.08.13)	Kontrolle: Evian WWZ: K/R2 2/1 nativ (06.08.13)	Kontrolle: Evian WWZ: K/R2 2/4 aufgestockt (06.08.13)	Kontrolle: Evian WWZ: K/R3 3/1 nativ (08.08.13)	Kontrolle: Evian WWZ: K/R3 3/4 aufgestockt (08.08.13)
Aufgetragenes Probenvolumen	5 µl	5 µl	5 µl	5 µl	1 µl	1 µl	1 µl	1 µl	1 µl	1 µl	1 µl
Hormonlösung	L6-E2	L4-E2	L5-E2	L7-E2	P5 (16.09.13)	K/R1 1/1 (05.08.13)	K/R1 1/4 (05.08.13)	K/R2 2/1 (06.08.13)	K/R2 2/4 (06.08.13)	K/R3 3/1 (08.08.13)	K/R3 3/4 (08.08.13)

Calibration with E2 standards

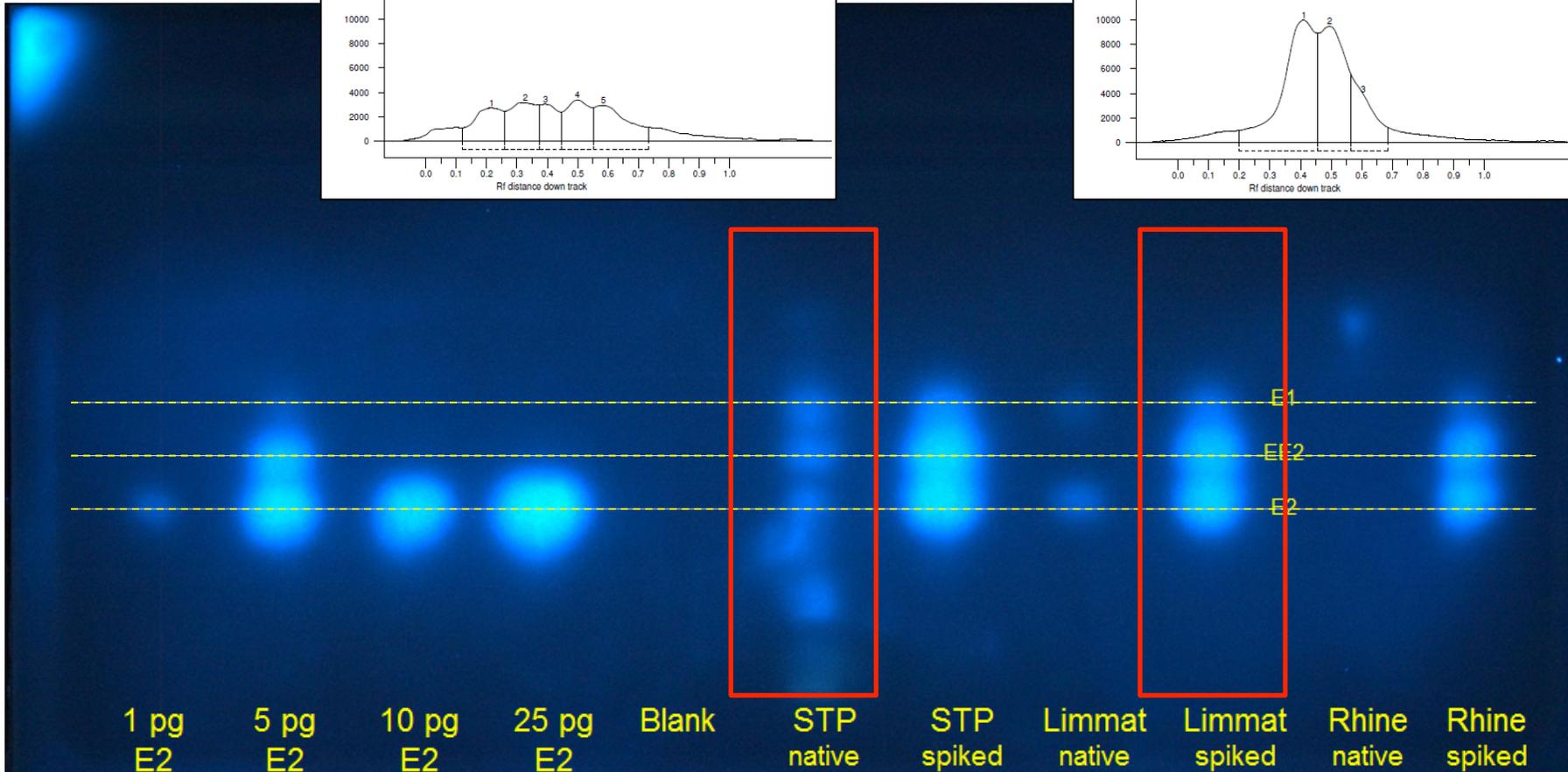
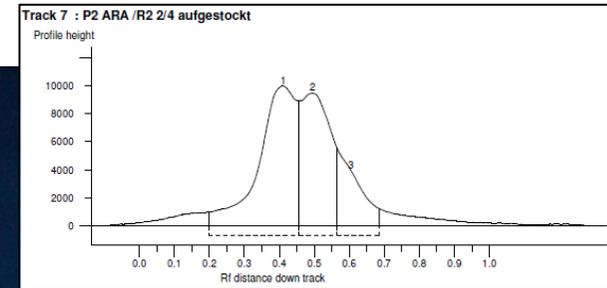
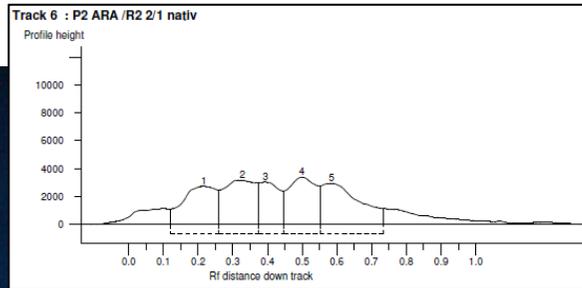


Limmat water (objective 1)

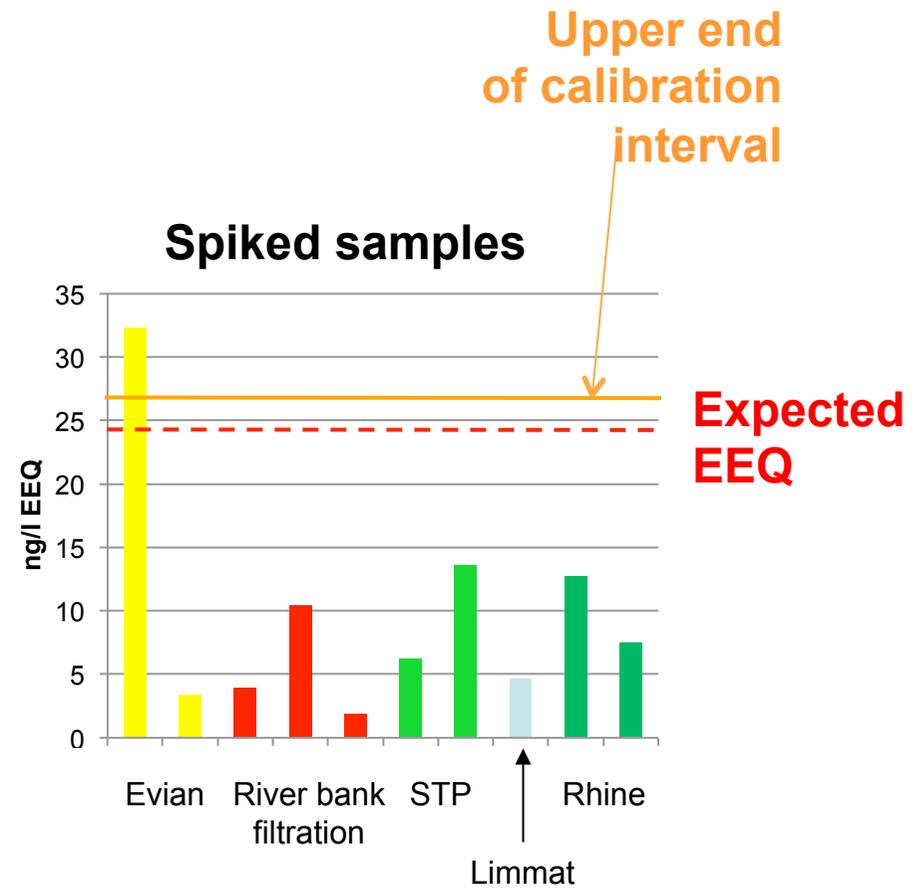
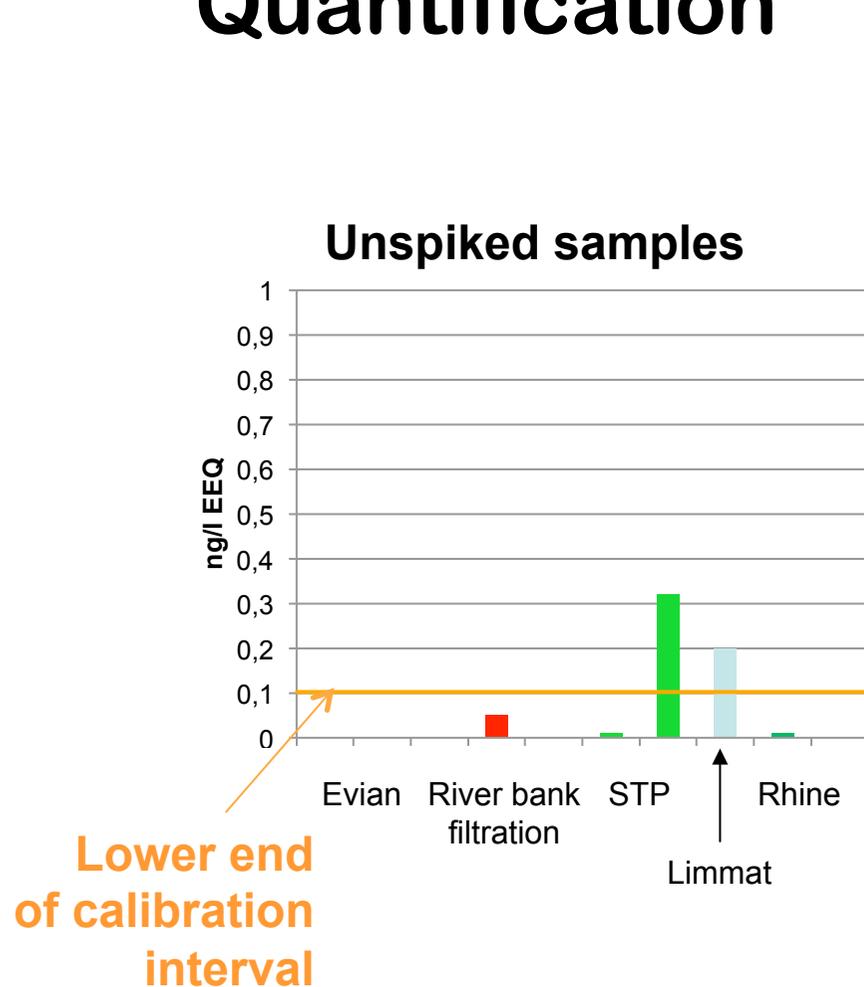


Spot-Nummer	1	2	3	4	5	6	7	8	9	10	11
Aufgetragene Proben	1 pg E2 (0.2 µg/l)	5 pg E2 (1.0 µg/l)	10 pg E2 (2.0 µg/l)	25 pg E2 (5.0 µg/l)	Lösemittel- kontrolle ZHAW	P3: LIM WVZ: P3/R1 1/1 nativ (21.08.13)	P3: LIM WVZ: P3/R1 1/4 aufgestockt (21.08.13)	P3: LIM WVZ: P3/R2 2/1 nativ (21.08.13)	P3: LIM WVZ: P3/R2 2/4 aufgestockt (21.08.13)	P3: LIM WVZ: P3/R3 3/1 nativ (23.08.13)	P3: LIM WVZ: P3/R3 3/4 aufgestockt (23.08.13)
Aufgetragenes Probenvolumen	5 µl	5 µl	5 µl	5 µl	1 µl	10 µl	1 µl	10 µl	1 µl	10 µl	1 µl
Hormonlösung	L6-E2	L4-E2	L5-E2	L7-E2	EtOH ZHAW	P3/R1 1/1 (21.08.13)	P3/R1 1/4 (21.08.13)	P3/R2 2/1 (21.08.13)	P3/R2 2/4 (21.08.13)	P3/R3 3/1 (23.08.13)	P3/R3 3/4 (23.08.13)

Origin of estrogenic activity (objective 2)

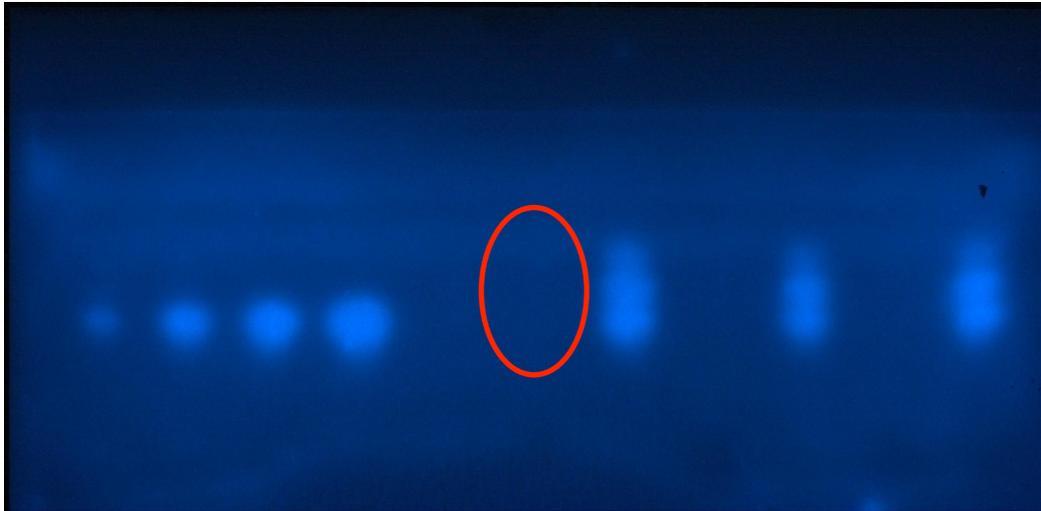


Quantification



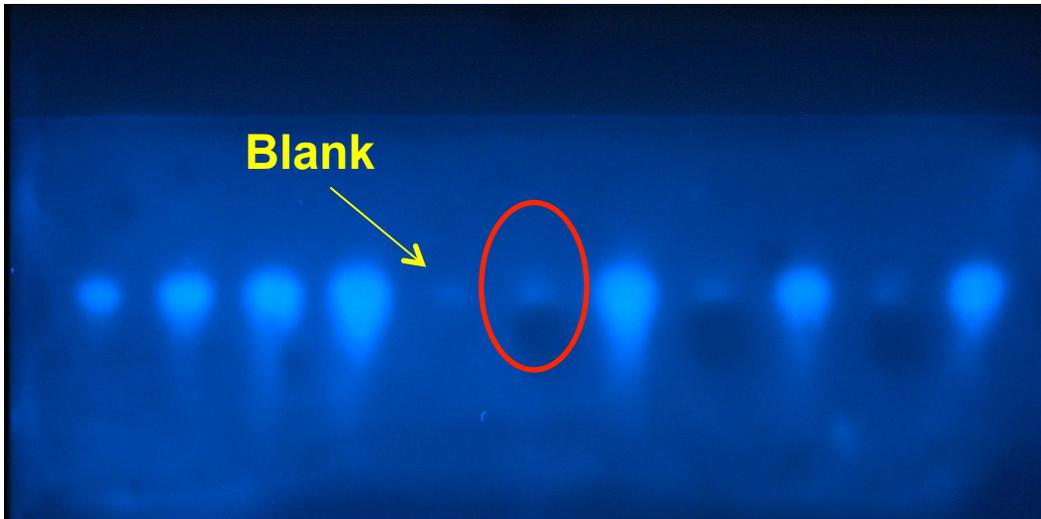
Comparison developed / undeveloped

(objective 3)



Rhine water

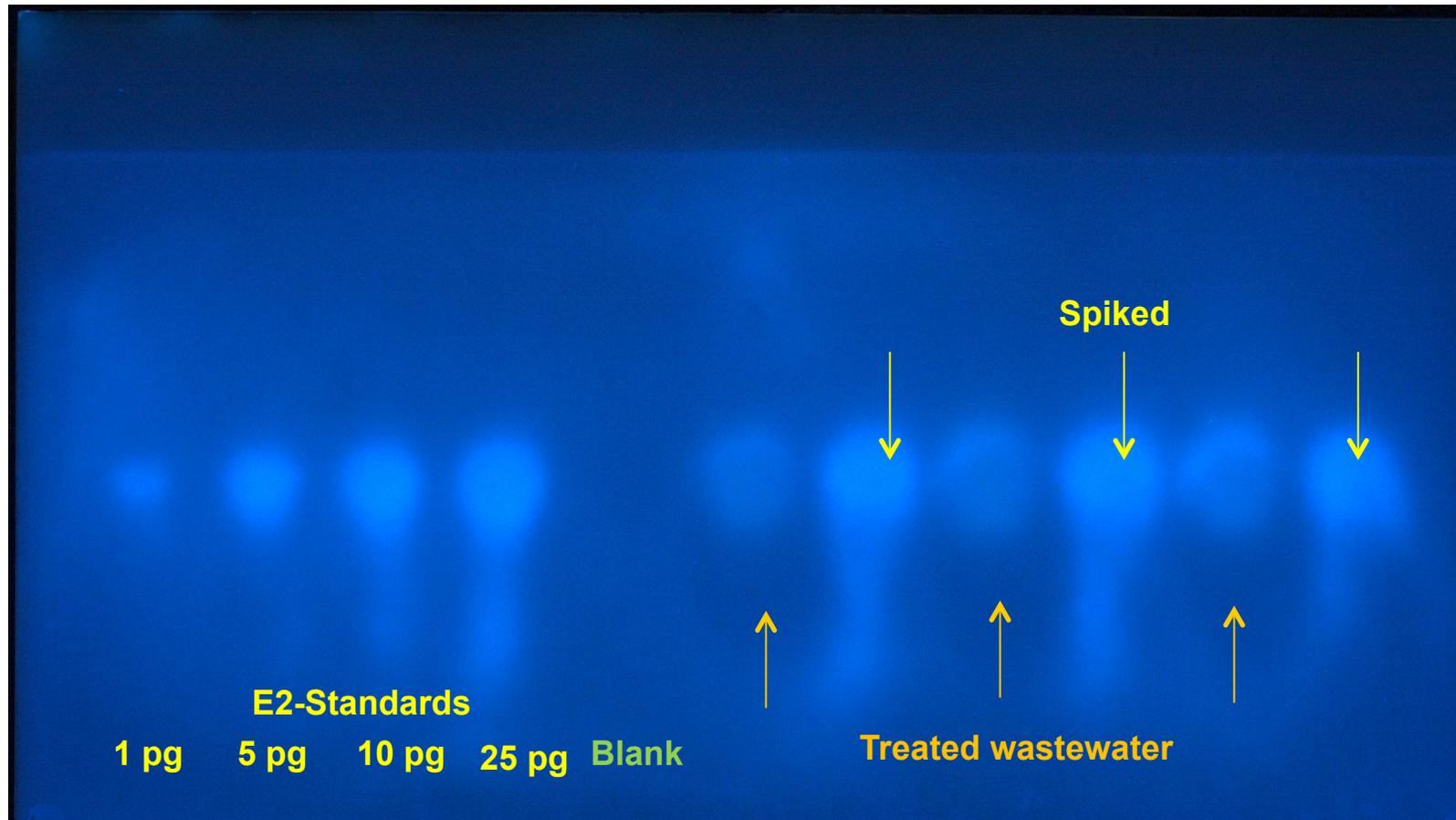
**Developed:
EEQ not detectable**



**Undeveloped:
EEQ detectable**

**Total-EEQ:
<< 1 pg / 10µl extract?**

Added value 2 (Treated wastewater)



Dimmed center of spot indicates inhibition!!!

Conclusion

- Net time demand: 1 day from sampling to result
- Robust, reproducible, 6-9 parallel samples
- LOQ < 1 pg/band EEQ (0.1 pg/band within reach)
- Potential to detect 1 ng/l EEQ in native water samples

Outlook

- Further refinement of procedure needed!
- Link to analytical chemistry difficult (low masses)
- Application to other matrices
(cosmetics, exudates from plastics...)
- ISO certification of planar-YES (?)

Thank you!

Zürich University
of Applied Sciences



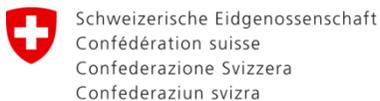
- ZHAW:

- Andrea Grimmer
- Dr. Evelyn Wolfram



- Wasserversorgung Zürich:

- Dr. Michael Kost
- Dr. Oliver Köster



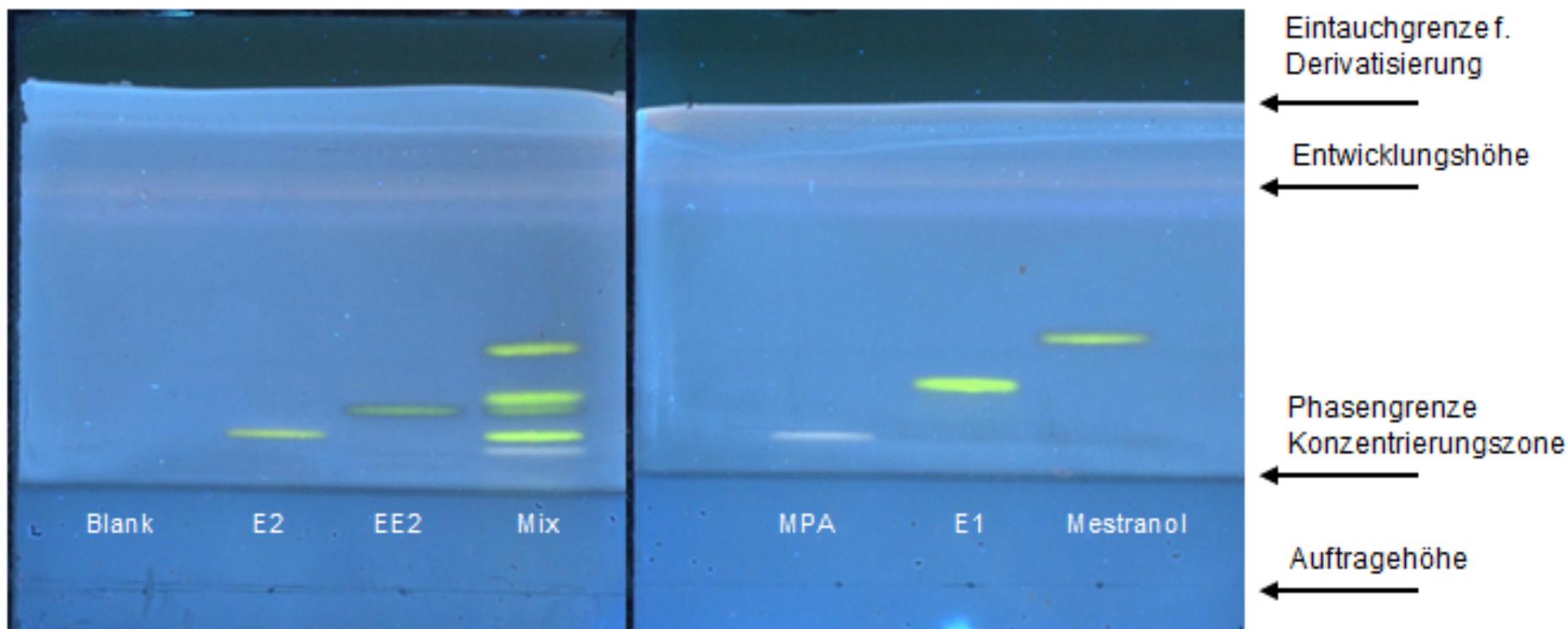
- Bundesamt für Gesundheit:

- Dr. Beat Brüscheiler

SPE vs. Clean-Up on HPTLC (objective 4)

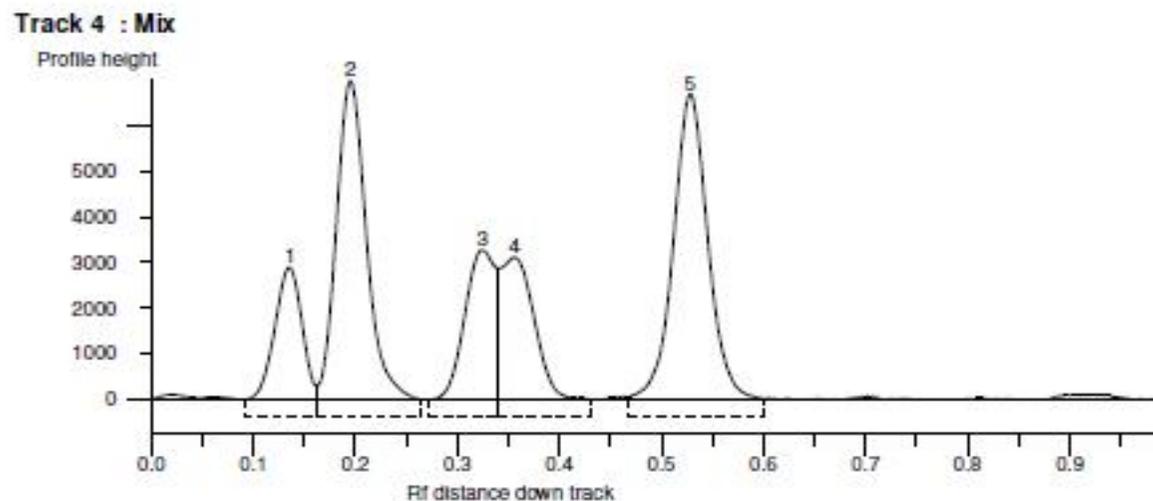
Method	Samp-ling	Extraction	Appli-cation	Drying	Focus-sing	Drying	Plate development
State of art	X	With SPE	X (SPE extract)	X	---	---	X
A. Clean-Up with focussing	X	---	X (water sample)	X	X (with CH ₃ OH)	X	X
B. Clean-Up on HPTLC plate with concentration zone	X	---	X (water sample)	X	---	---	X
Time demand	---	ca. 1 d / 1-6 samples	ca. 1,5 h / plate		ca. 0,5 h / plate		ca. 1,5 h / plate

Focussing on HPTLC-K plates



275 ng/Spot, dissolved in 5 μ l ethanol (manual application). Height: 1 cm (in concentration zone). Development to 7,5 cm with ADC2 (Camag). Mobile phase: n-Hexan 4 / Ethylacetate 1,5. Derivatization with mit sulfuric acid reactant. Light: 366 nm. Exposure time 3665.97ms, Gain 1.

Separation on HPTLC-K plates



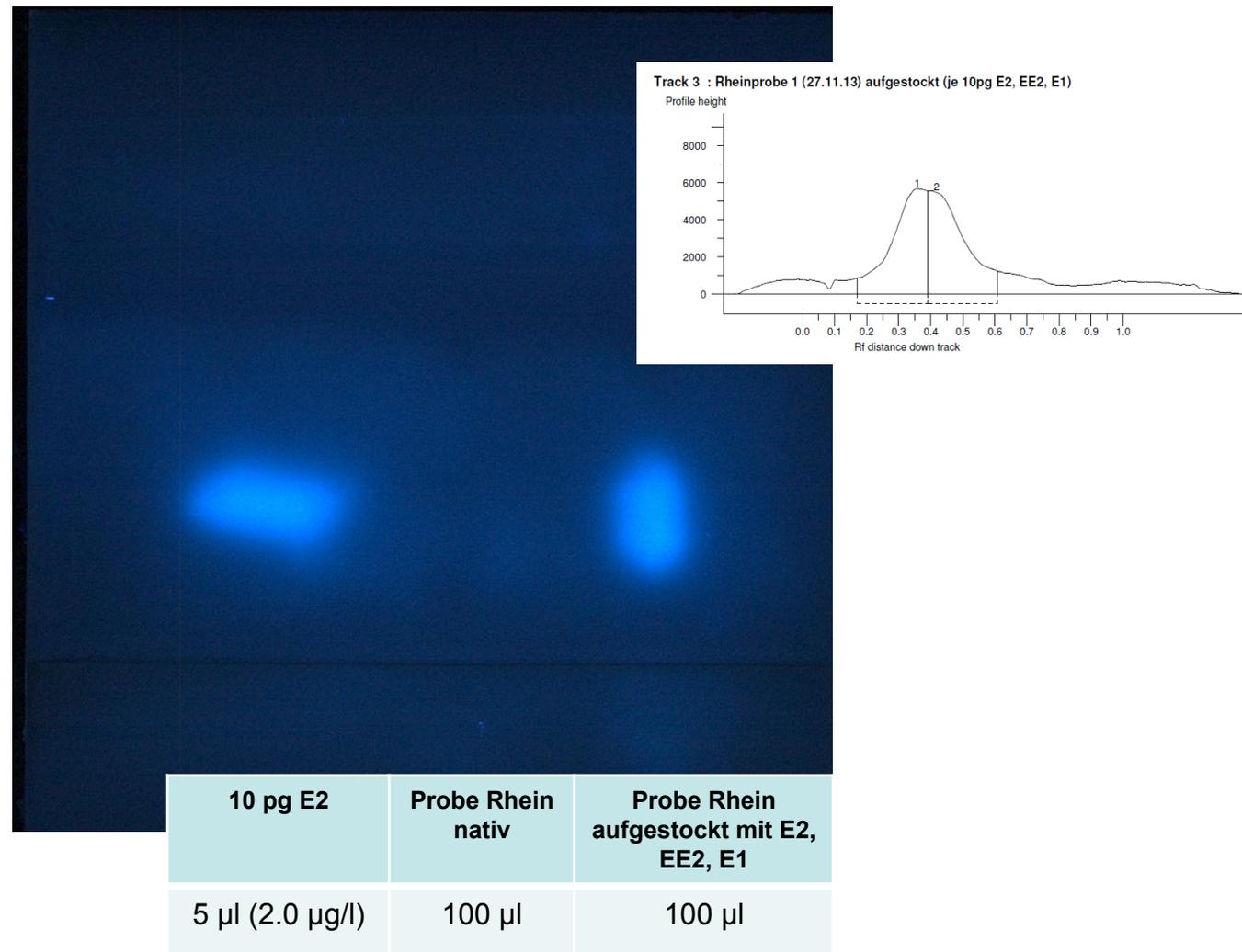
Track 4

Peak #	Start Rf	Start H	Max. Rf	Max. H	Max. [%]	End Rf	End H	area A	area [%]	Subst Name
1	0.091	0.0	0.136	2893.6	12.60	0.163	299.7	39316.6	10.89	
2	0.163	299.7	0.195	6975.1	30.38	0.263	7.7	104191.8	28.87	
3	0.272	0.0	0.324	3275.3	14.27	0.340	2864.2	49708.4	13.77	
4	0.340	2864.2	0.356	3117.2	13.58	0.431	9.2	52129.0	14.44	
5	0.467	75.8	0.528	6697.9	29.17	0.601	25.9	115575.7	32.02	

Total Height 22959.1 Total Area : 360922

275 ng/Spot, dissolved in 5 µl ethanol (manual application). Height: 1 cm (in concentration zone). Development to 7,5 cm with ADC2 (Camag). Mobile phase: n-Hexan 4 / Ethylacetate 1,5. Derivatization with mit sulfuric acid reactant. Light: 366 nm. Exposure time 3665.97ms, Gain 1.

Proof-of-concept: Clean-Up on HPTLC-K



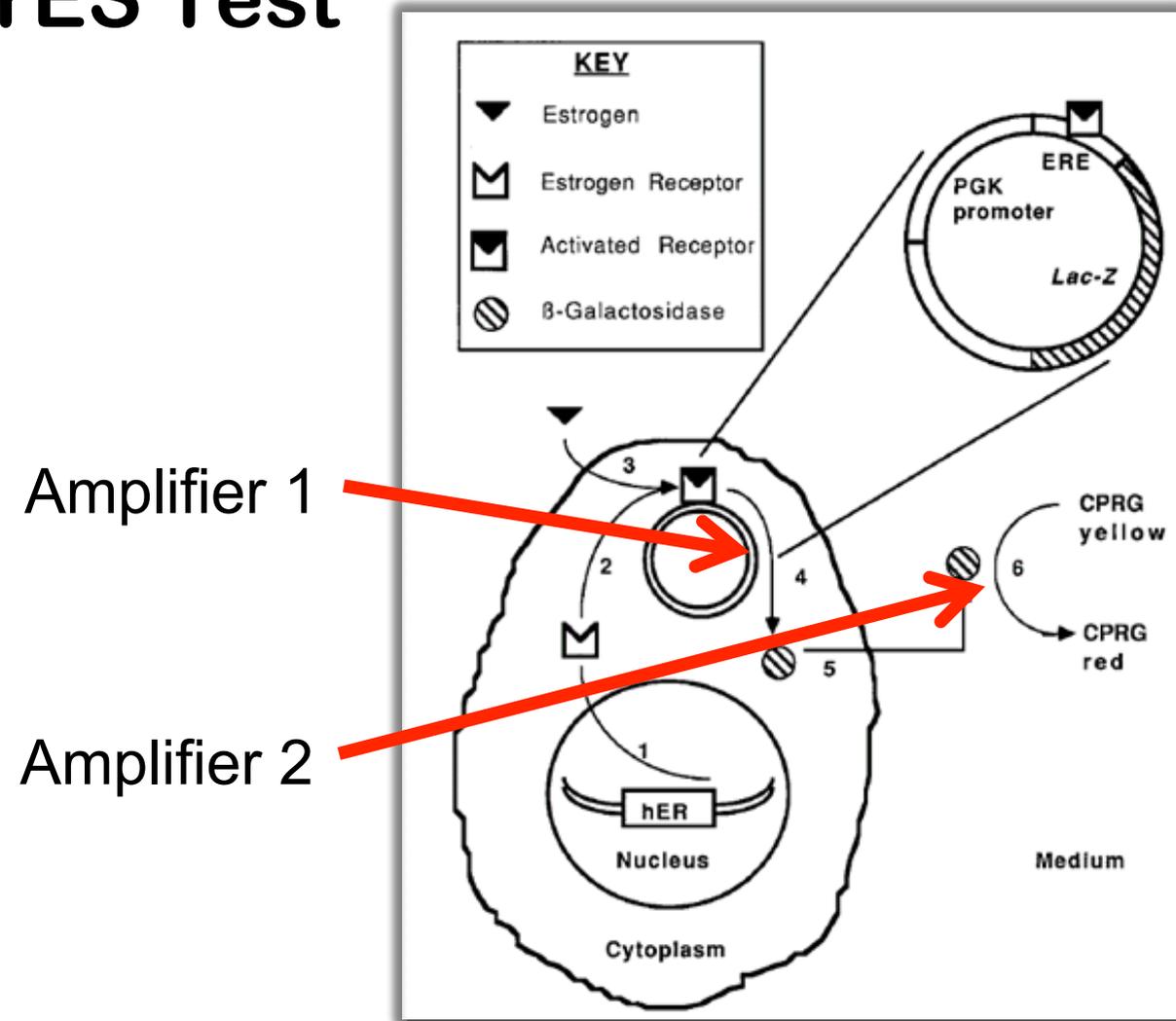
Potential endocrine disruptors (WHO)

- Persistente und bioakkumulierende Halogenide: Langlebige organische Schadstoffe (z. B. Polychlorierte Dibenzodioxine und Dibenzofurane, Polychlorierte Biphenyle, Hexachlorbenzol, Perfluorooctansulfonat, Polybromierte Diphenylether, Polybromierte Biphenyle, Chlordan, Mirex, Toxaphen, Dichlordiphenyltrichlorethan, Lindan, Endosulfan); andere persistente und bioakkumulative Stoffe (z. B. Hexabromcyclododecan, Chlorparaffine, Perfluorierte Carbonsäuren, Octachlorostyrene, PCB Methyl Sulfone)
- weniger persistente und weniger bioakkumulative Stoffe: Weichmacher und Additive (z. B. Phthalsäureester, Triphenylphosphat, Diethylhexyladipat, Butylbenzole, Triclocarban, Butylhydroxyanisol); Polycyclische aromatische Kohlenwasserstoffe (z. B. Benzopyren, Benz(a)anthracen, Pyren, Anthracen); Halogenphenole (z. B. 2,4-Dichlorphenol, Pentachlorophenol, Hydroxy-PCB, Hydroxy-PBDE, Tetrabrombisphenol A, 2,4,6-Tribromphenol, Triclosan); nicht-Halogenphenole (z. B. Bisphenol A, Bisphenol F, Bisphenol S, Nonylphenol, Octylphenol, Resorcinol)
- Pestizide, Arzneimittel und Kosmetika: gegenwärtig genutzte Pflanzenschutzmittel (z. B. 2,4-Dichlorphenoxyessigsäure, Atrazin, Carbaryl, Malathion, Mancozeb, Vinclozolin, Prochloraz, Propylidon, Chlorpyrifos, Fenitrothion, Linuron); Arzneimittel, Wachstumshormone und Kosmetika (z. B. Diethylstilbestrol, Ethinylestradiol, Tamoxifen, Levonorgestrel, Fluoxetin, Flutamid, 4-Methylbenzyliden-campher, Octylmethoxycinnamat, Parabene, Octamethylcyclotetrasiloxan, Decamethylcyclopentasiloxan, Dodecamethylcyclohexasiloxan, Galaxolid, 3-Benzyliden-Campher)
- Andere Chemikalien: Metalle und Organometalle (z. B. Arsen, Cadmium, Blei, Quecksilber, Methylquecksilber, Tributylzinnhydrid, Triphenylzinn); natürliche Hormone (z. B. 17-beta-Estradiol, Estron, Testosteron); Phytoöstrogene (z. B. Isoflavone, Coumestan, Mykotoxine, Prenylflavonoide)

Zahlen zu den Rohwasser-Messungen

- 6 Messdurchgänge (davon 2 x als Vorversuch)
- 20 auswertbare HPTLC Platten (10 x 20 cm, 10 x 10 cm)
- 206 aufgebrauchte Tracks, davon:
 - 112 Extrakte (mit und ohne DC)
 - 74 E2-Standards (mit und ohne DC)
 - 20 Lösungsmittelblanks (mit und ohne DC)
- 271 auswertbare Datensätze
 - 99 gut auswertbare Peaks (37%)
 - 101 mit Vorbehalt auswertbare Peaks (37%)
 - 34 Tracks ohne detektierbare Peaks (13%)
 - 37 nicht auswertbare Tracks (14%)

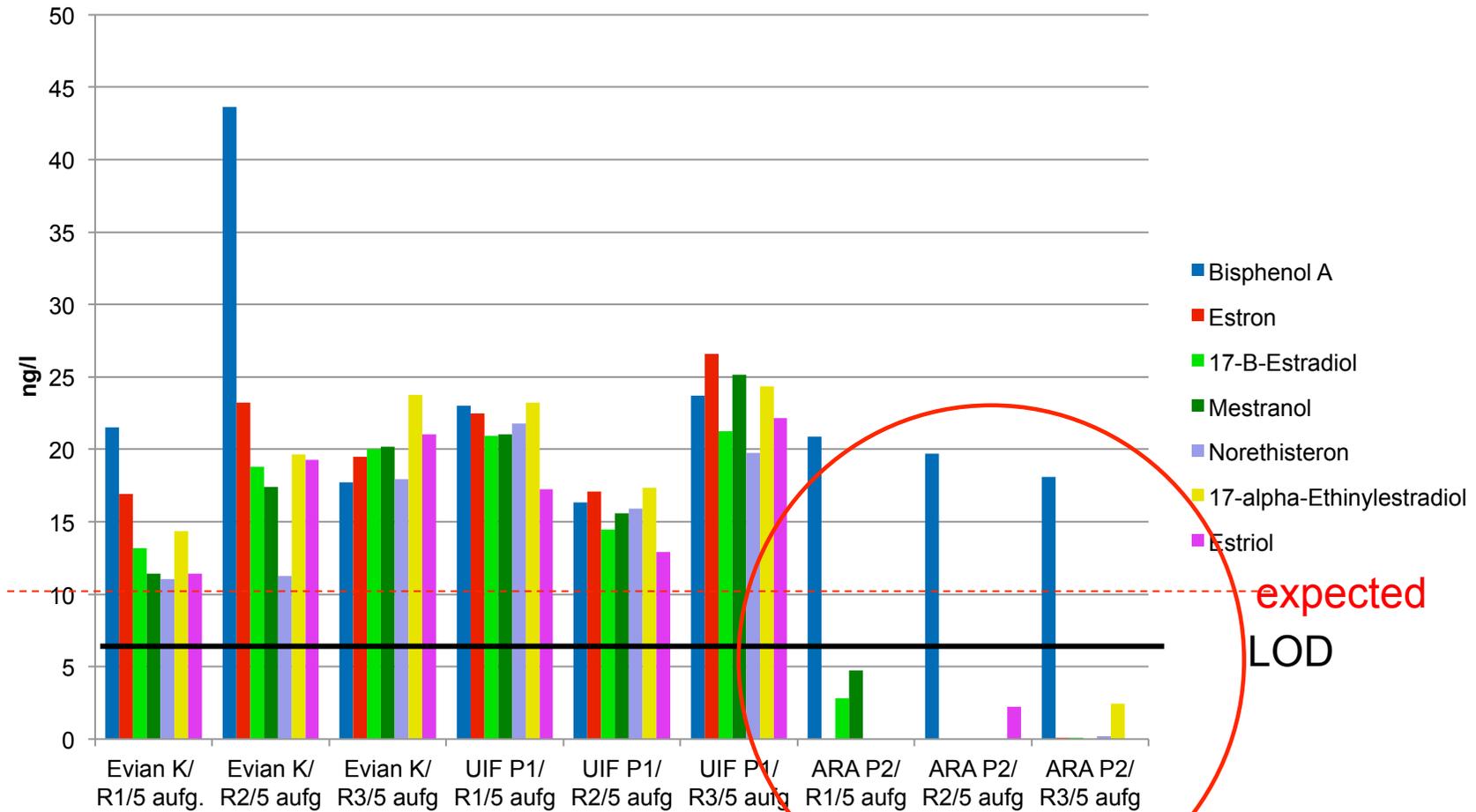
Prinzip YES Test



Routledge, E. J. & Sumpter, J. P., 1996, *Env. Toxic. Chem.*, 15(3), 241 - 248

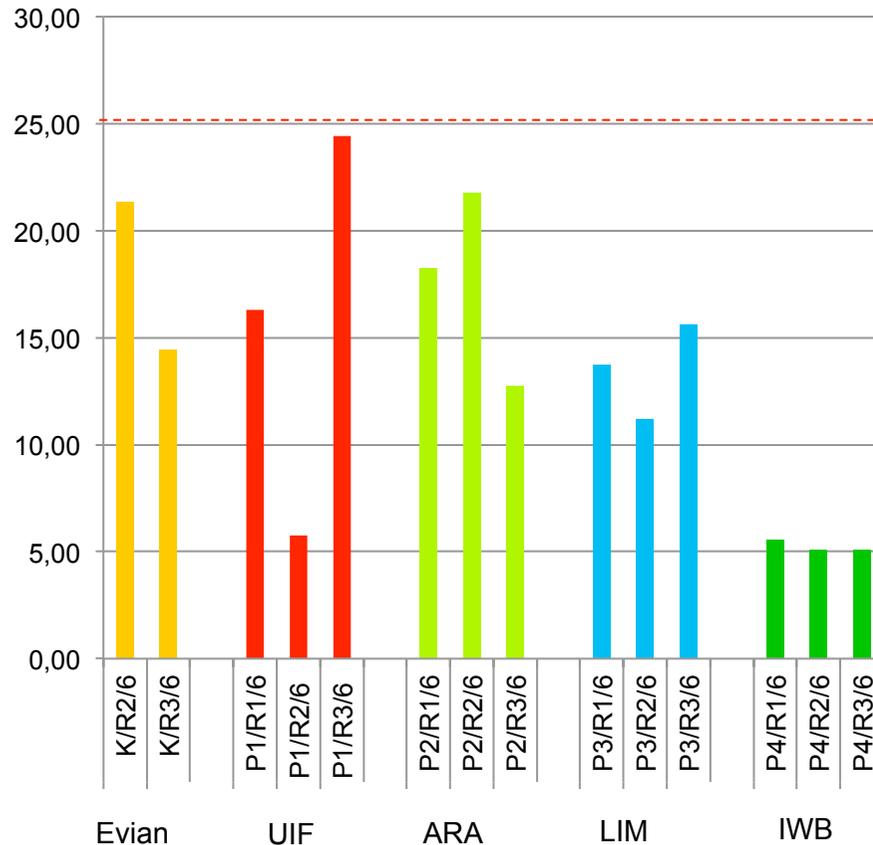


Target analytics of spiked samples

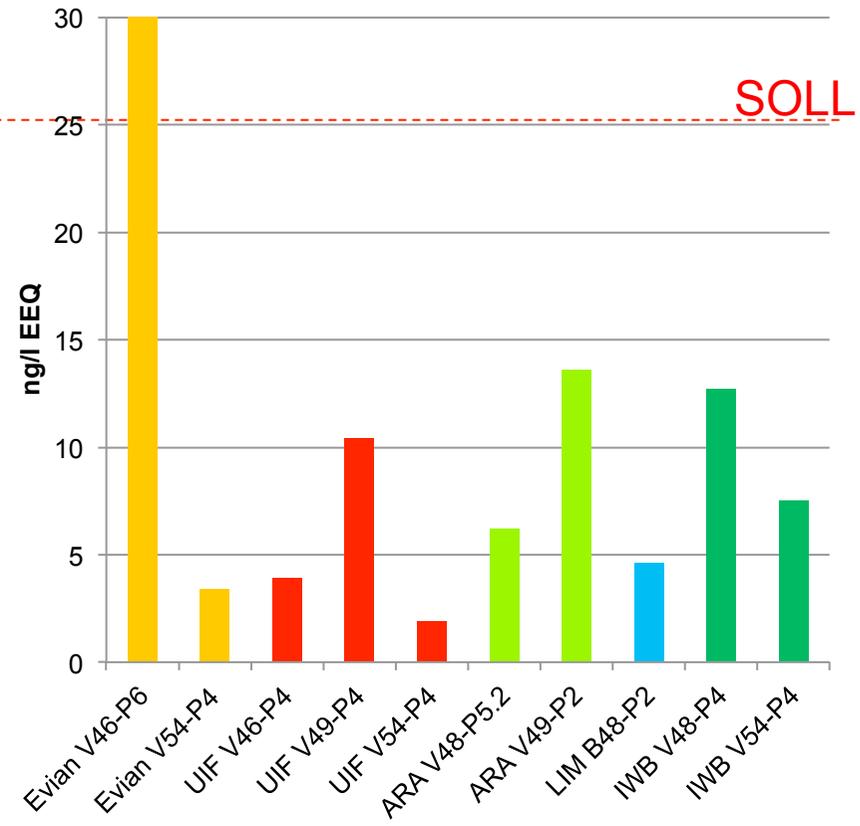


Spiked samples EEQ (ng/l)

LYES (Xenoscreen)

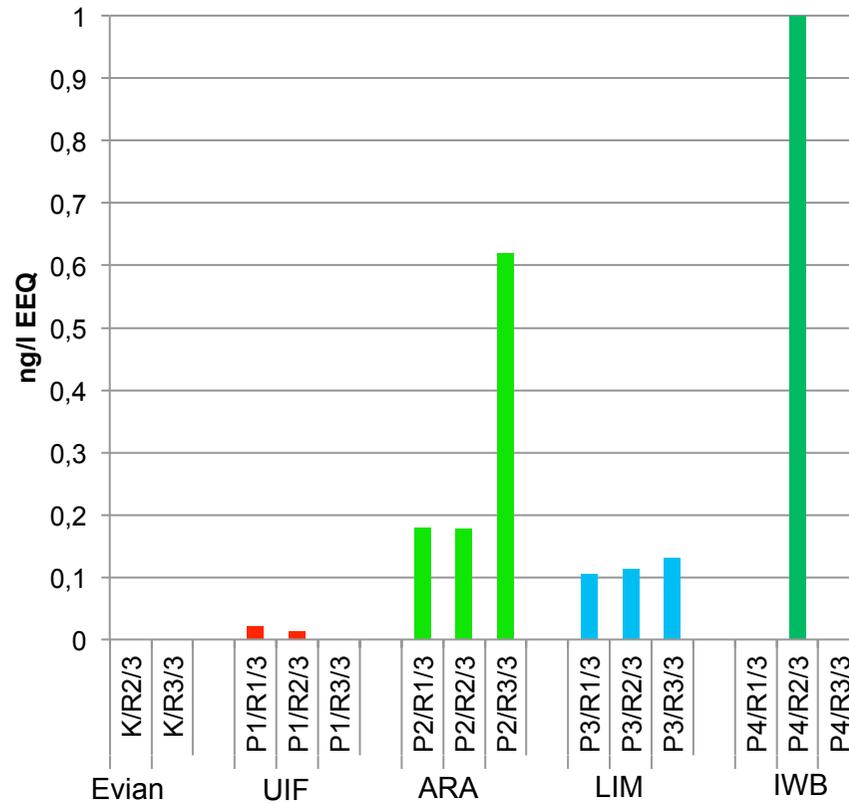


planar-YES (ZHAW)

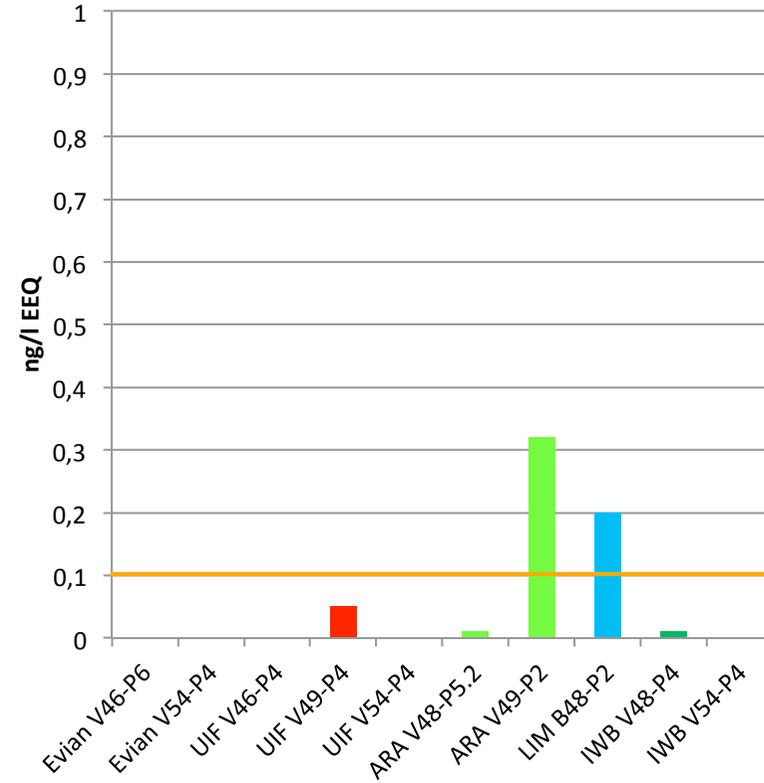


Native Proben EEQ (ng/l)

LYES (Xenoscreen)



planar-YES (ZHAW)



Why estrogens?

