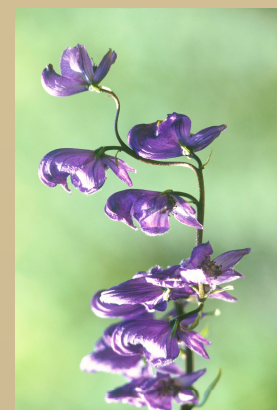




Detection of the decomposition of aconitine in Aconitum napellus mother tincture V.2a and identification of the main cleavage products using HPTLC-MS



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Aconitine, the main alkaloid of Aconitum napellus is one of the most toxic phytochemicals. The toxicity decreases from aconitine to its degradation products induced by hydrolytic cleavage of acetate and / or benzoate groups. So the decomposition of aconitine in aged tinctures is of great interest when used for pharmaceutical purposes.

Sample preparation for HPTLC: 2 ml of the tincture were mixed with 2 ml of water *R* and 1 ml of diluted ammonia solution *R* 3. The mixture has been put onto a kieselguhr cartridge and eluted twice with 8 ml of ether *R*. The extract was evaporated to dryness under reduced pressure. The residue was dissolved in 0,5 ml of methanol *R*.

HPTLC method: The sample solutions were applied on a HPTLC silica gel 60 (20 x 10 cm) glass plate (Merck, Darmstadt, Germany) by ATS4 (CAMAG, Berlin, Germany). Using a mobile phase of cyclohexane / ethylacetate / diethylamine (70:20:10 v/v/v) the plate was developed twice over a migration distance of 60 mm. For detection the plate was sprayed with a mixture of a solution of tartaric acid in water and the solution of iodine and iron-III-chloride in acetone. Evaluation in vis.

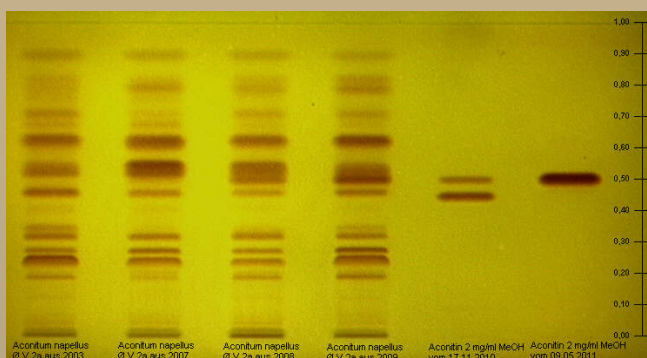


Figure 1: HPTLC of tinctures from 2003, 2007, 2008 and 2009 and reference solutions (new and aged)

Sample preparation for HPLC:

The visible zones (Figure 2) were marked on an underivatized plate at the matching R_f value and extracted with methanol *R* (2 min, flow rate 0,3 mL · min⁻¹) using a TLC - MS - interface (CAMAG, Berlin, Germany). All sample solutions of the zones were separately collected in glass vials.

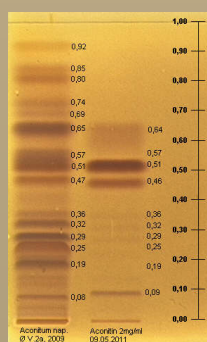


Figure 2: HPTLC of tincture and reference solution showing the extracted zones

Instrumentation for HPLC:

HPLC was performed with a Thermo Fisher Scientific Accela (Thermo Fisher Scientific, Dreieich, Germany) instrument with diode-array detector. The equipment was controlled by means of Xcalibur, Version 2.07. Compounds were separated on a 100 x 200 mm i.d., 1,9 µm particle, Hypersil Gold RP 18 column (Thermo Fisher Scientific) maintained at 25 °C.

Table 1: Gradient for HPLC measurement

Time	Percentage mobile phase B (%)	Remarks
0 → 30	2 → 50	Start of linear gradient 2-50 % B in 30 min
30 → 30,1	50 → 100	
30,1 → 35	100	Washing step
35 → 37	100 → 2	
37 → 43	2	Reequilibration

The mobile phase was a multi step, 43 min, linear gradient prepared from 0,1% HCOOH (A) and CH₃CN (B) (Table 1); the flow rate was 0,35 mL · min⁻¹. The volume of sample solution injected was 5 µL. Peaks were recorded in the range of 200 – 600 nm.

Instrumentation for HPLC-MS analysis:

The HPLC system described above was coupled with an LTQ Orbitrap XL (Thermo Fisher Scientific, Dreieich, Germany). Ionisation was performed with a heated electrospray interface equipped with a metal needle kit. The conditions used for mass spectrometry were as follows: capillary voltage 35 V, spray voltage 3.00 kV, tube lens 120 V. Nitrogen was used as drying gas. The capillary temperature was set at 300 °C. Collision-induced dissociation spectra (CID) were obtained with a collision energy of 35 and an activation time of 30 ms. Full scan spectra from m/z 150-2000 were obtained in positive mode. Aconitine (Figure 3) was used for the optimization of ionization parameters.

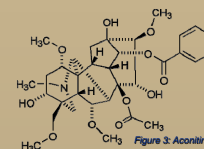


Figure 3: Aconitine

Table 2: LC/MS data of Aconitum napellus tincture

HPTLC zone (R _f) tincture	HPLC Retention time (min)	[M+H] ⁺ (m/z)	Major fragment ions m/z
0.08	7.2	500	482 / 468 / 450 / 436 / 418
	7.9	500	482 / 468 / 450 / 436 / 418
	19.0	618	586 / 558 / 536 / 526
0.47	19.8	618	586 / 568 / 536 / 522
	21.9	632	600 / 582 / 568 / 550
	16.3	604	n.d.
0.51	19.7	618	600 / 586 / 568 / 554 / 536 / 522
	20.3	586	568 / 554 / 536 / 522
	21.7	646	n.d.
0.65	11.4	480	462 / 430 / 412 / 402

Table 3: LC/MS data of an aged reference solution of aconitine

HPTLC zone (R _f) reference solution	HPLC Retention time (min)	[M+H] ⁺ (m/z)	Major fragment ions m/z
0.09	19.1	618	586 / 558 / 536 / 526 / 494
0.46	17.9	590	572 / 558 / 540 / 526 / 508 / 494 / 476 / 444
	19.8	618	586 / 568 / 554 / 536 / 522
0.51	16.3	604	586 / 572 / 554 / 540 / 522
	19.7	618	600 / 586 / 568 / 554 / 536 / 522
	20.2	586	568 / 554 / 536 / 522 / 504
	21.7	646	586 / 568 / 554 / 536 / 526 / 522
0.64	9.6	482	464 / 450 / 432
	19.8	618	600 / 586 / 568 / 554 / 537 / 522 / 504
	20.2	586	568 / 554 / 536 / 522 / 504 / 490

Comparison of the significant zones:

- 1) The zones at R_f 0,08 and 0,09 are corresponding in the detected mass 618 at a retention time of 19,0 and 19,1 minutes. A possible identity of the substance according to the detected mass and its major fragments is described in literature as N-deethyl-aconitine [5]. The detected mass at the retention times 7,2 or 7,9 could be identified as aconitine.
- 2) The zones at R_f 0,47 and 0,46 are corresponding in the detected mass 618 at a retention time of 19,8 minutes. A possible identity of the substance according to the detected mass and its major fragments is described in literature as 8-deacetylunaconitine [3].
- 3) The zones at R_f 0,51 are corresponding in all of the four detected masses at different retention times. The identity of the substance with m/z 646 could be proofed according to its fragmentation as aconitine [3]. The detected mass at the retention time 16,3 could be identified as benzoylaconitine.
- 4) The zones at R_f 0,65 and 0,64 do not correspond in their detected masses.

Conclusion:

The main products of the reference substance aconitine solved in methanol *R* could be detected in aged tincture of Aconitum napellus as well. Most of the examined zones were a mixture of substances with different masses.