

A large, central water droplet is shown falling into a pool of water, creating a dynamic splash with radiating ripples and bubbles. The background is a soft, out-of-focus blue.

**Trinkwasser für Baden-Württemberg**

# **Characterisation of natural waters using HPTLC and toxicity-directed analysis**

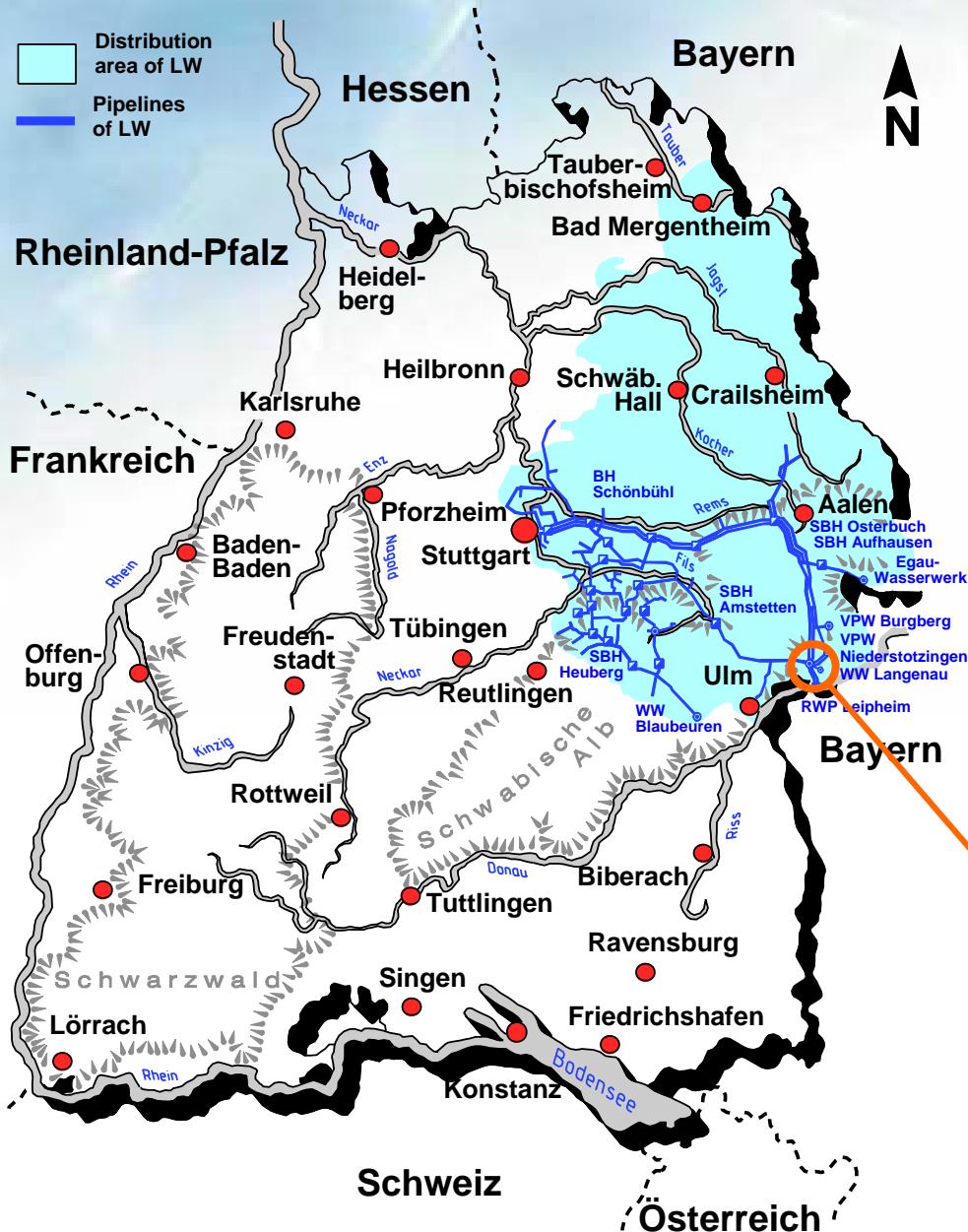
**Sonja Knödler, Wolfram Seitz, Wolfgang Schulz, and Walter H. Weber**

Zweckverband Landeswasserversorgung, Langenau

- **Introduction**
- **Luminescence inhibition test**
- **Data evaluation**
- **Investigation of natural waters**
- **Conclusions**

# Distribution area of LW in South West Germany

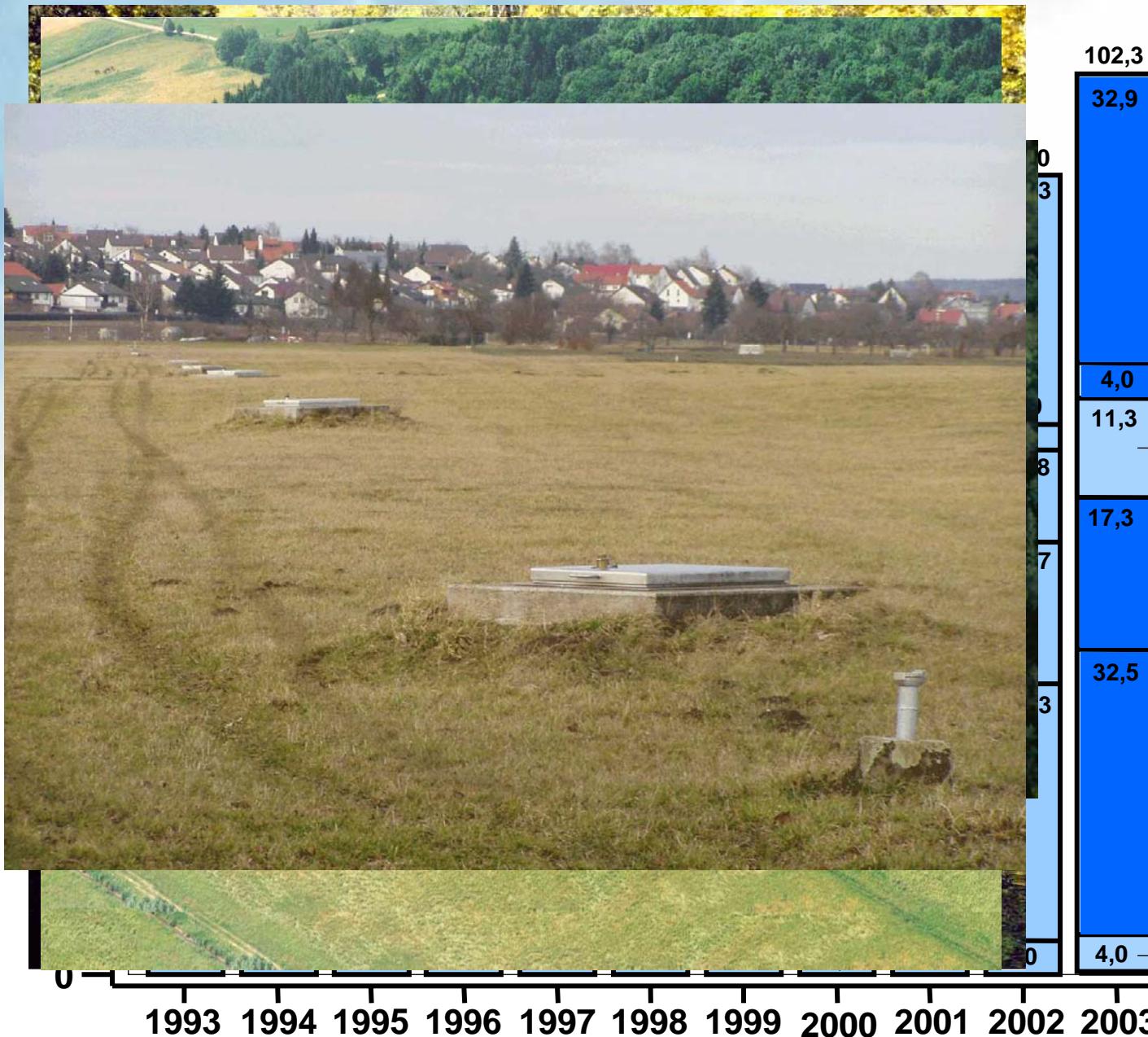
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- Distribution area in South West Germany
  - 3 Million customers within distribution area
  - Long-distance water fraction approx. 50%
- Langenau Waterworks

# Water production 1993 - 2003

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Surface water  
(Danube)

Ground water  
Blaubeuren  
Ground water  
Burgberg

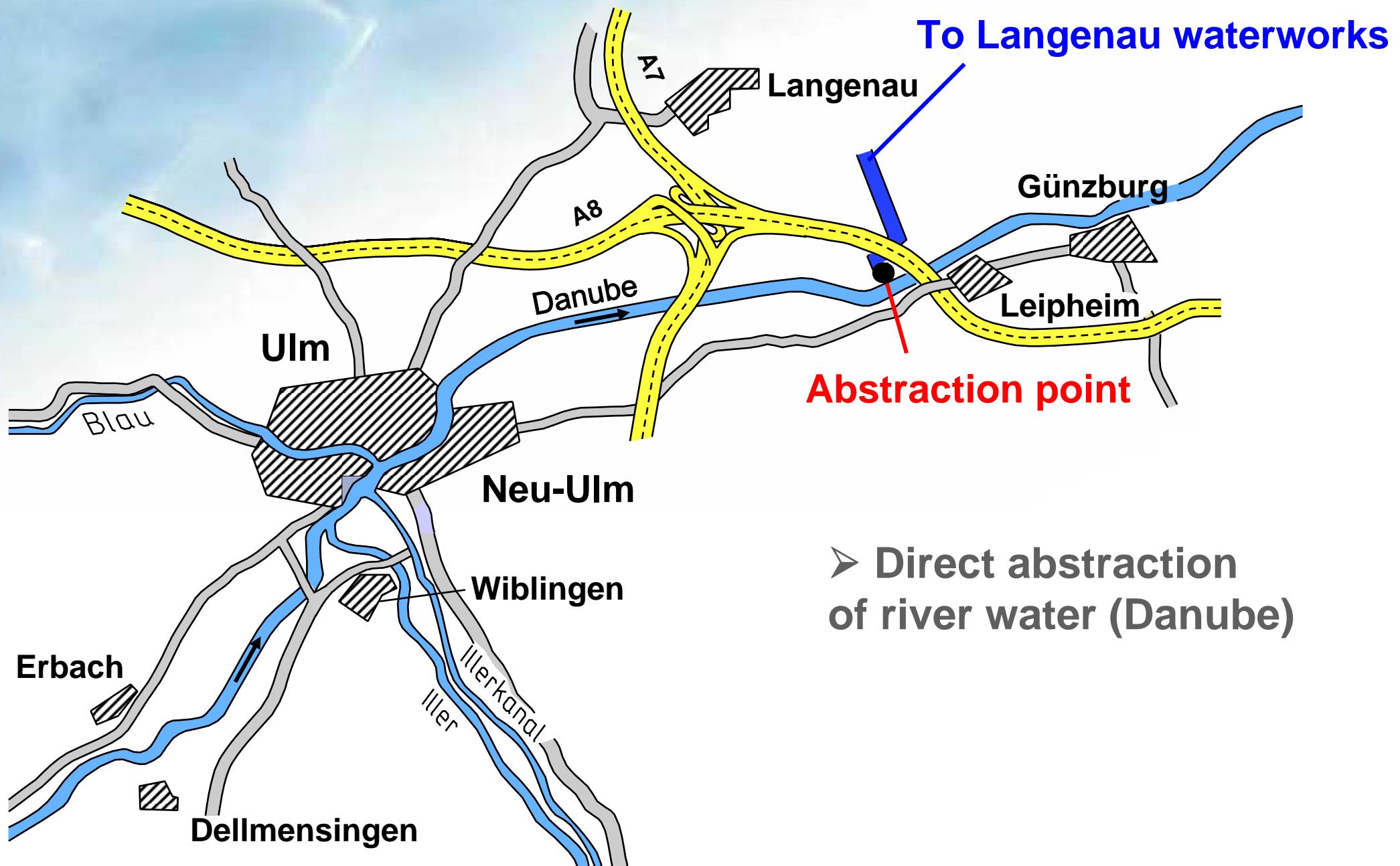
Spring water  
Egau waterworks

Ground water  
Danube Reed

Other

# Langenau waterworks and Leipheim abstraction point

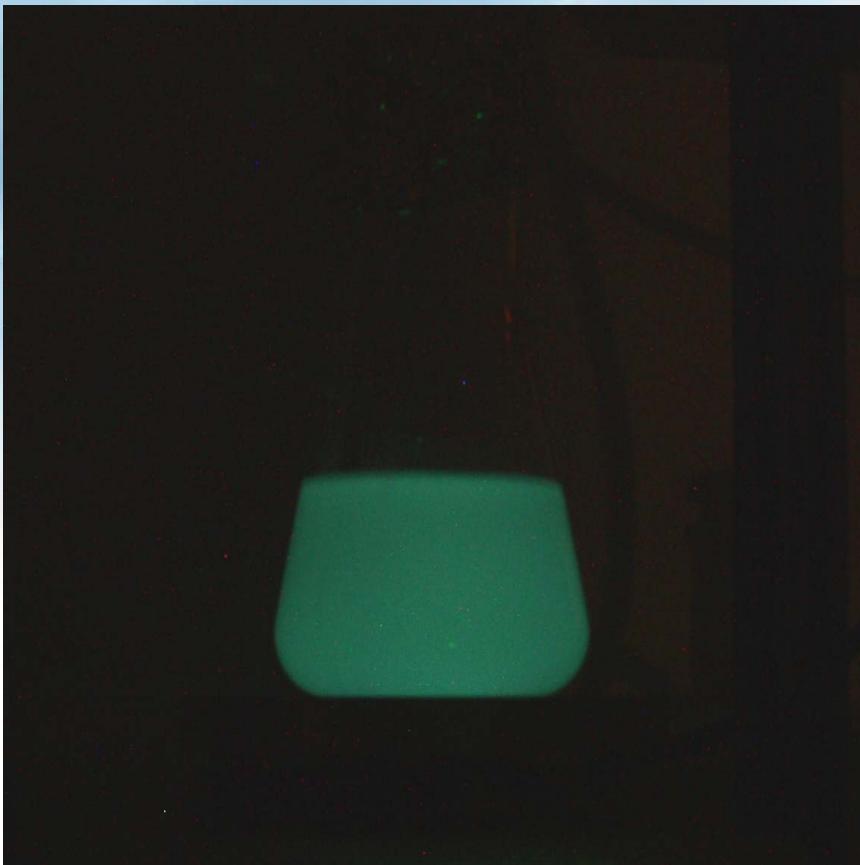
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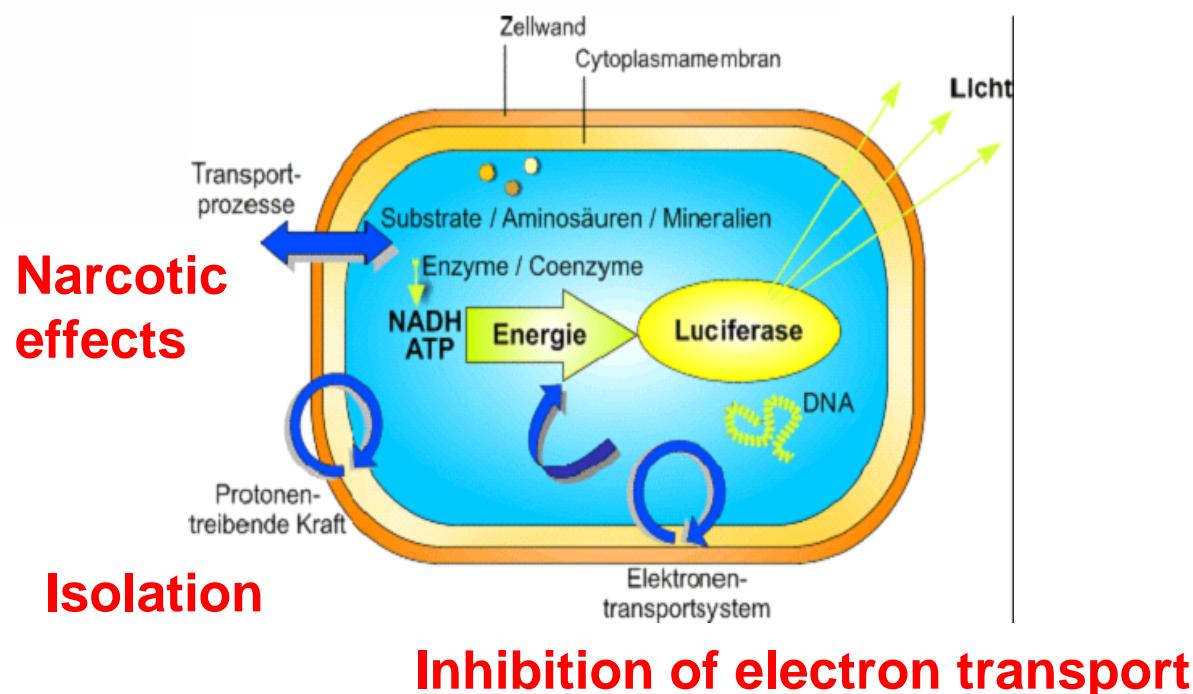
# Luminescence bacteria *Vibrio Fischeri*

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Suspension of *Vibrio fischeri*  
in water

- Marine bacterium
- Lives in symbiosis with marine life forms
- Continuous bioluminescence
- Bioluminescence is coupled to energy metabolism



- Classic application: Cuvette test (DIN 11348)
  - Detection of combination effects of toxic compounds (synergistic effects)



(1) LUMIStox 300  
(2) LUMISterm

# Luminescence inhibition test on TLC plates

Sample  
(extract)



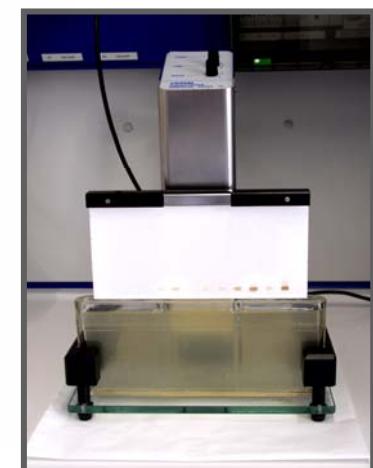
Application  
(TLC Sampler)



Development

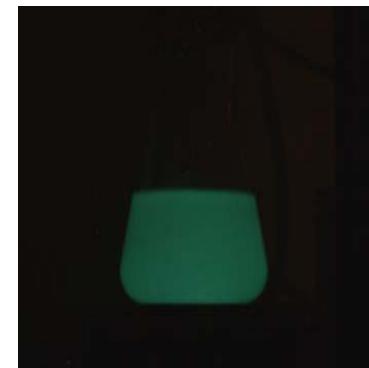
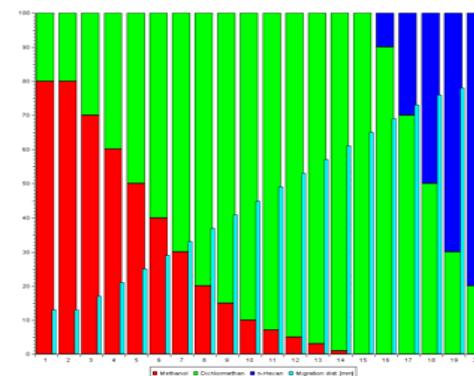


Immersion



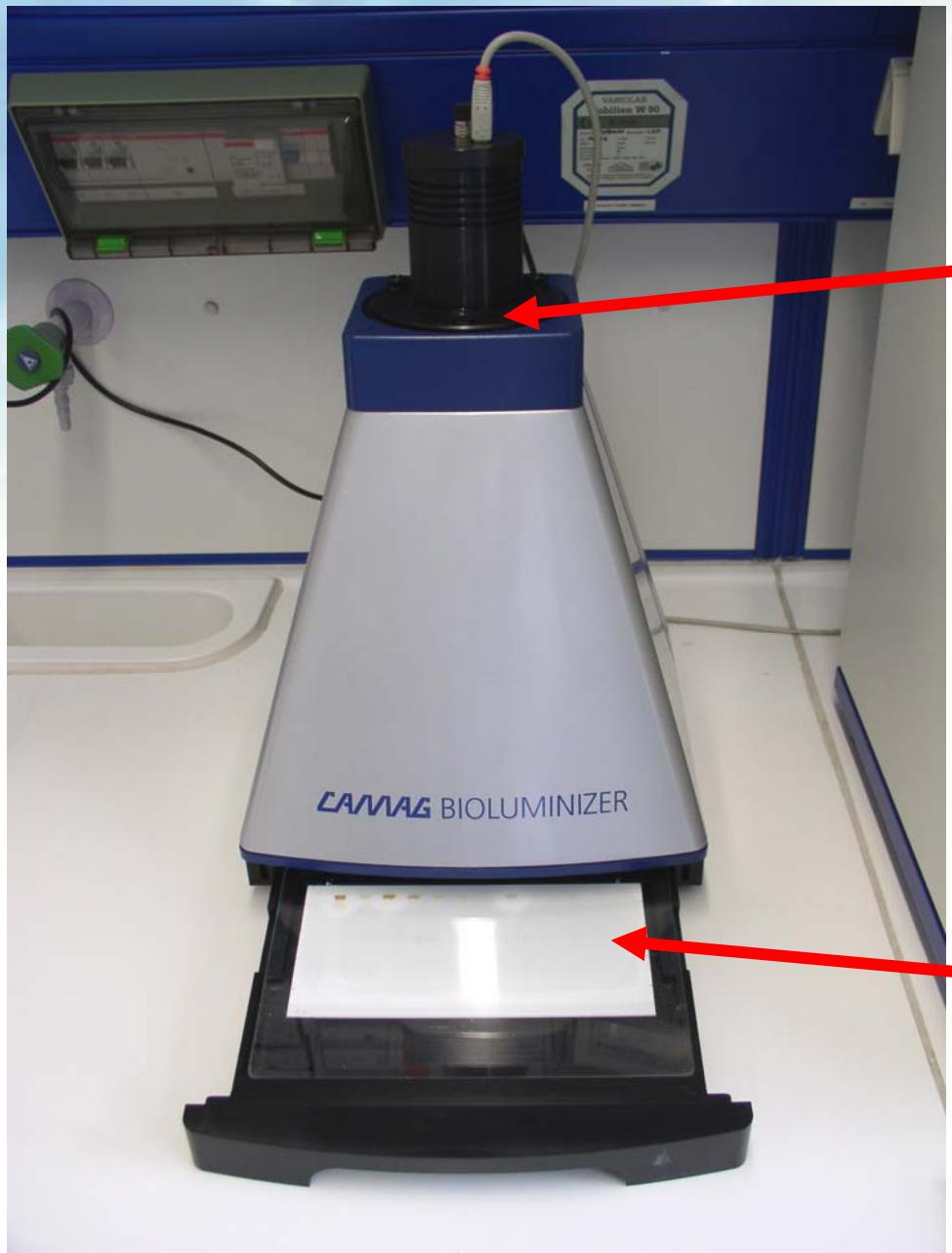
Reference:

Weisemann, C., Kreiss, W., Rast, H-G., Eberz, G.;  
“Analytical Method for Investigating Mixtures for Toxic Components.” European Patent No: EP 0 588 139 B1.



# Detection of luminescence

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**CCD camera**  
**Camag Bioluminizer**  
**(typical detection time 40 sec)**

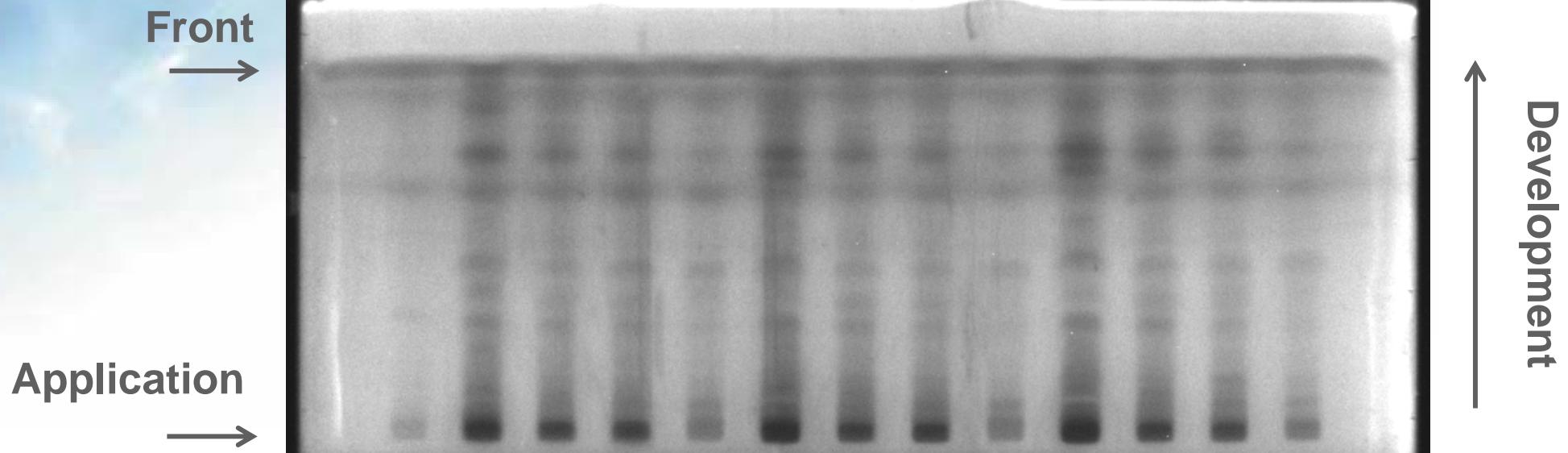
**TLC plate**  
**after immersion**  
**into bacteria suspension**

# Luminescence of TLC plate

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Application example: Analysis of different waste water extracts



Front

↑

Development

Effluent (pilot plant)

Effluent (A)

Effluent

Influent

Effluent (pilot plant)

Effluent (A)

Effluent

Influent

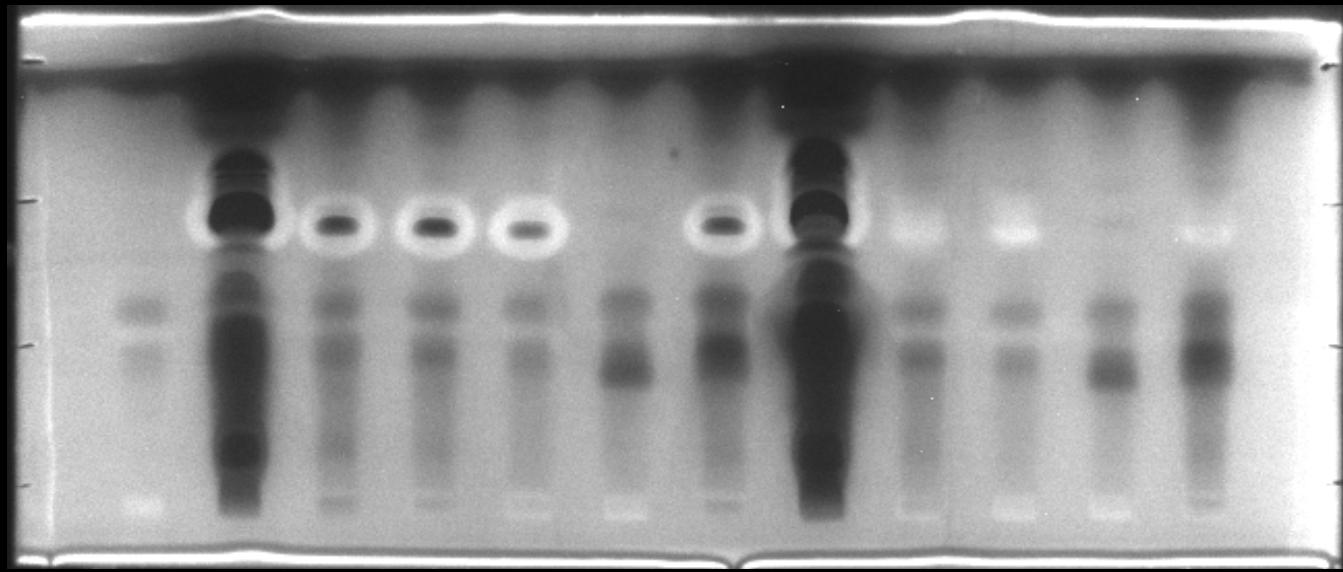
Effluent (pilot plant)

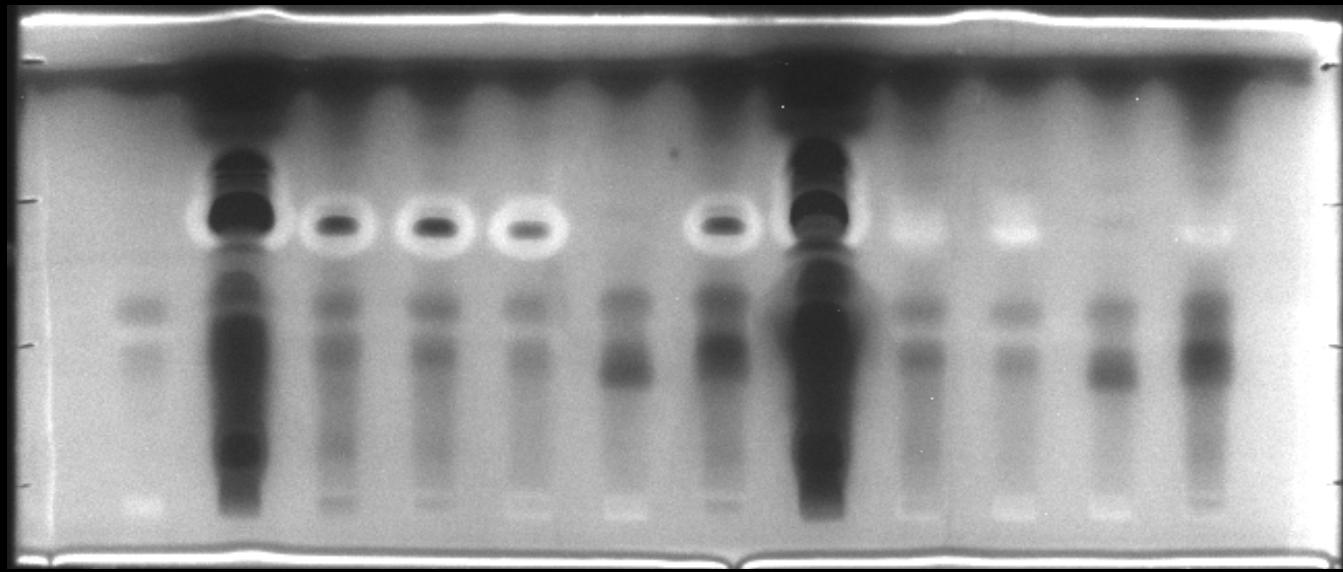
Effluent (A)

Effluent

Influent

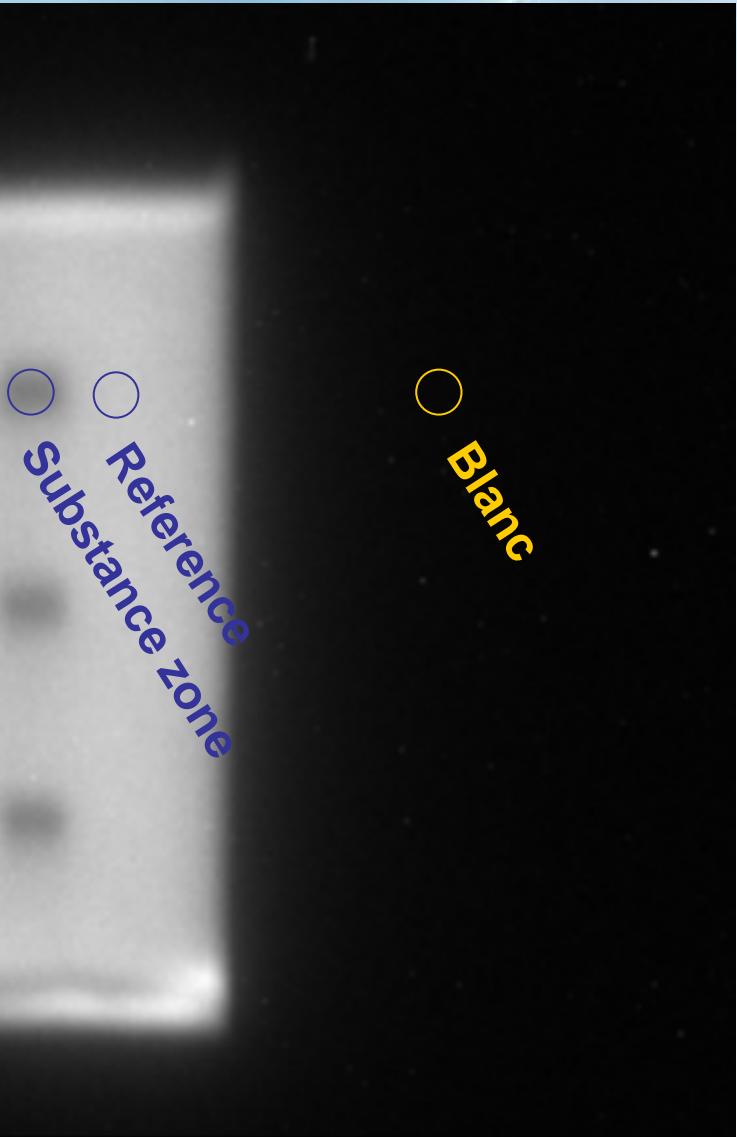
Blanc





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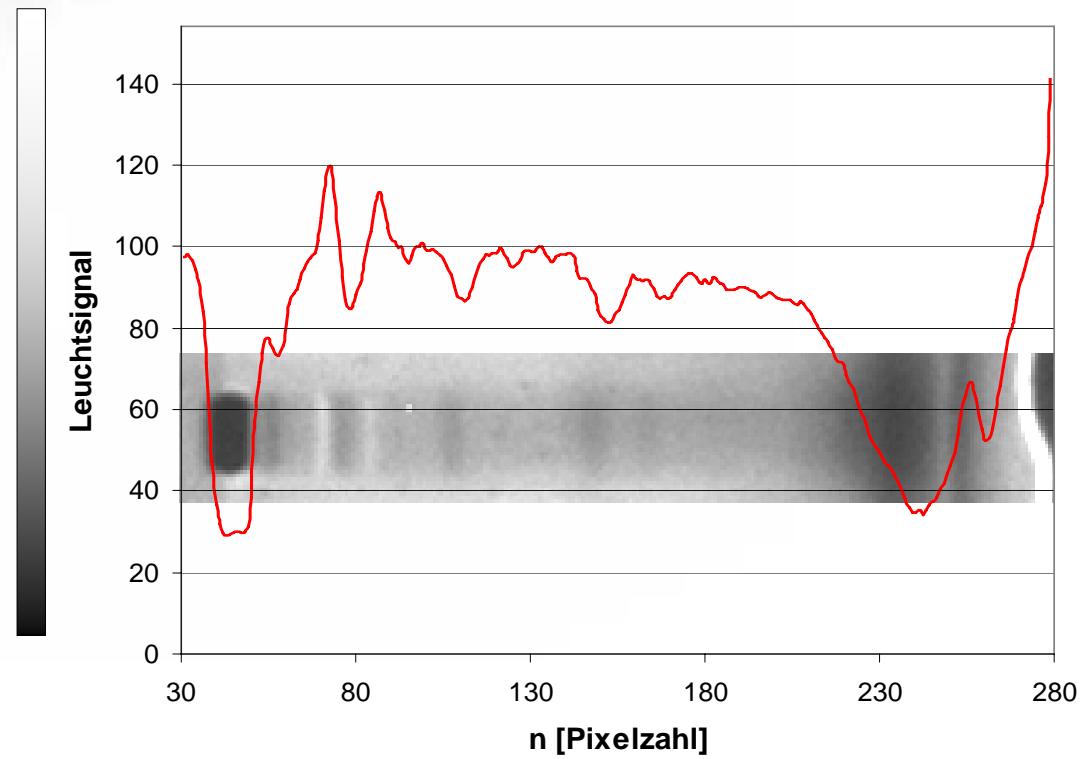
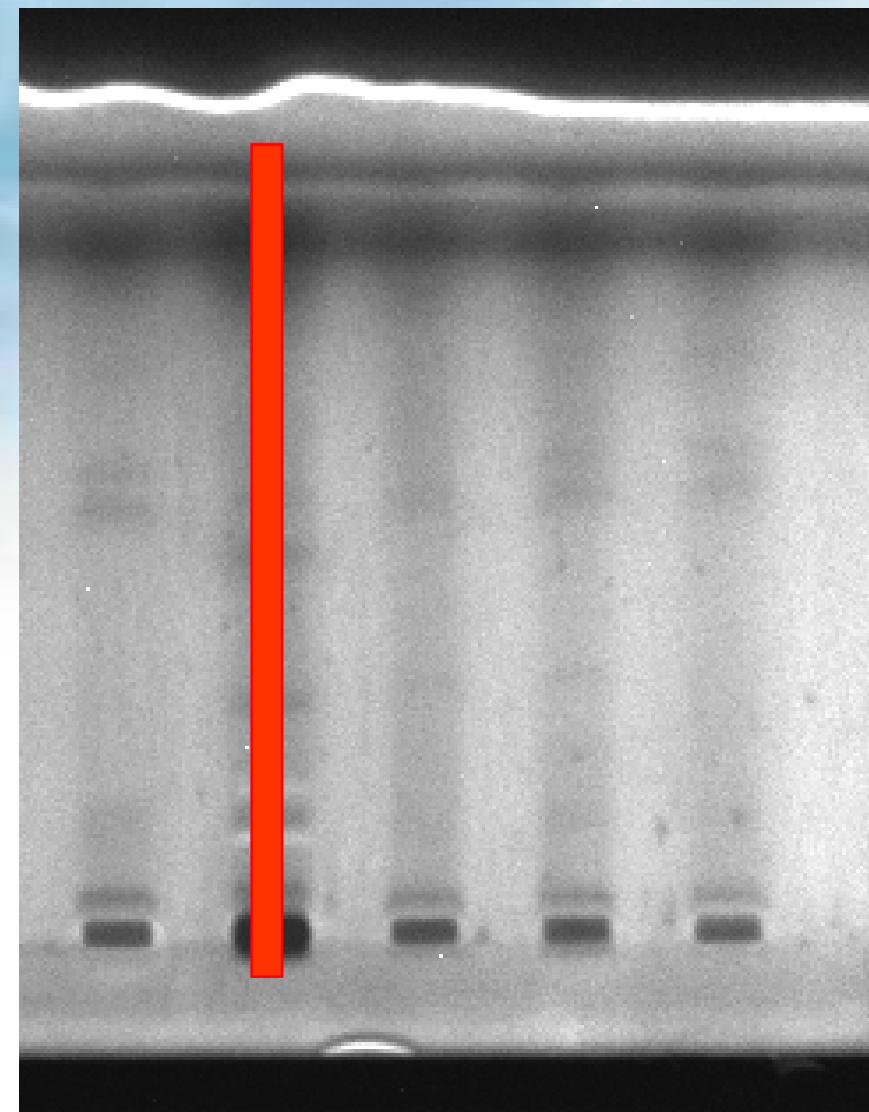
# Calculation of inhibition values



$$\text{Inhibition} = \frac{\text{Reference intensity} - \text{Substance intensity}}{\text{Reference intensity} - \text{Blanc intensity}}$$

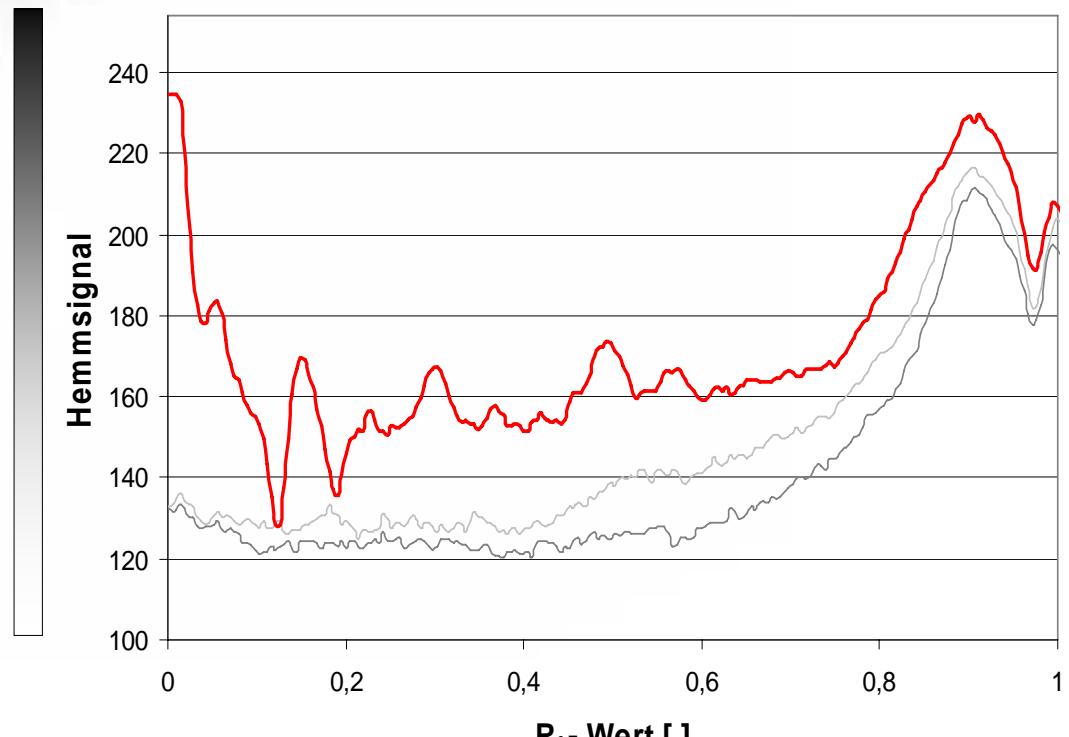
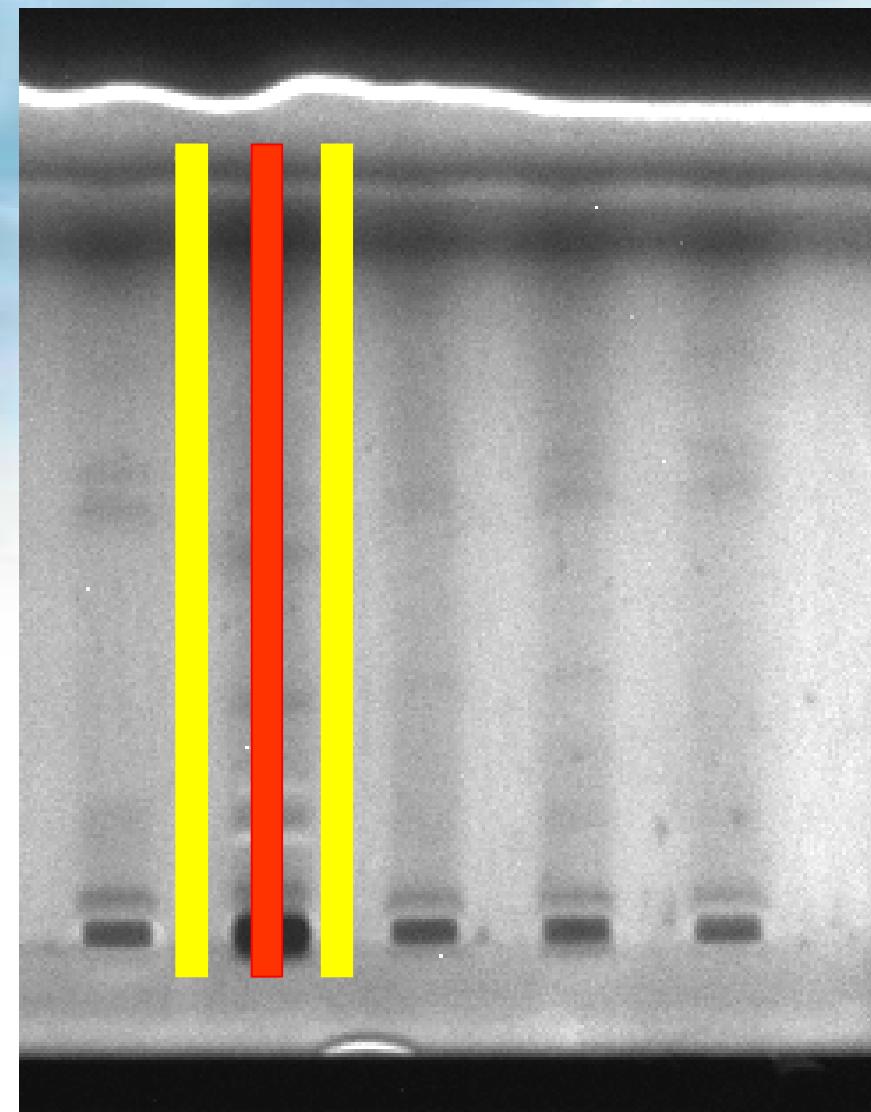
- **Simple method to evaluate single substance zones**

# Advanced data evaluation



- Selection of sample line (light intensity)

# Advanced data evaluation

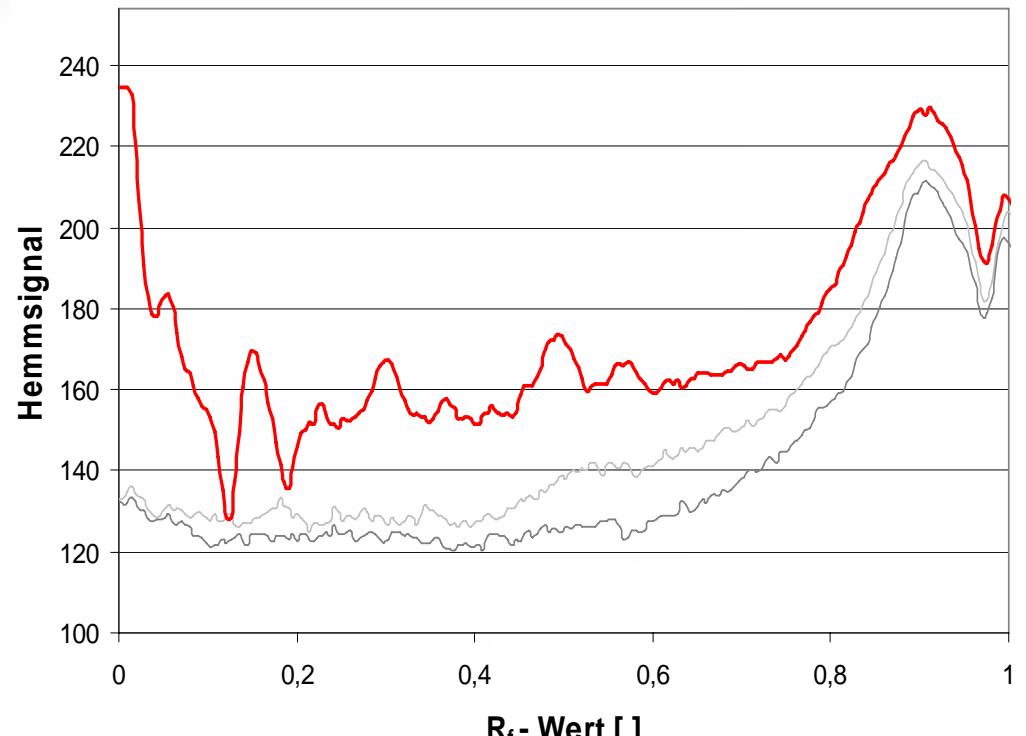


- Selection of reference lines
- Inversion of light intensity into inhibition

# Advanced data evaluation

$$h^U = \frac{\sum_{n=n_1}^{n_2} (i_0 - i_n^U)}{(n_{2+1} - n_1) \cdot i_0} = 1 - \frac{\sum_{n=n_1}^{n_2} i_n^U}{(n_{2+1} - n_1) \cdot i_0}$$

$$h^P = \frac{\sum_{n=n_1}^{n_2} (i_0 - i_n^P)}{(n_{2+1} - n_1) \cdot i_0} = 1 - \frac{\sum_{n=n_1}^{n_2} i_n^P}{(n_{2+1} - n_1) \cdot i_0}$$

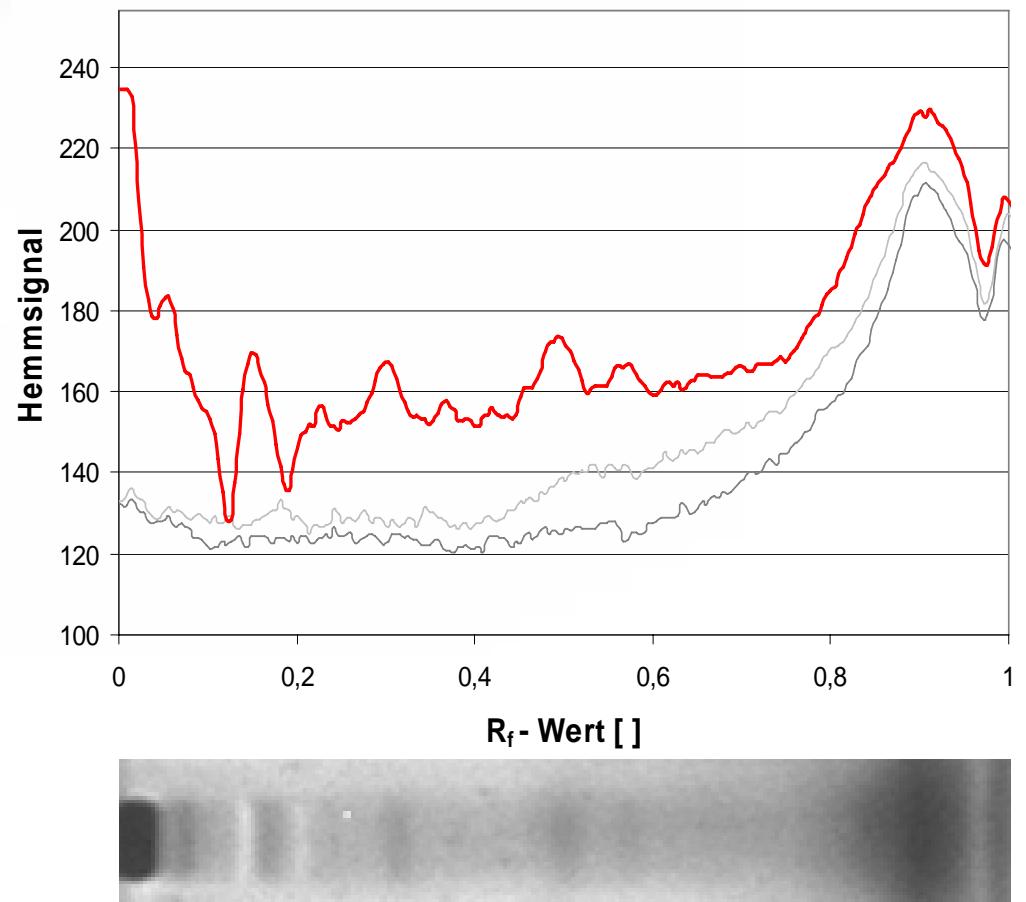


- Calculations ...

# Advanced data evaluation

$$H^P = \frac{h^P - h^U}{1 - h^U} = \frac{\frac{\sum_{n=n_1}^{n_2} i_n^U}{(n_{2+1} - n_1) \cdot i_0} - \frac{\sum_{n=n_1}^{n_2} i_n^P}{(n_{2+1} - n_1) \cdot i_0}}{\frac{\sum_{n=n_1}^{n_2} i_n^P}{(n_{2+1} - n_1) \cdot i_0}}$$

$$= \frac{\frac{\sum_{n=n_1}^{n_2} i_n^U}{(n_{2+1} - n_1) \cdot i_0} - \frac{\sum_{n=n_1}^{n_2} i_n^P}{(n_{2+1} - n_1) \cdot i_0}}{\frac{\sum_{n=n_1}^{n_2} i_n^P}{(n_{2+1} - n_1) \cdot i_0}} = 1 - \frac{\frac{\sum_{n=n_1}^{n_2} i_n^P}{(n_{2+1} - n_1) \cdot i_0}}{\frac{\sum_{n=n_1}^{n_2} i_n^U}{(n_{2+1} - n_1) \cdot i_0}}$$



- Calculations ...

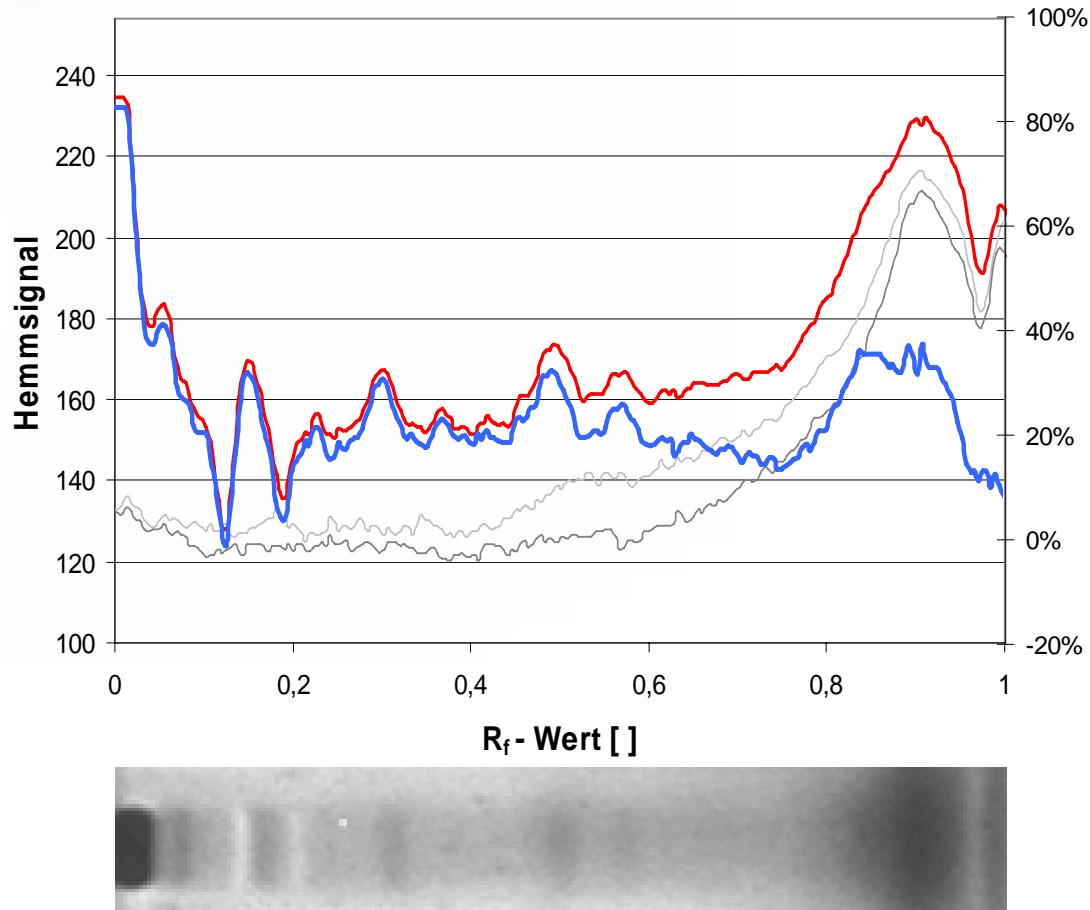
# Advanced data evaluation – inhibition values

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$$I_n^S = 1 - \frac{i_n^S}{i_n^R}$$

I = Inhibition value  
i = light intensity  
S = Sample  
R = Reference  
n = number of pixel



- Calculation of chromatograms  
(Inhibition value vs. migration distance)

# Poster “Quantification of luminescence inhibition on TLC plates”

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**Quantification of luminescence inhibition on TLC plates**  
Wolfgang Schulz, Wolfram Seitz, and Walter H. Weber

**Zweckverband Landeswasserversorgung LW**

**Objectives**

- Development of an algorithm for evaluation of luminescence inhibition on TLC plates
- Comparison of TLC plate test with classic cuvette test
- Application in water analysis

**Conclusions**

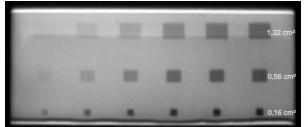
- Sensitivity of luminescence inhibition test on TLC plate was found to be very high compared to cuvette test
- New luminescence inhibition was detected in treated process waste waters using TLC test

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**Method comparison**

**Luminescent inhibition test on TLC plates**

- Application of substance onto silica gel plates
- Submerging into bacteria suspension
- Taking a picture After the Incubation time of 10 min
- Determination of the Inhibition values by a special software.



$I^S = 1 - \frac{\sum_{n_1}^{n_2} i_n^S}{\sum_{n_1}^{n_2} i_n}$

Dose effect relationship of Bromoxynil – TLC plate test –  
EC<sub>50</sub> = 77 ng/cm<sup>2</sup>

**Luminescent inhibition cuvette test**

- Sample + 2 % NaCl
- Addition of bacteria suspension
- Incubation time of 30 min at 15 °C (LUMIStherm)
- Detection (LUMISTox 300)



(1) LUMISTox 300 (2) LUMIStherm

Dose effect relationship of Bromoxynil – Cuvette test –  
EC<sub>50</sub> = 15 ng/µl

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**Sensitivity of TLC test is increased by a factor (F) of 1440 for pesticide Bromoxynil**

O=C#Cc1ccc(Br)c(O)cc1

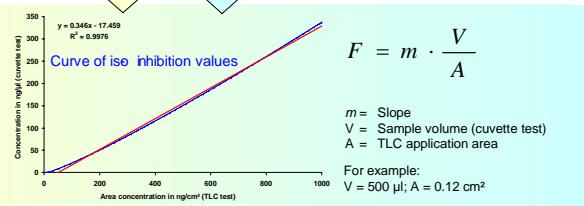
$y = 0.344x + 17.459$   
 $R^2 = 0.9976$

$F = m \cdot \frac{V}{A}$

m = Slope  
V = Sample volume (cuvette test)  
A = TLC application area

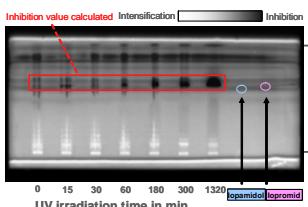
For example:  
V = 500 µl; A = 0.12 cm<sup>2</sup>

**Curve of iso inhibition values**

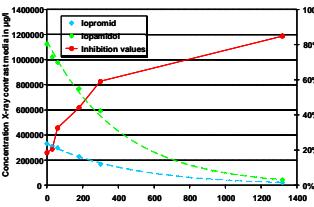


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**Application example: Investigation of UV oxidation by-products**



Inhibition value calculated  
Intensification  
Inhibition  
Rf = 1  
Chromatography  
Rf = 0  
Iopamidol  
Iopromid  
UV irradiation time in min



Concentration X-ray contrast medium (ng/ml)  
Iopamidol  
Iopromid  
Inhibition values  
Irradiation time in min  
Luminescent bacteria inhibition value

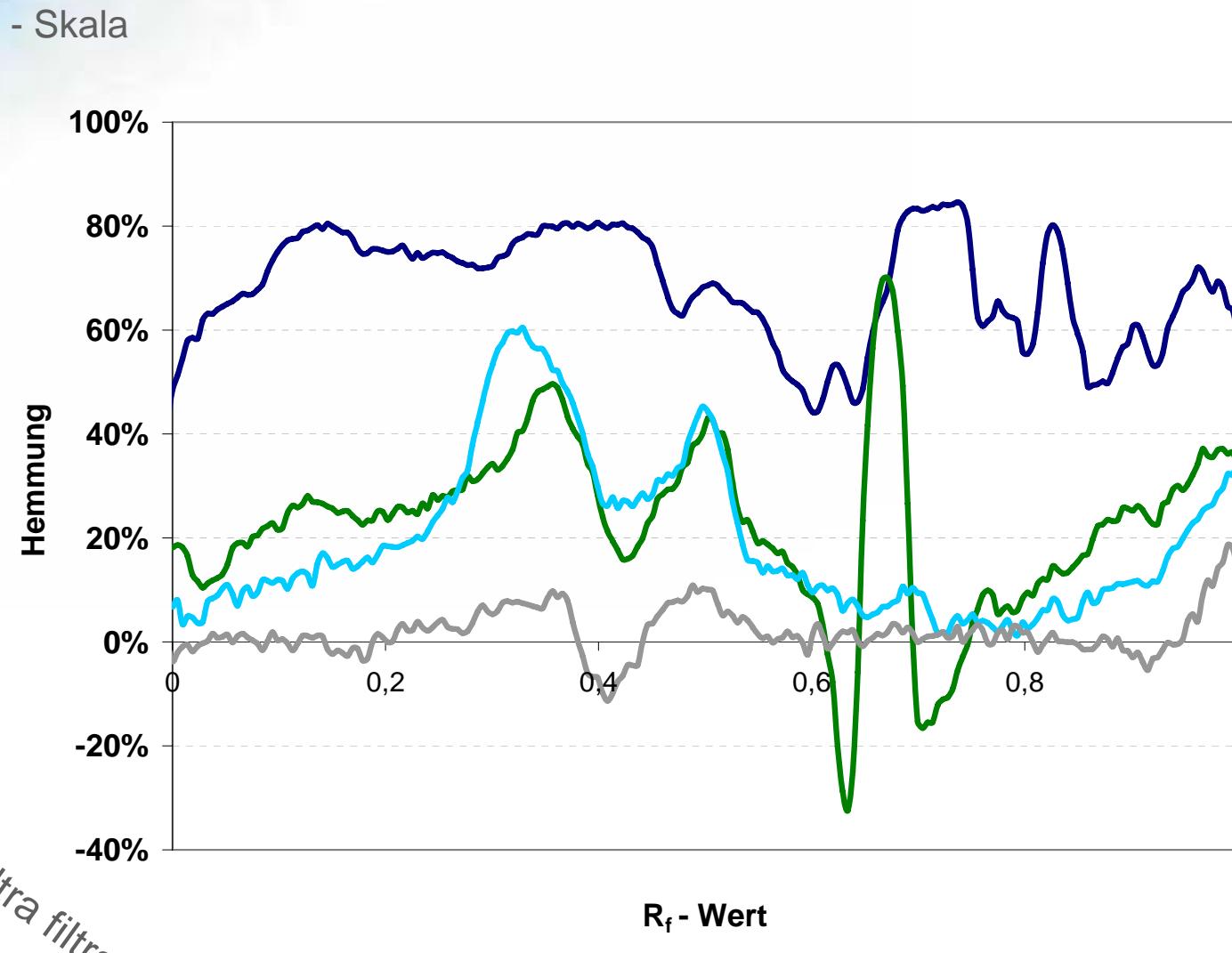
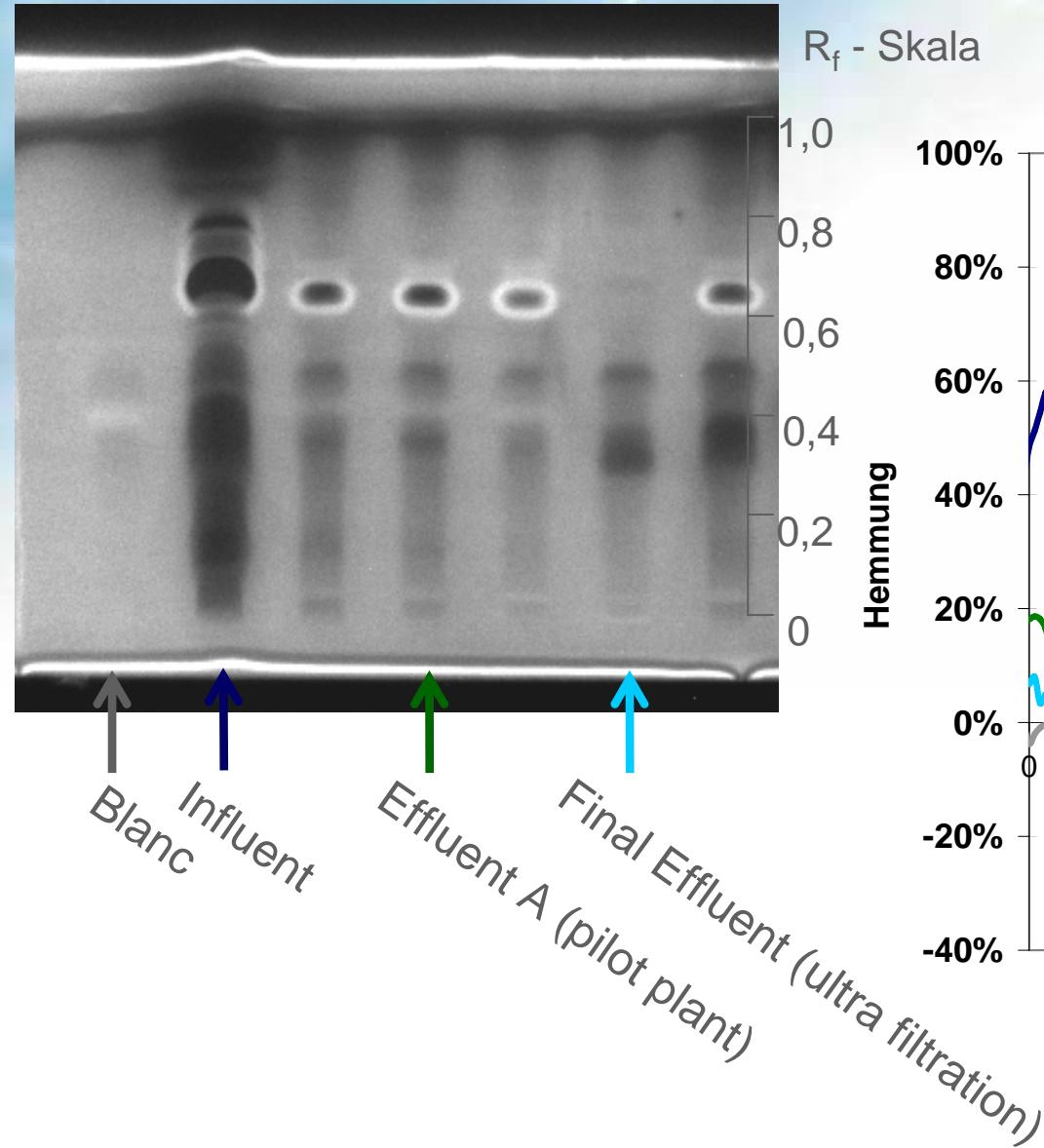
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Tel. +49 (0) 7345-9638-2291 E-Mail: schulz.w@lw-online.de

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# Different inhibition chromatograms of waste water extracts\*

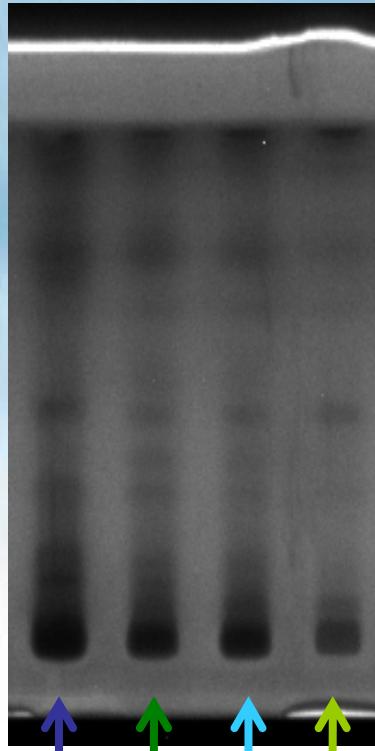
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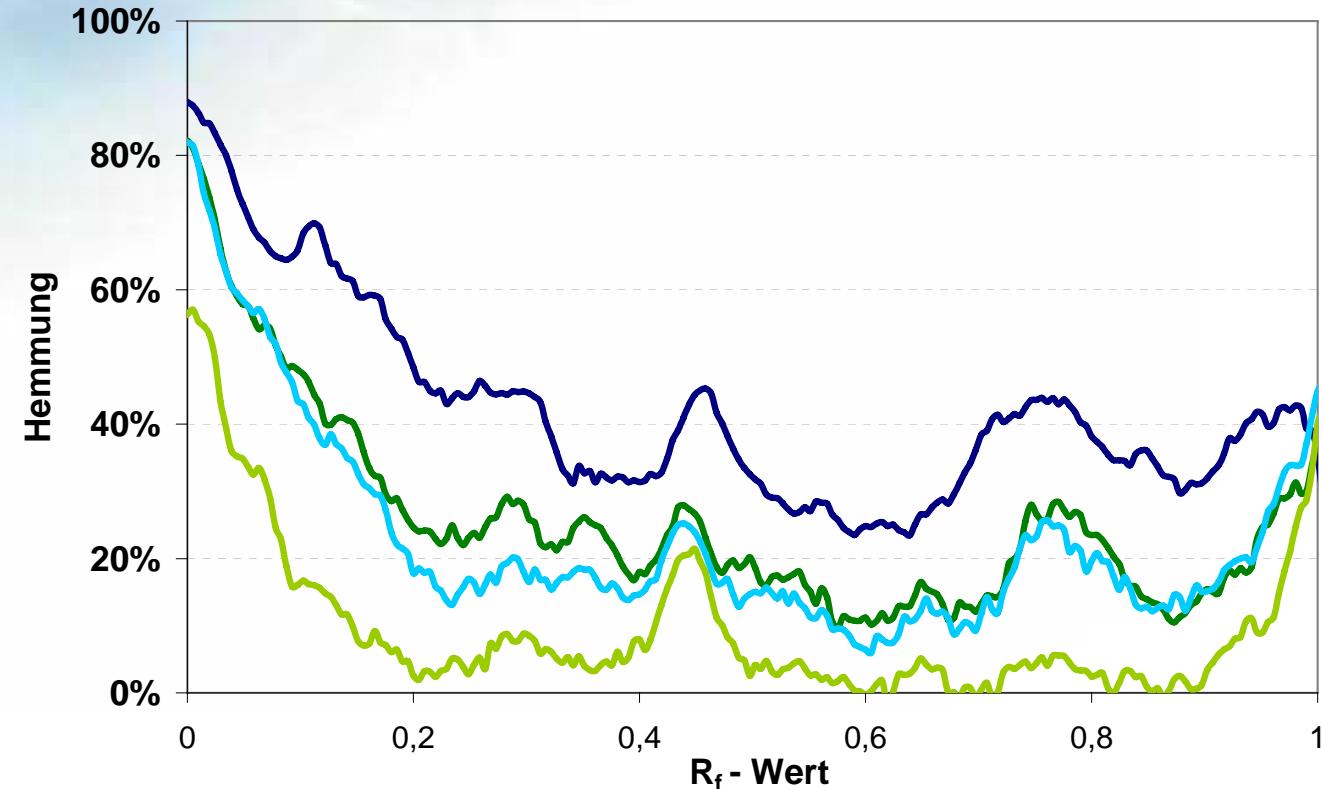
\* Enrichment factor = 1000

# Comparison of waste water influent and effluents\*

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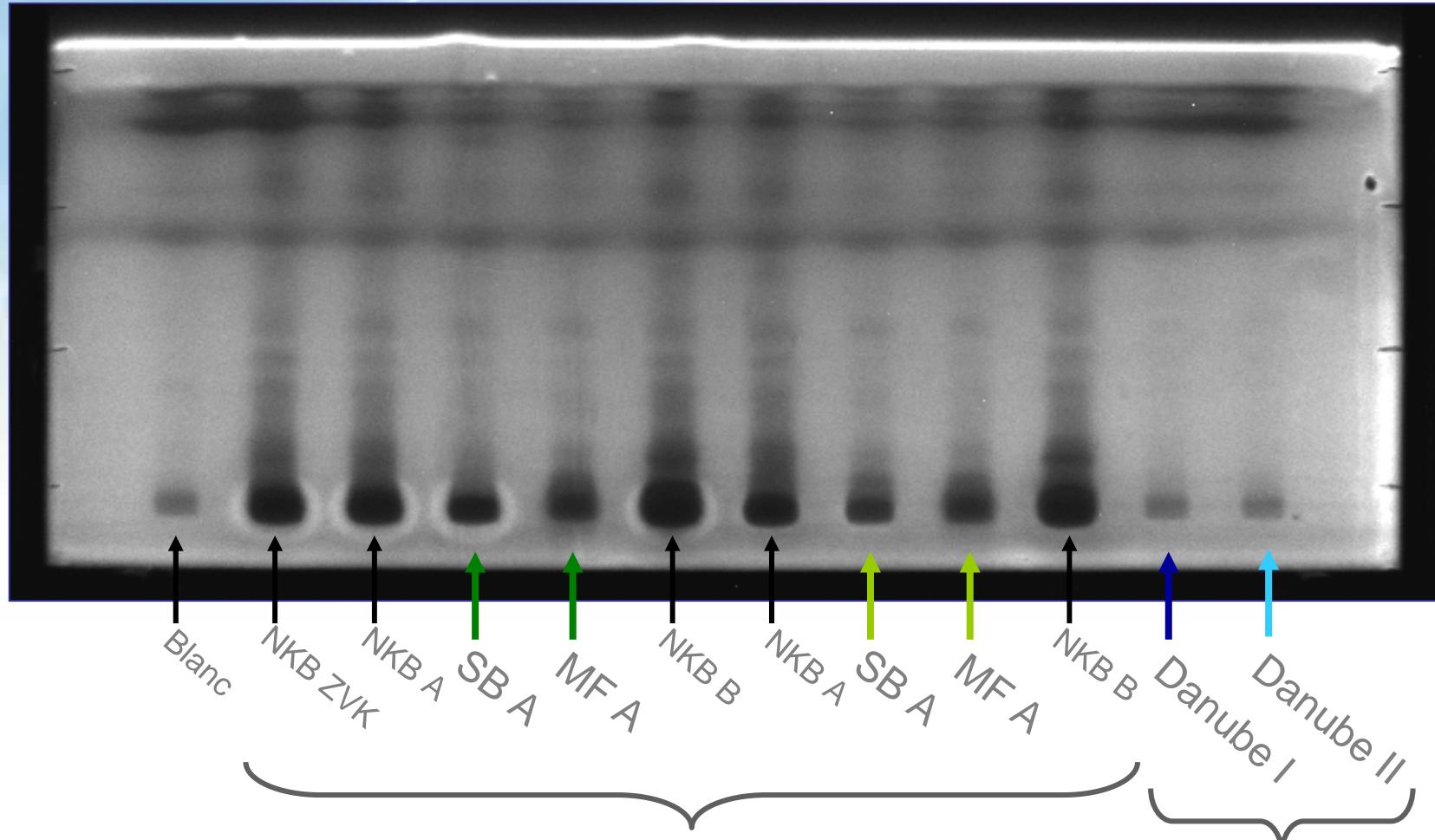
Influent  
Secondary effluent  
Effluent A (Pilot plant)  
Final Effluent (sedimentation)



\* Enrichment factor = 1000

Dosage of powdered activated carbon: 10 mg/L

# Comparison of waste waters with river water\*



**Waste water extracts**

**River water extracts**

# Investigation of process waste waters from X-ray contrast media production

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**UV irradiation time in min**

Co-operating laboratory:  
Heinrich-Sontheimer-Laboratorium,  
Karlsruhe

# Analysis of process waste waters using HPTLC and luminescence inhibition detection\*

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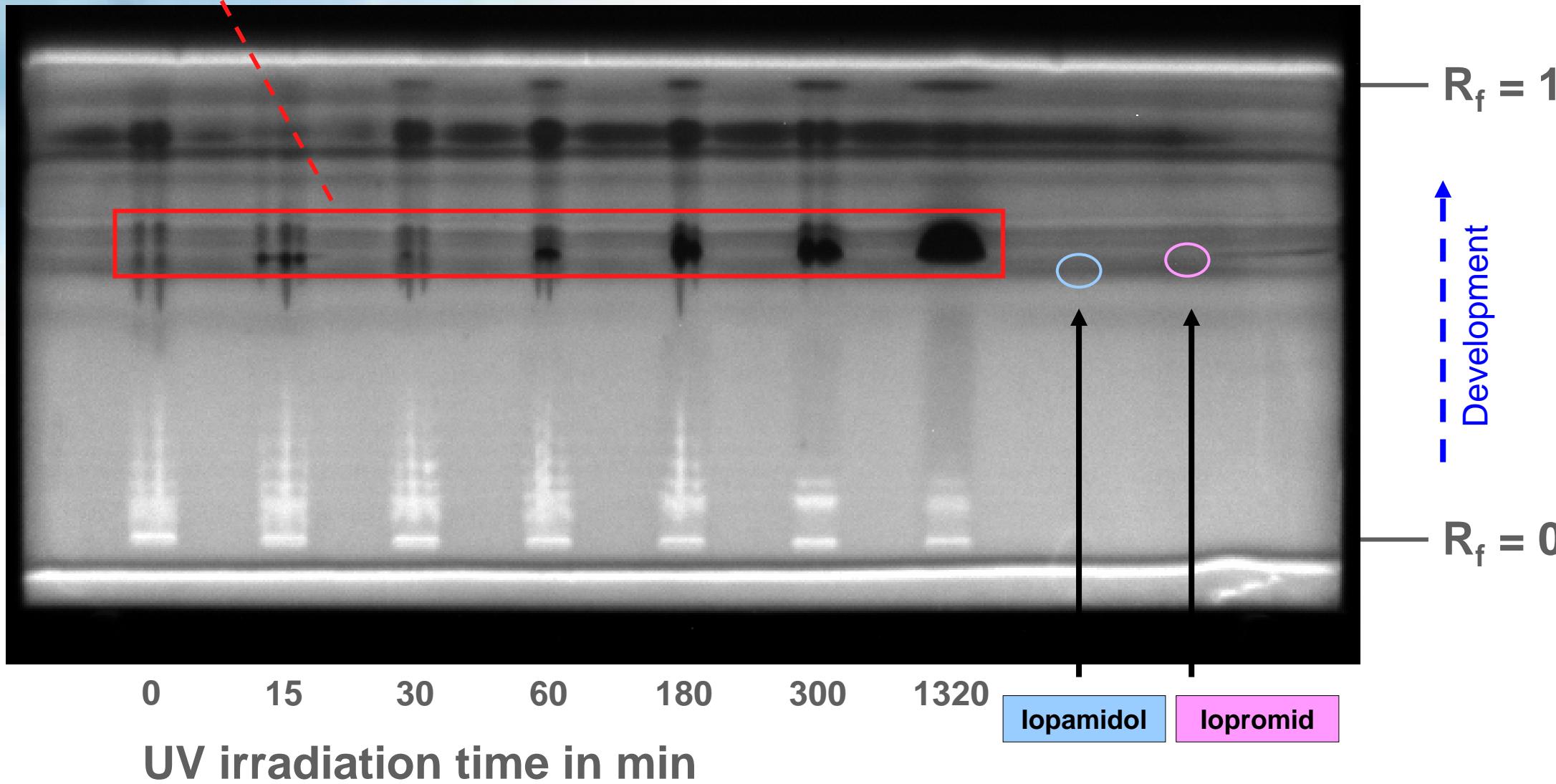


Inhibition values calculated

Intensification

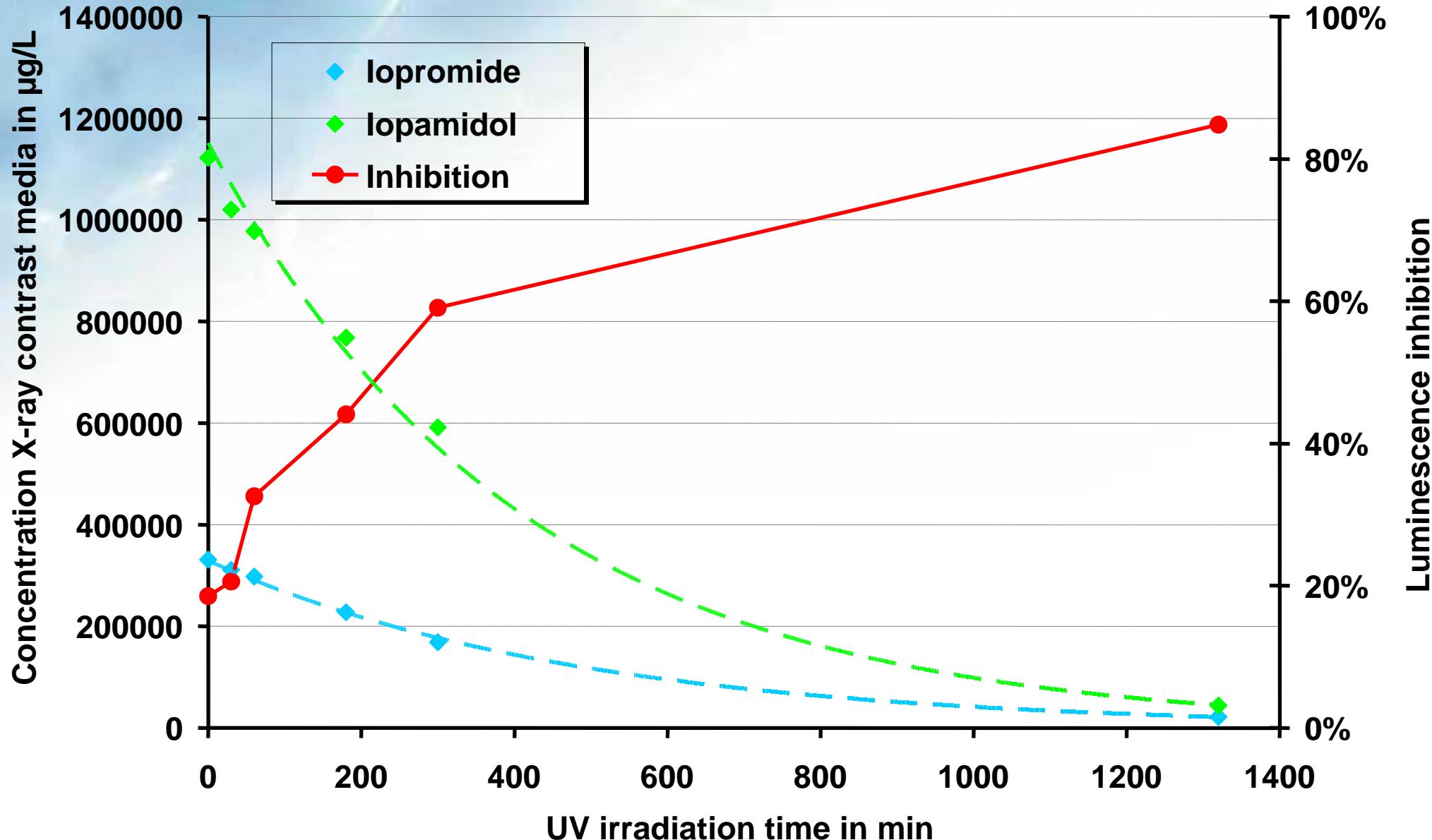


Inhibition



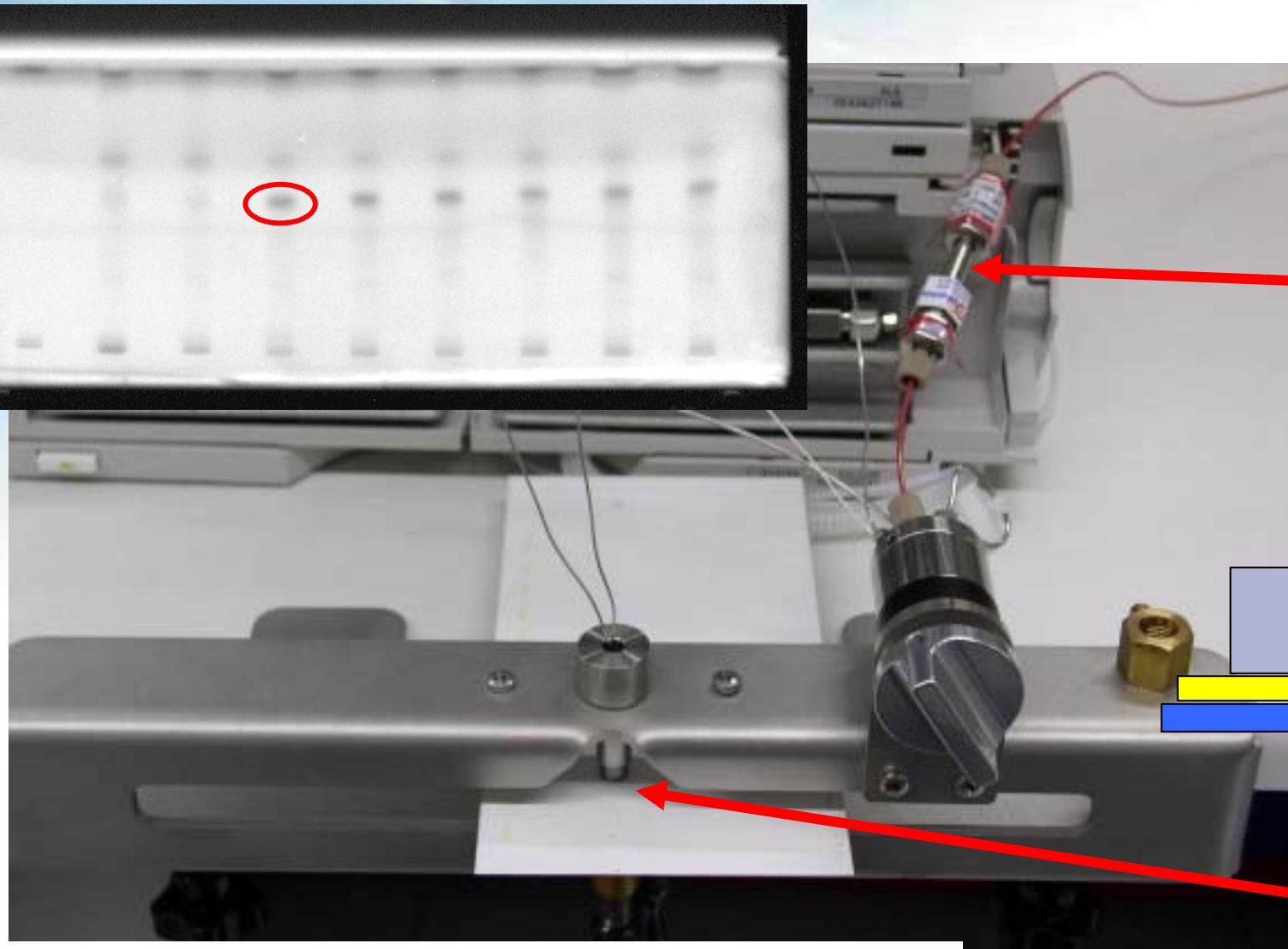
\* Direct application

# Behaviour of luminescence inhibition during the UV treatment process



# Extraction of TLC spots for further investigation using MS detection

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HPLC column

Plunger with  
cutting edge  
and filter

TLC plate

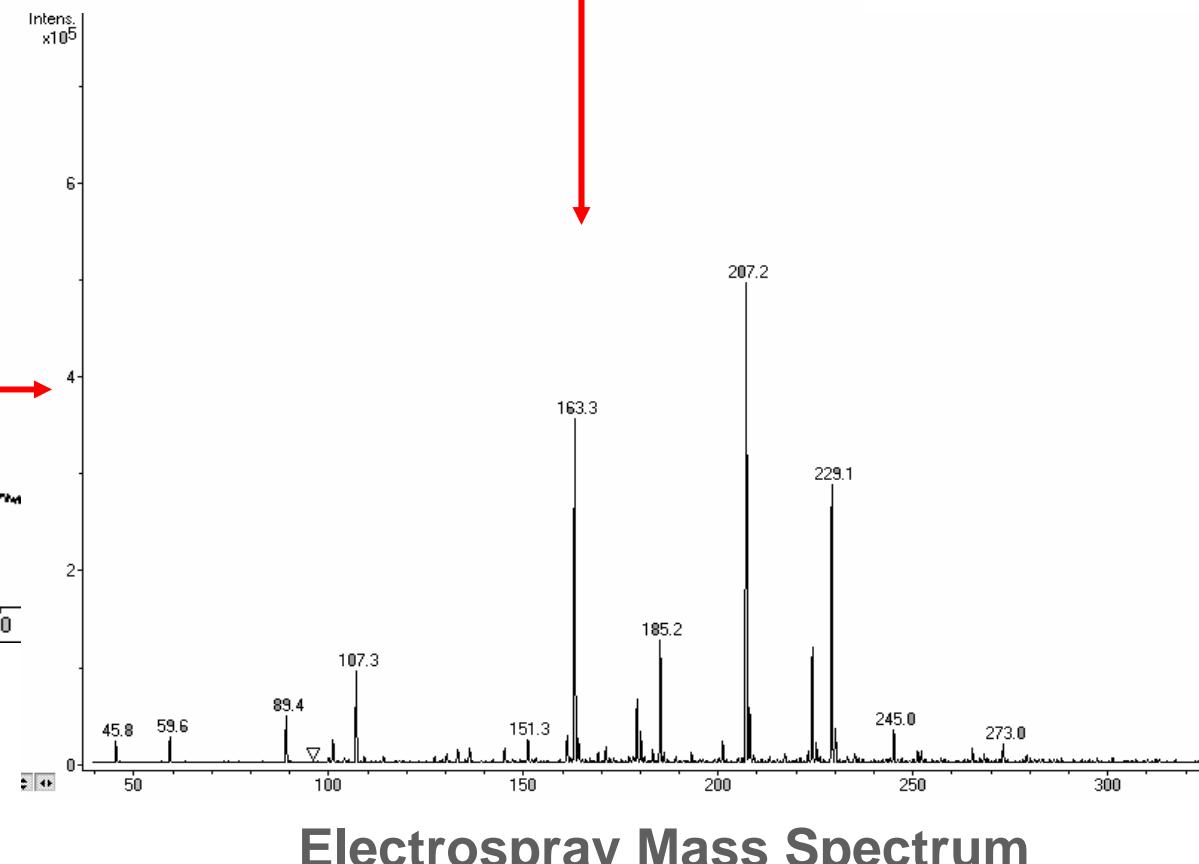
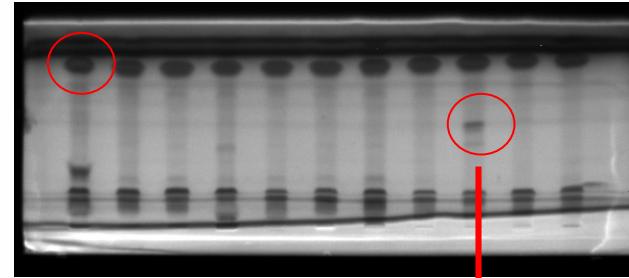
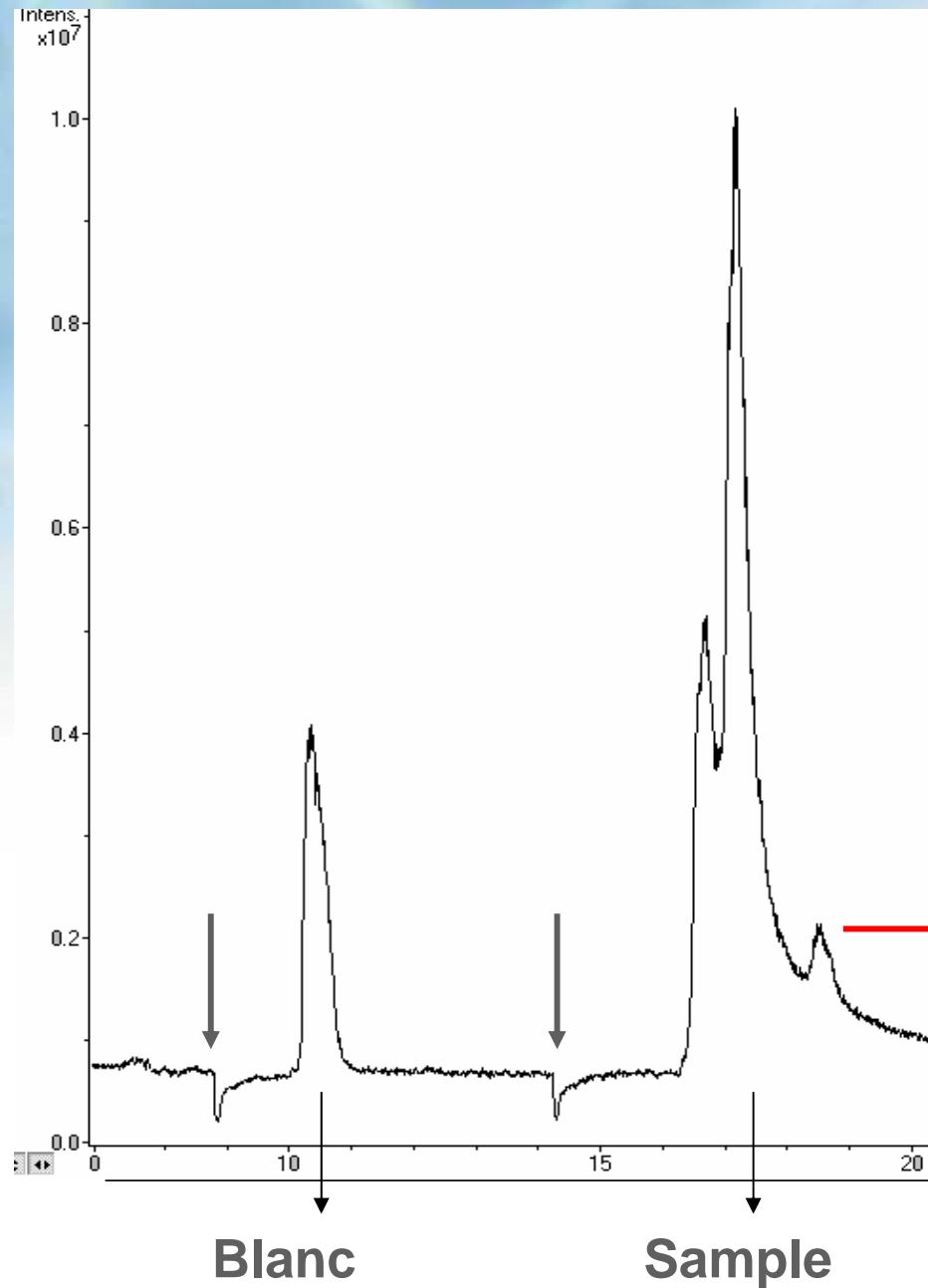
Extractor

Luftmann, H. (2004); A simple device for the extraction of  
TLC spots: direct coupling with an electrospray mass spectrometer,  
Journal of Analytical Bioanalytical Chemistry, 378, 964-968

TLC Extractor ChromeXtract  
(ChromAn, Leipzig, Germany)

# TLC extraction with Online-LC-MS detection

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- CDD camera showed high sensitivity
- Evaluation of single spots and calculation of chromatograms possible
- Interpretation of results is still challenging
- Further bio-activity based detection systems for HPTLC

**Thank you for your attention!**

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Landeswasserversorgung



Laboratory for Operation Control and Research, Langenau



A large, central water droplet is shown falling into a pool of water, creating a dynamic splash with radiating ripples and bubbles. The background is a soft, out-of-focus blue.

**Trinkwasser für Baden-Württemberg**

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