



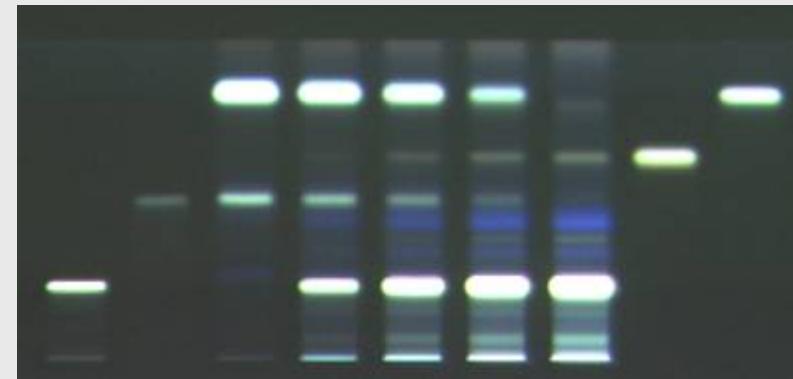
# Standardized HPTLC Methods

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For identification of botanicals and detection of adulteration

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## What is HPTLC?

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**High Performance** Thin-Layer Chromatography

### Key elements

- ▶ Instruments for all steps
  - Application
  - Development
  - Documentation
  - Densitometry
- ▶ Standardized methodology
- ▶ Validated methods
- ▶ Full cGMP compliance

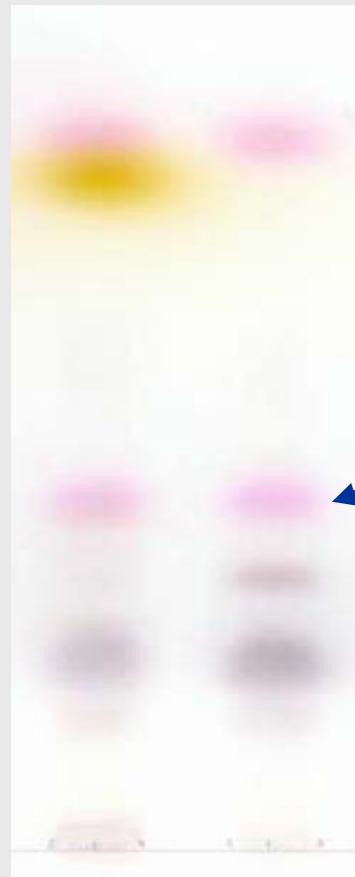


# Standardized Methodology

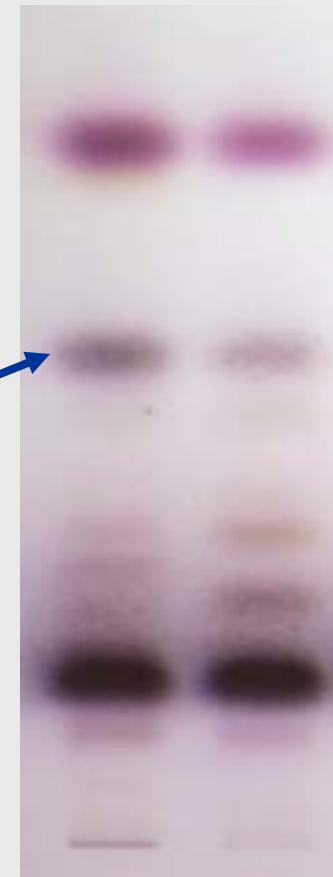
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## Is there a need for standardization?

Lab A



Lab B



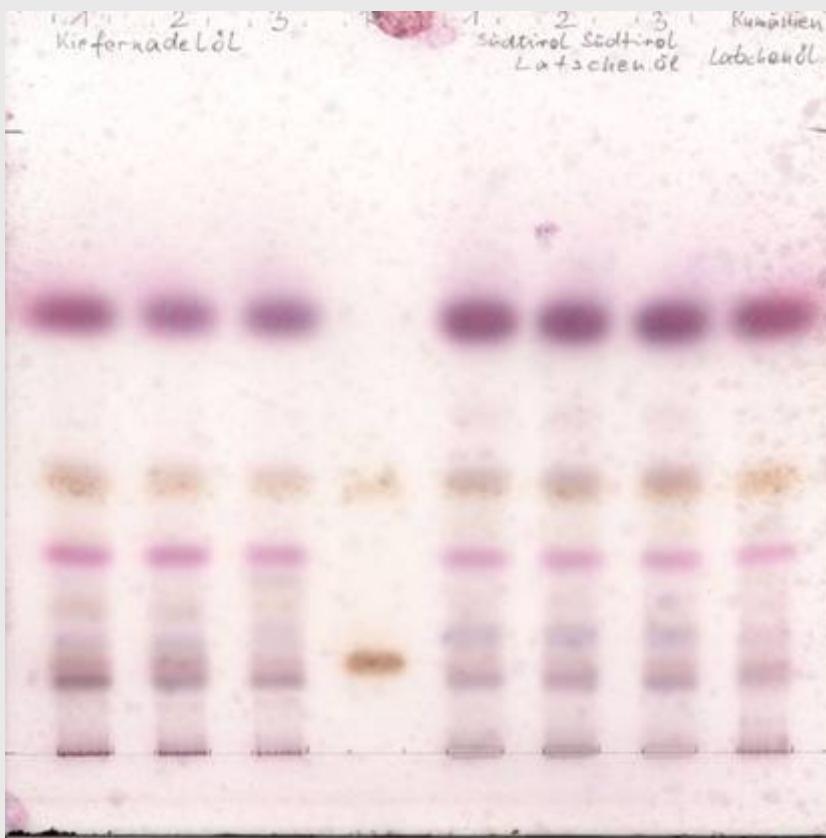
?



Identification test for Peppermint and Spearmint oil, Ph.Eur.5

## TLC or HPTLC?

TLC

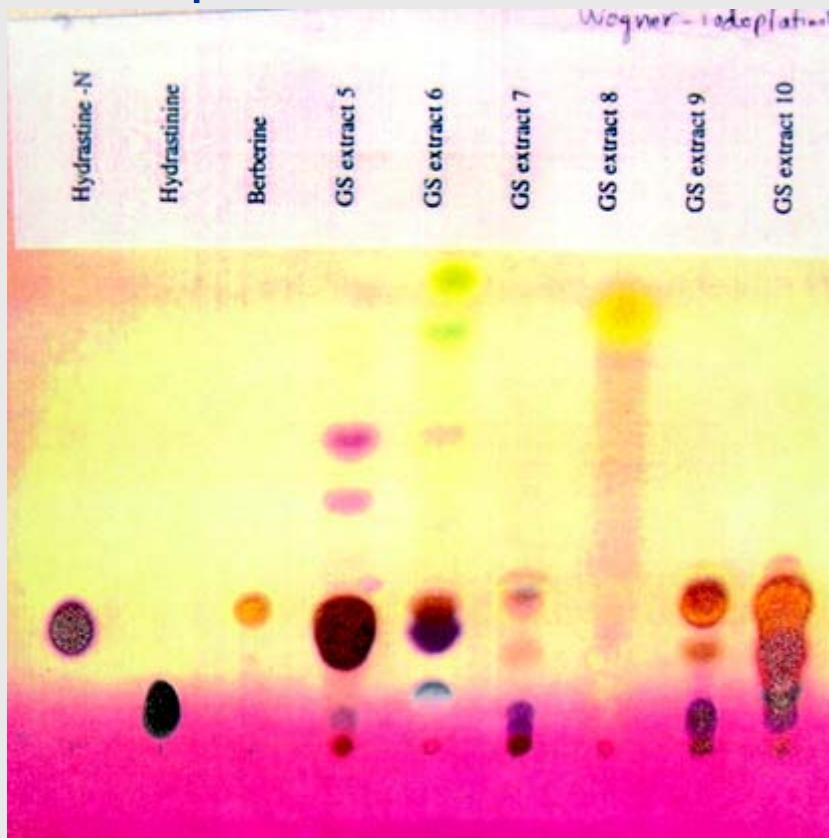


HPTLC

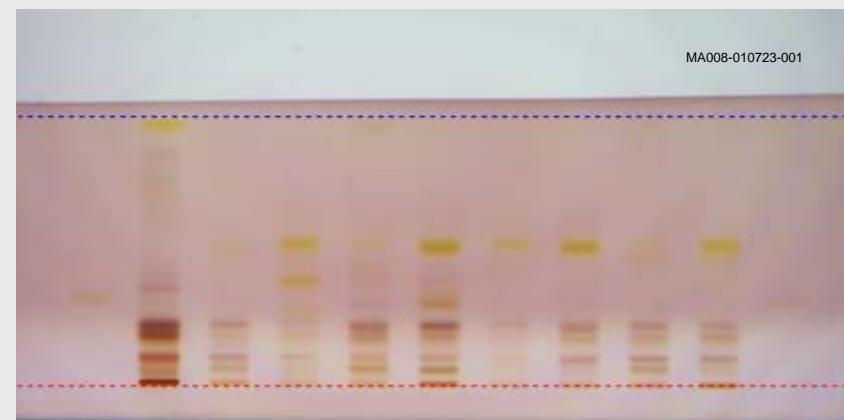


# Spot or band application?

Spot

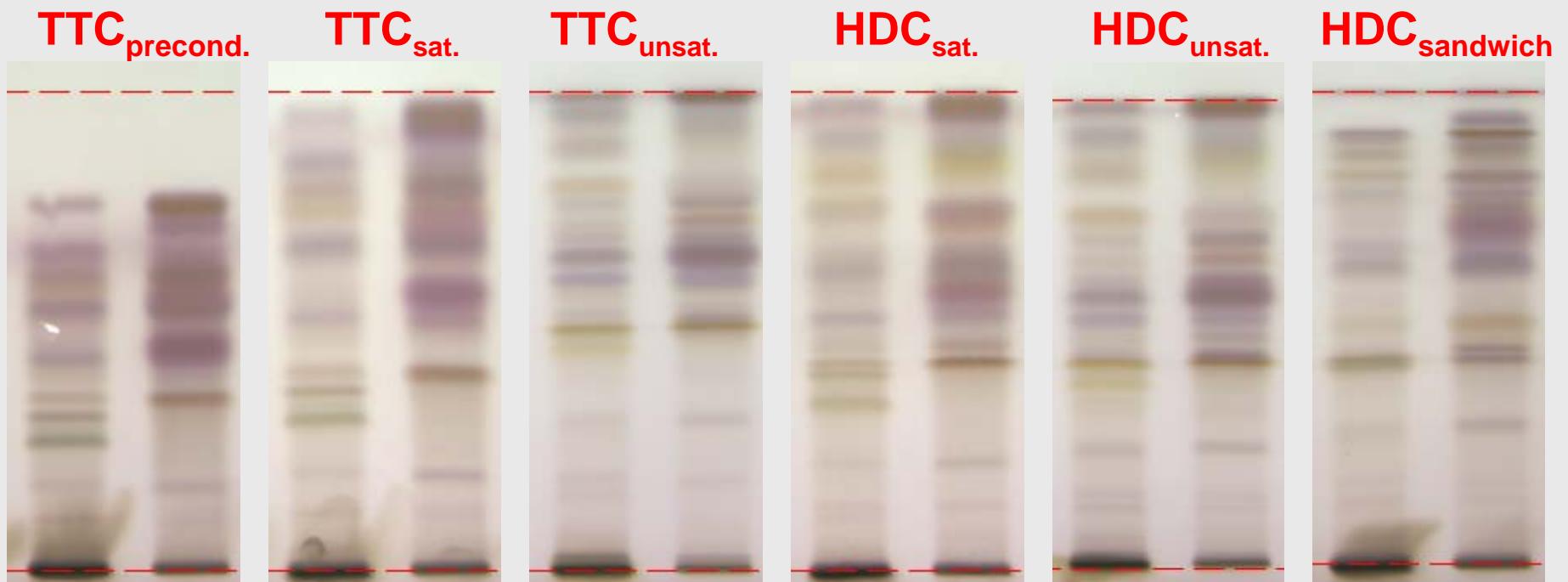


Band



## Effects of chamber configuration/saturation

TTC = Twin Trough Chamber      HDC= Horizontal Developing Chamber

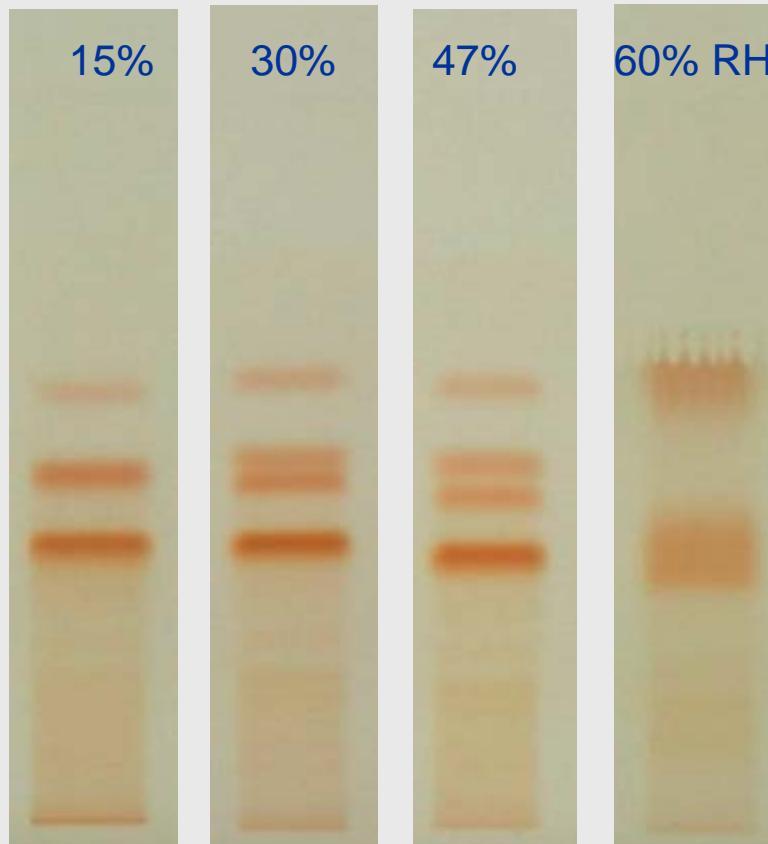


HPTLC silica gel 60 F<sub>254</sub>, toluene - ethyl acetate - acetic acid (70 : 33 : 3)

Left: *Schisandra chinensis*, right: *Schisandra sphenanthera*

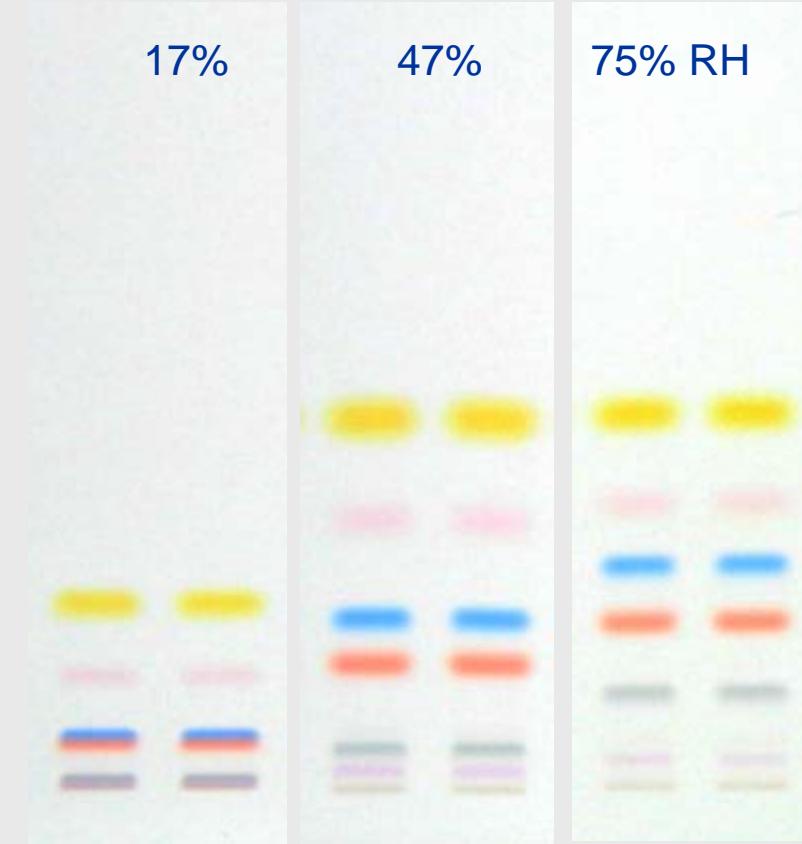
## Effects of rel. humidity /activity

Green Tea: Polyphenols



Toluene, Acetone, Formic Acid (4.5:4.5:1)

Test Dye



Toluene

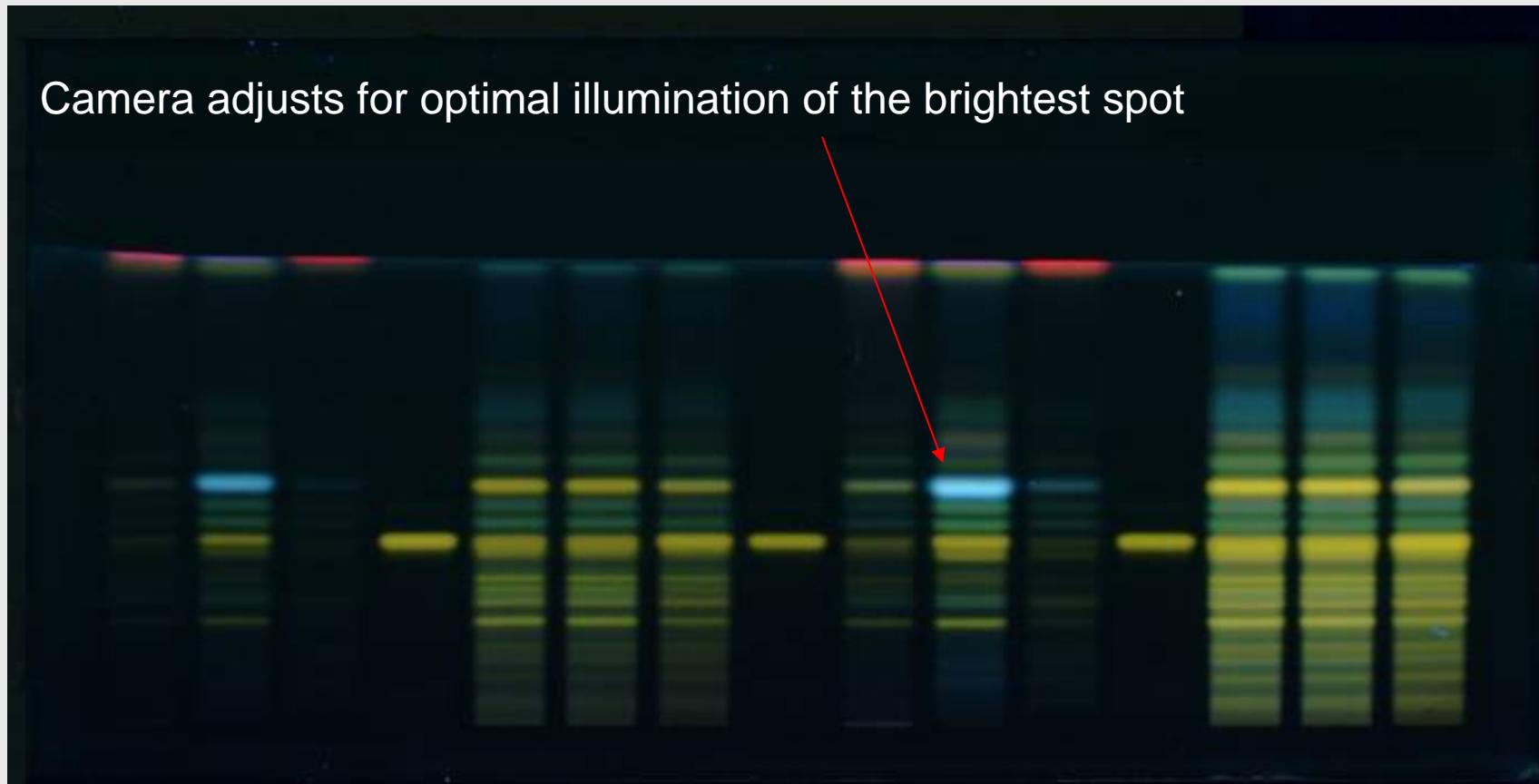
## Spraying or dipping?



Dichloroquinone  
chloroimide reagent  
and ammonia vapor



## Automatic image optimization



## Our approach: SOP for HPTLC

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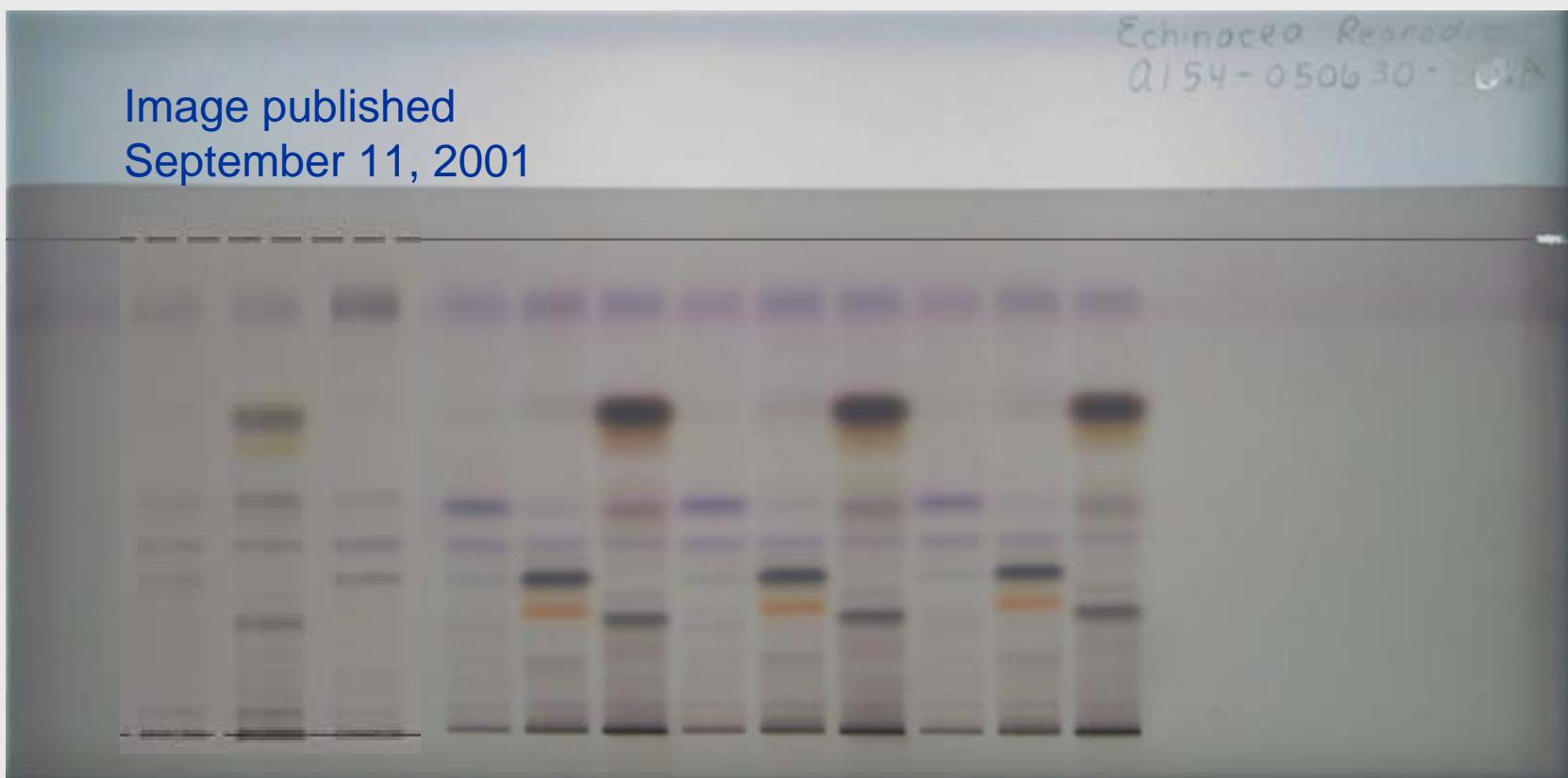
- ▶ Plate material & labeling
  - pre-coated HPTLC plates, 20x10 / 10x10 cm
  - Project number\_year/month/day\_plate number
- ▶ Parameters for sample application
  - 8 mm bands, spray-on, 8 mm (lower edge), 15 mm (left right edge)
- ▶ Detailed description of development
  - 6 cm, 20 min saturation (with filter paper)
- ▶ Derivatization
  - Dipping whenever possible
- ▶ Densitometry
  - MWL scan
  - Scan at the max. WL
- ▶ Digital documentation
  - UV 254 nm / 366 nm / white light (prior to and after derivatization)

## Successful standardization – *Echinacea*

May 06, 2005 – CSI Laboratory

Image published  
September 11, 2001

*Echinacea* Reprodic  
Q154-050630-103A





# Validated Methods

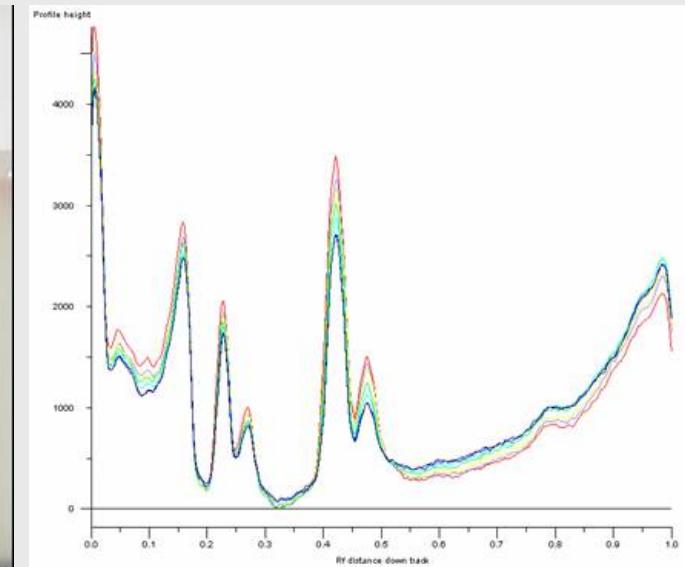
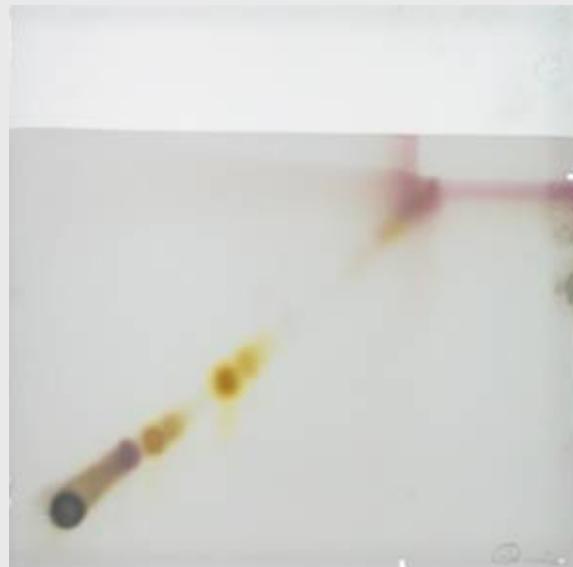
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## Validation of qualitative methods



## Stability: Licorice

- ▶ During chromatography
- ▶ In solution and on plate (3 h each)
- ▶ Result 30 min

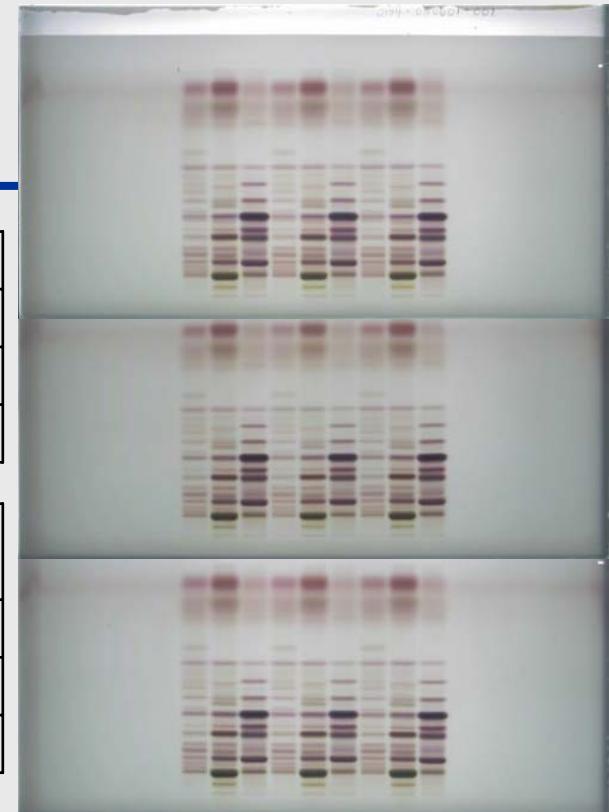


## Precision: Rf of Ginsenosides

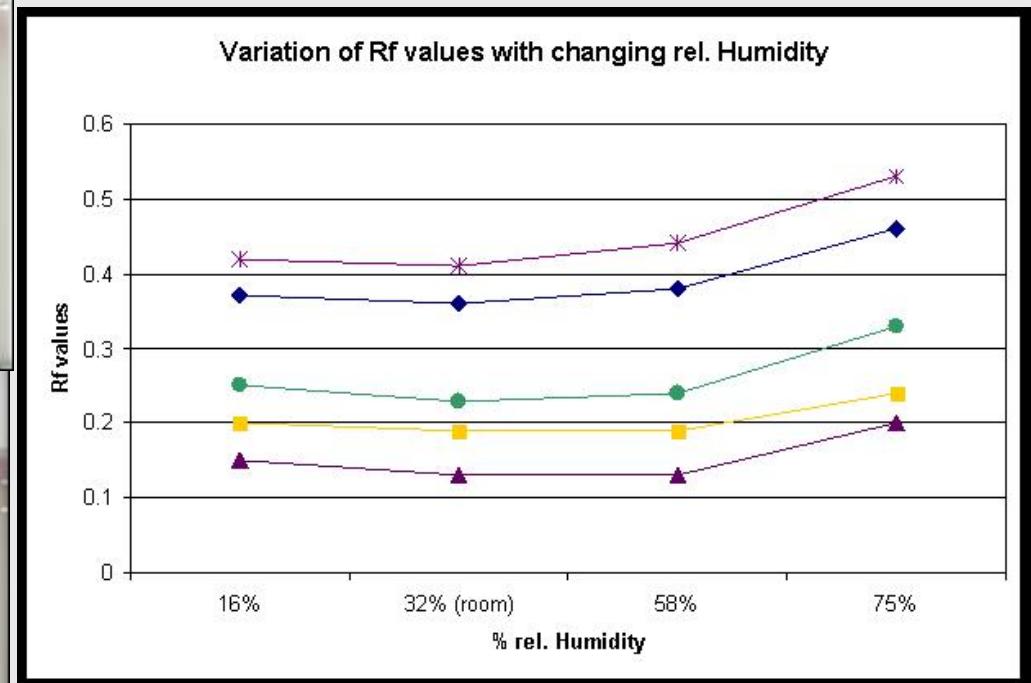
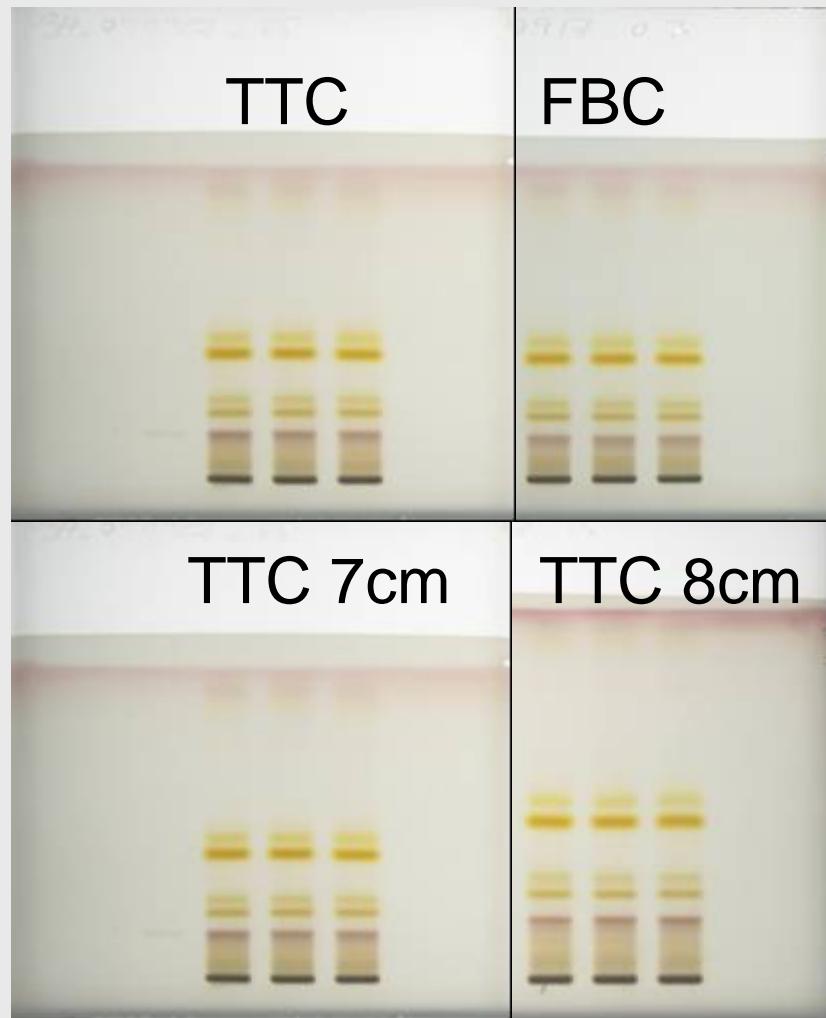
Same day	A154-050531-01	A154-050531-02	A154-050531-03	$\Delta Rf$
Rg1	0.35	0.36	0.36	0.01
Re	0.26	0.26	0.27	0.01
Rb1	0.15	0.16	0.16	0.01

3 days	A154-050531-01 (Comparison)	A154-050601-01	A154-050606-01	$\Delta Rf$
Rg1	0.35	0.35	0.35	0.0
Re	0.26	0.26	0.26	0.0
Rb1	0.15	0.15	0.16	0.01

Two Labs	Plate P47_05072 6_03	Plate P47_05072 6_04	Plate P47_05072 6_05	$\Delta Rf$	A154-050531-01 (Comparison)	$\Delta Rf$
Rg1	0.38	0.38	0.36	0.02	0.35	0.03
Re	0.28	0.29	0.27	0.02	0.26	0.03
Rb1	0.16	0.18	0.16	0.02	0.15	0.03



## Robustness: Licorice



## Validated methods for identification (of single herbs)

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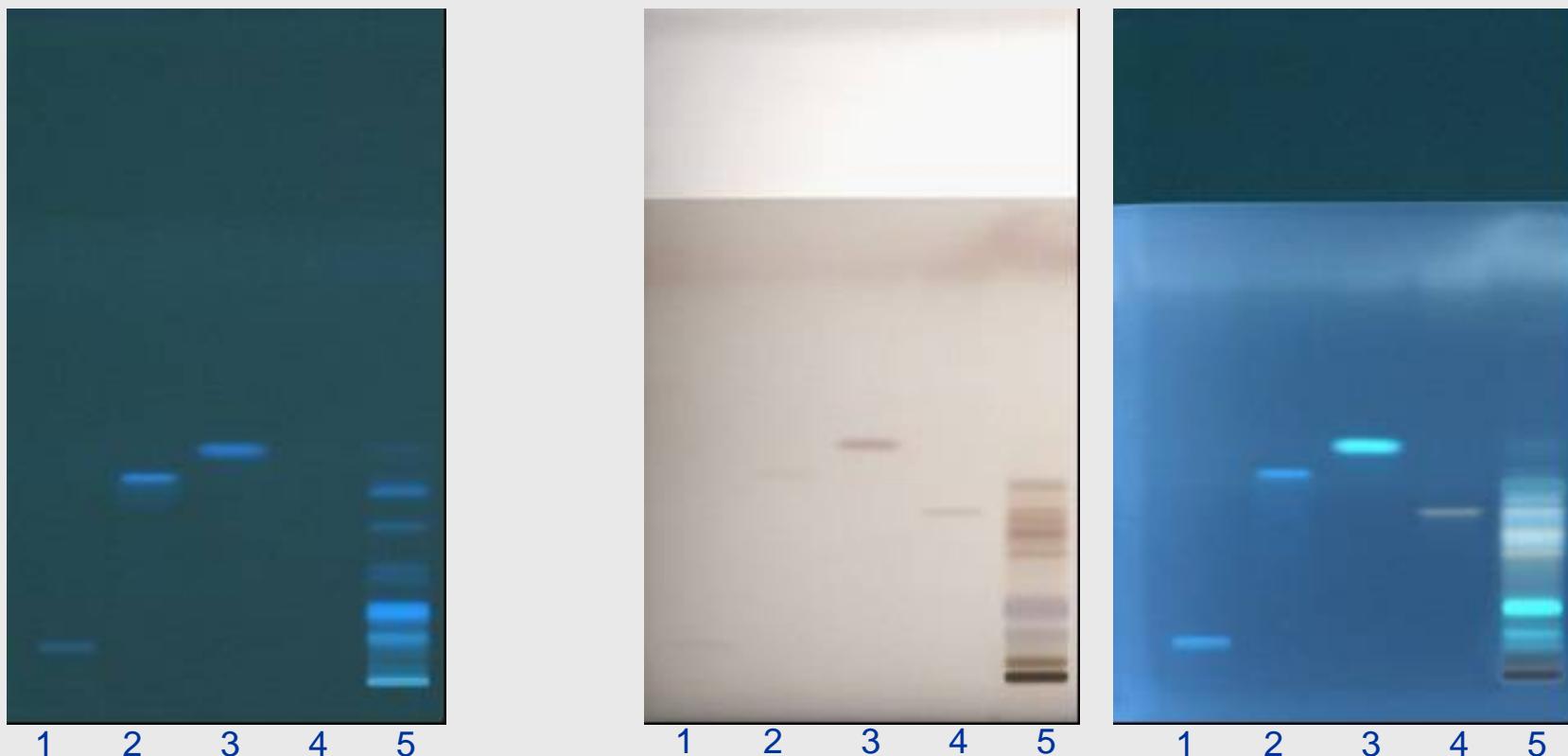
- ▶ Licorice In the process
- ▶ Feverfew
- ▶ Milk thistle
- ▶ Kava kava
- ▶ Ginger
- ▶ Green tea
- ▶ Ginseng
- ▶ Eleuthero
- ▶ Echinacea
- ▶ Black Cohosh
- ▶ St.Johns Wort
- ▶ Saw Palmetto
- ▶ Hoodia
- ▶ Skullcap
- ▶ Star Anise
- ▶ Other Cimicifuga species



# Identification of single species

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## Identification of Black Cohosh (validated, AHP)



1 Chlorogenic acid; 2 Caffeic acid; 3 Isoferulic acid; 4 Actein; 5 *Actaea racemosa* BRM

## Variability?



- 1: Chlorogenic acid; 2: Caffeic acid; 3: Isoferulic acid; 4, 16: Actein  
5: *Actaea racemosa* BRM; 6: *Actaea racemosa* ; 7: *Actaea* (wild);  
8: *Actaea* (cultivated); 9: *Actaea* (cultivated)\*; 10: *Actaea* (cultivated);  
11: *Actaea* (wild); 12: *Actaea rhizoma* conc.; 13: *Actaea racemosa* extract;  
14: *Actaea racemosa* powder; 15: *Actaea racemosa* powder;

## Other species



- 1: Chlorogenic acid, actein, caffeic acid, and isoferulic acid (increasing Rf values);  
2: *Actaea racemosa* BRM;      3-6: *Actaea podocarpa*;      7-8: *Actaea pachypoda*;  
9: *Actaea pachypoda* (leaves);      10: *Actaea rubra*;      11: *Actaea foetida*  
12-13: *Actaea dahurica*;      14: *Actaea heracleifolia*;      15-16: *Actaea* sp unknown  
17: *Caulophyllum thalictroides*      18-19: Guang Shengma (no *Actaea*).



# Identification of mixtures

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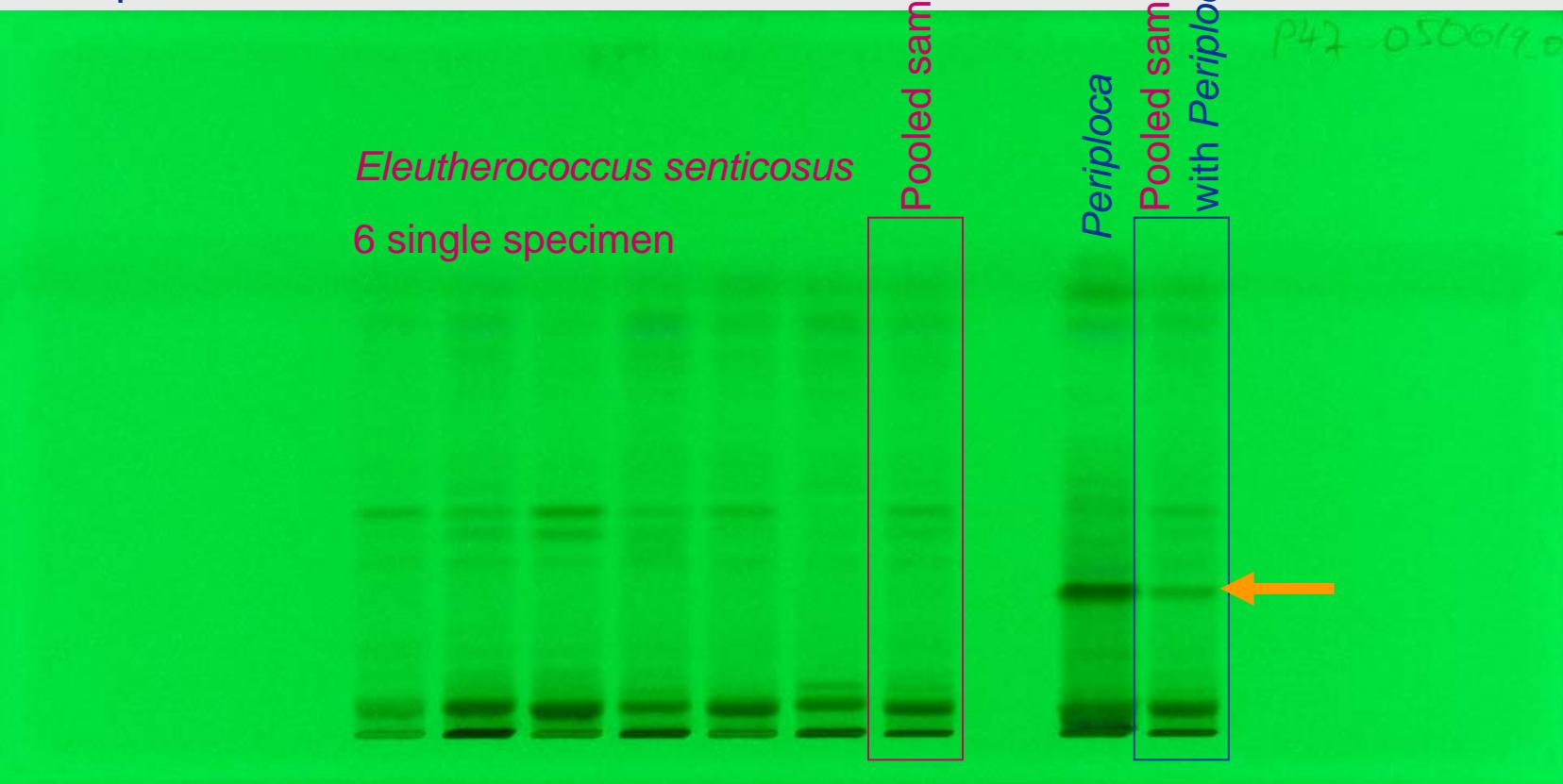
## Detection of adulterated mixtures

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- ▶ Great challenge
- ▶ (Not just) identification of markers
- ▶ Representative „pooled samples“
- ▶ Representative fingerprint for adulterant
- ▶ Improved specificity
- ▶ Detection limits

## Single specimen vs. pooled sample

Multiple detection 1: UV 254 nm



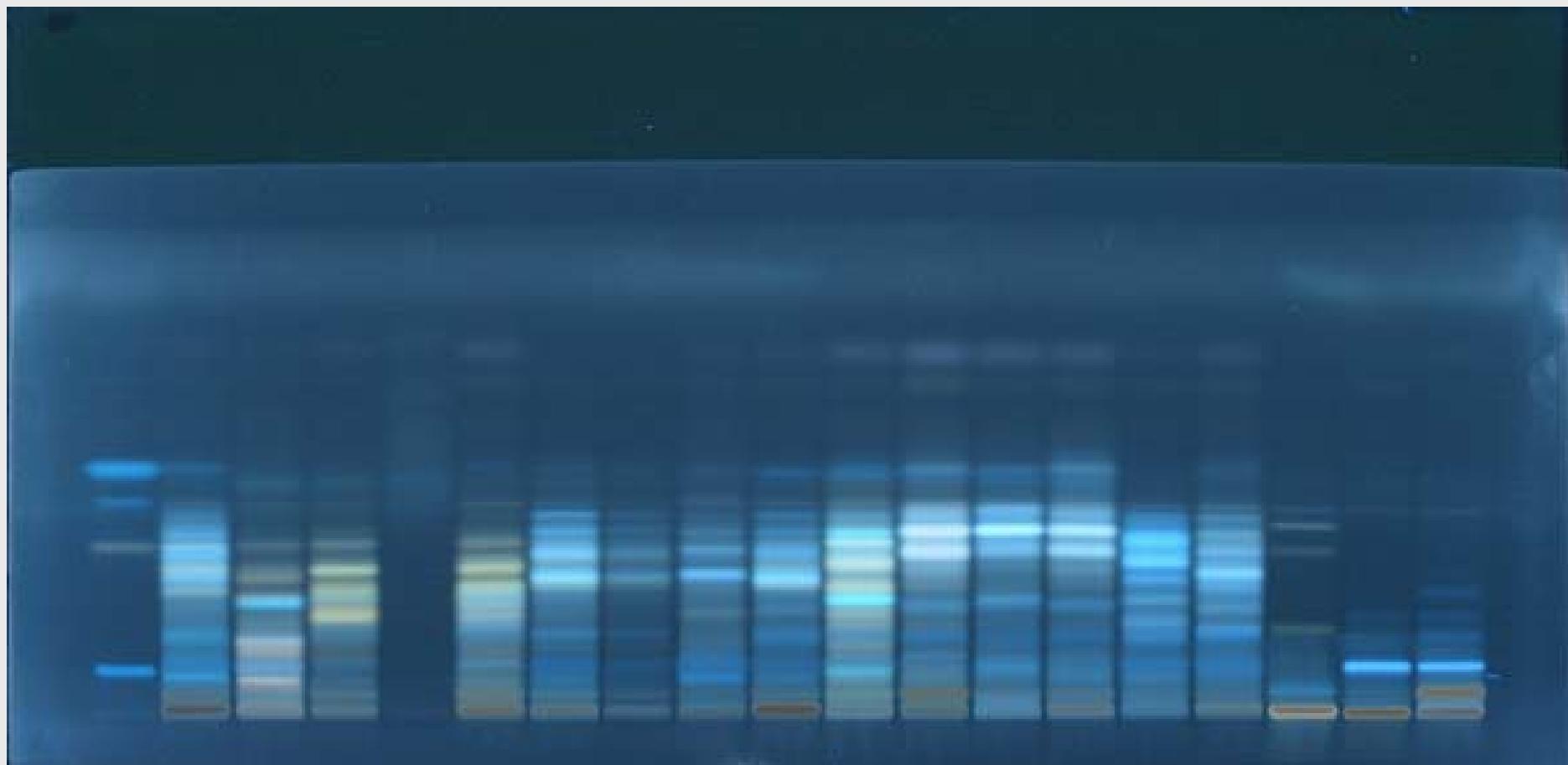
## Single specimen vs. pooled sample

Multiple detection 2: Sulfuric acid reagent



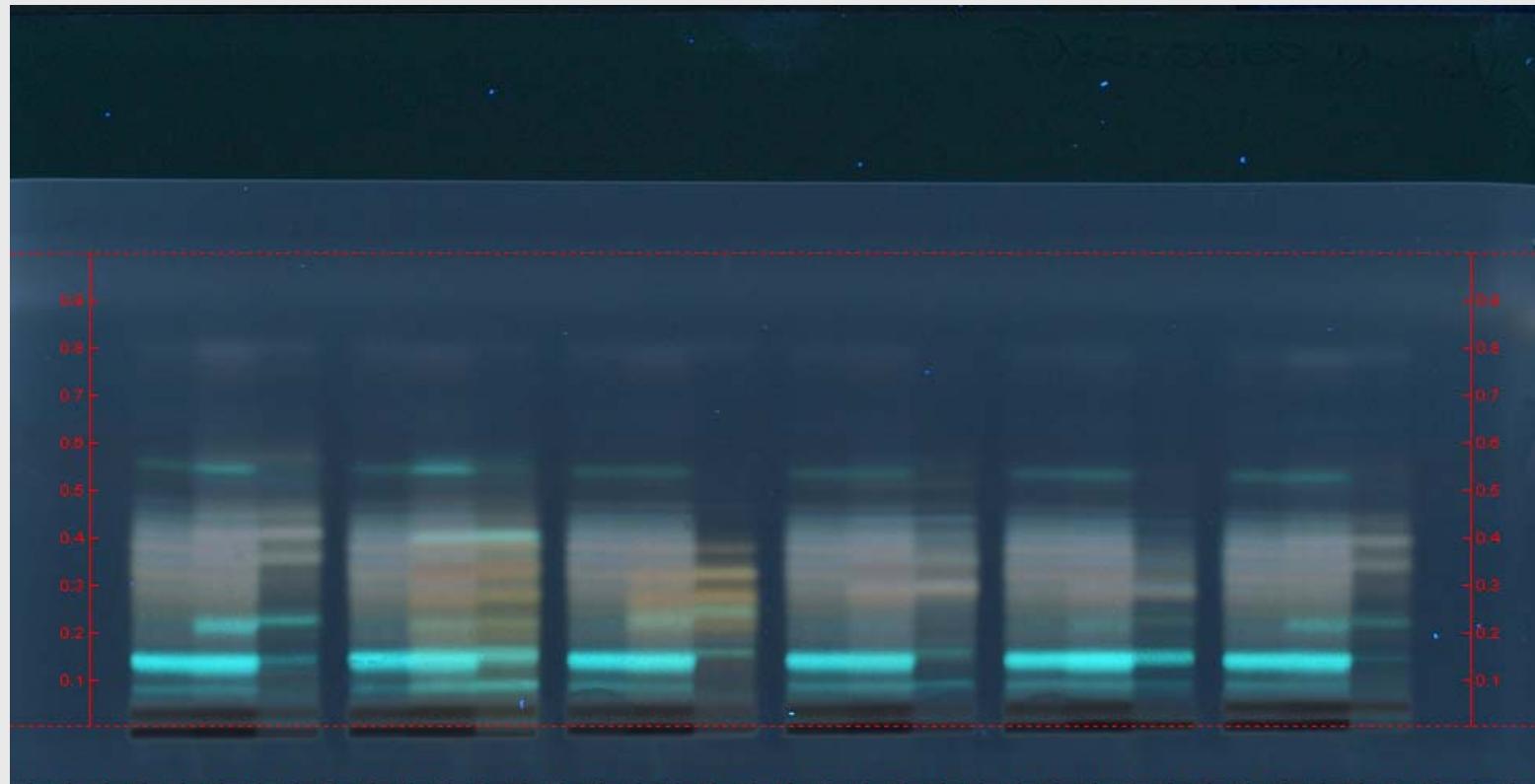
## Again: Black Cohosh and adulterants

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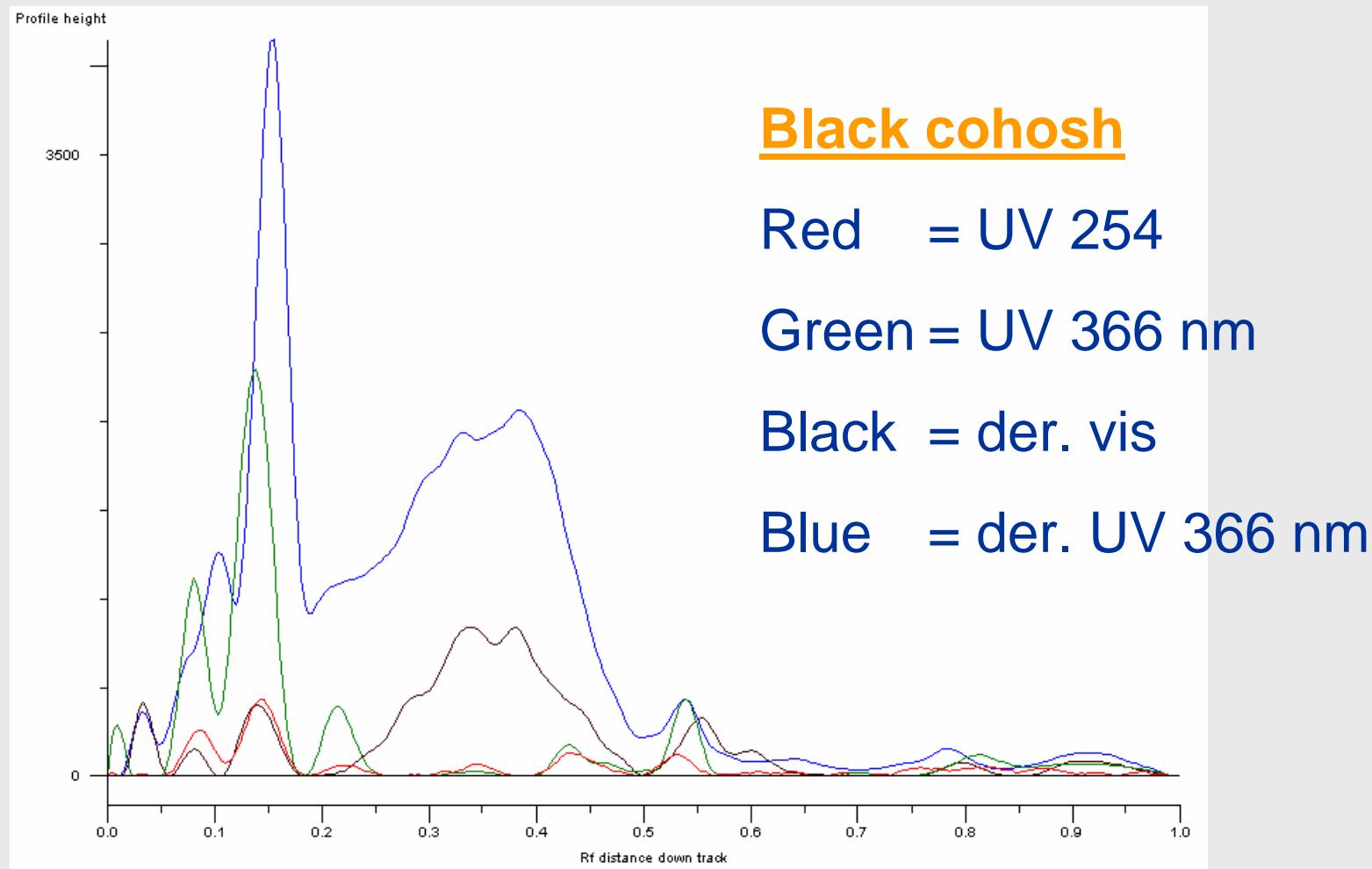
## Adulteration of Black Cohosh

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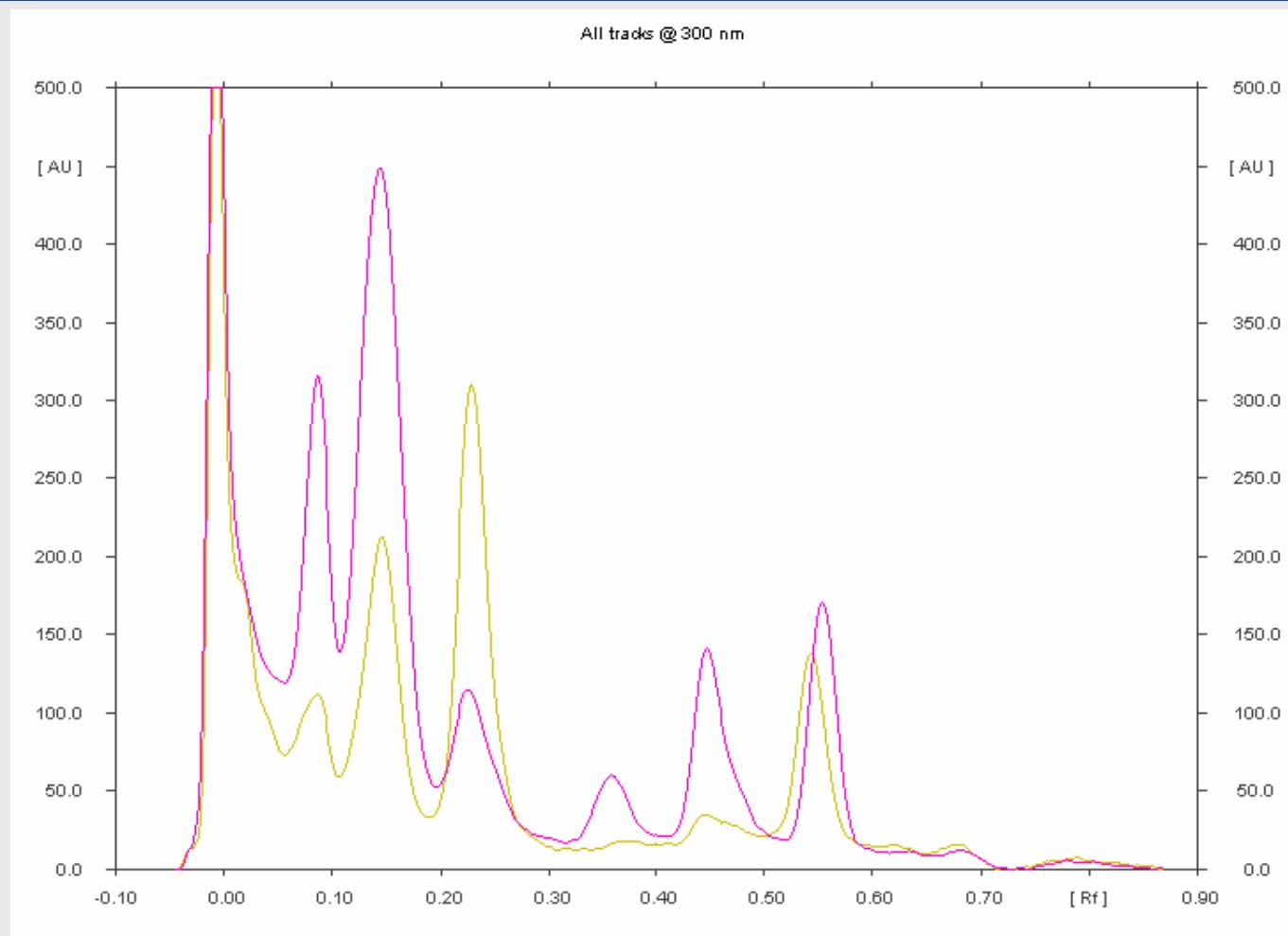


*Actaea hera.*    *foet.*    *podo.*    *pach.*    *rubr.*    *dahu.*

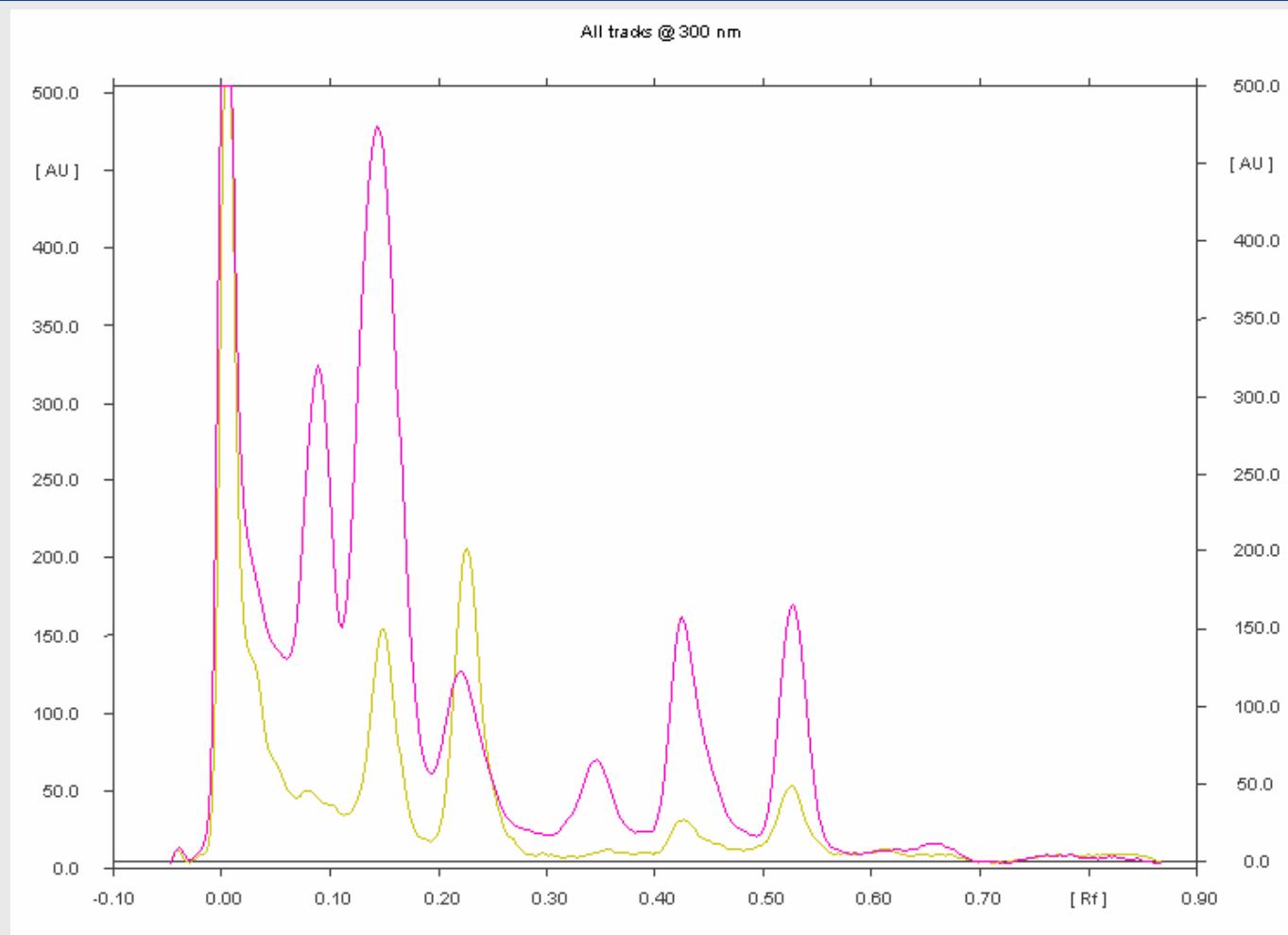
## Profile comparison (multiple detection)



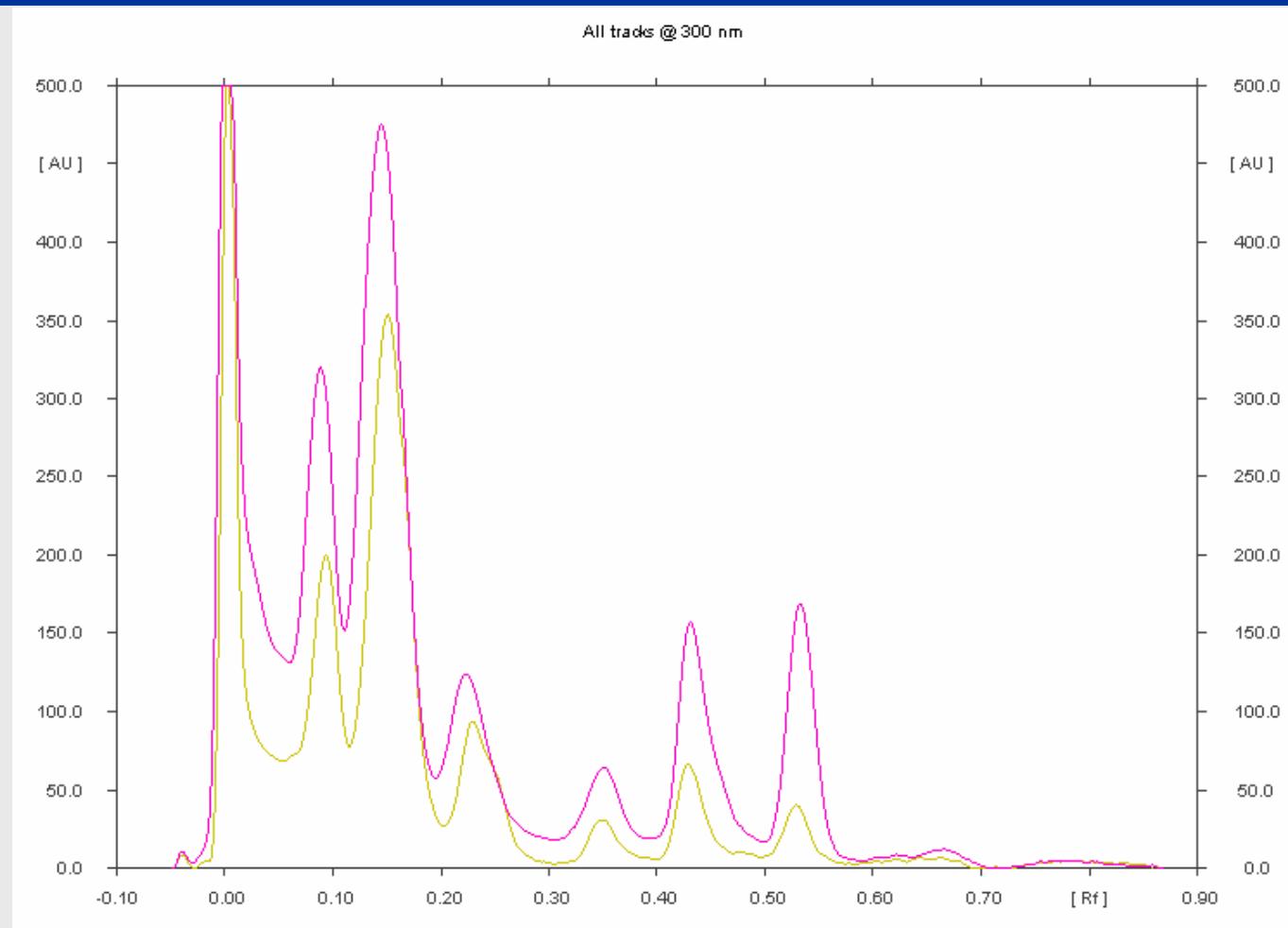
## *A. racemosa* vs *A. heracleifolia*



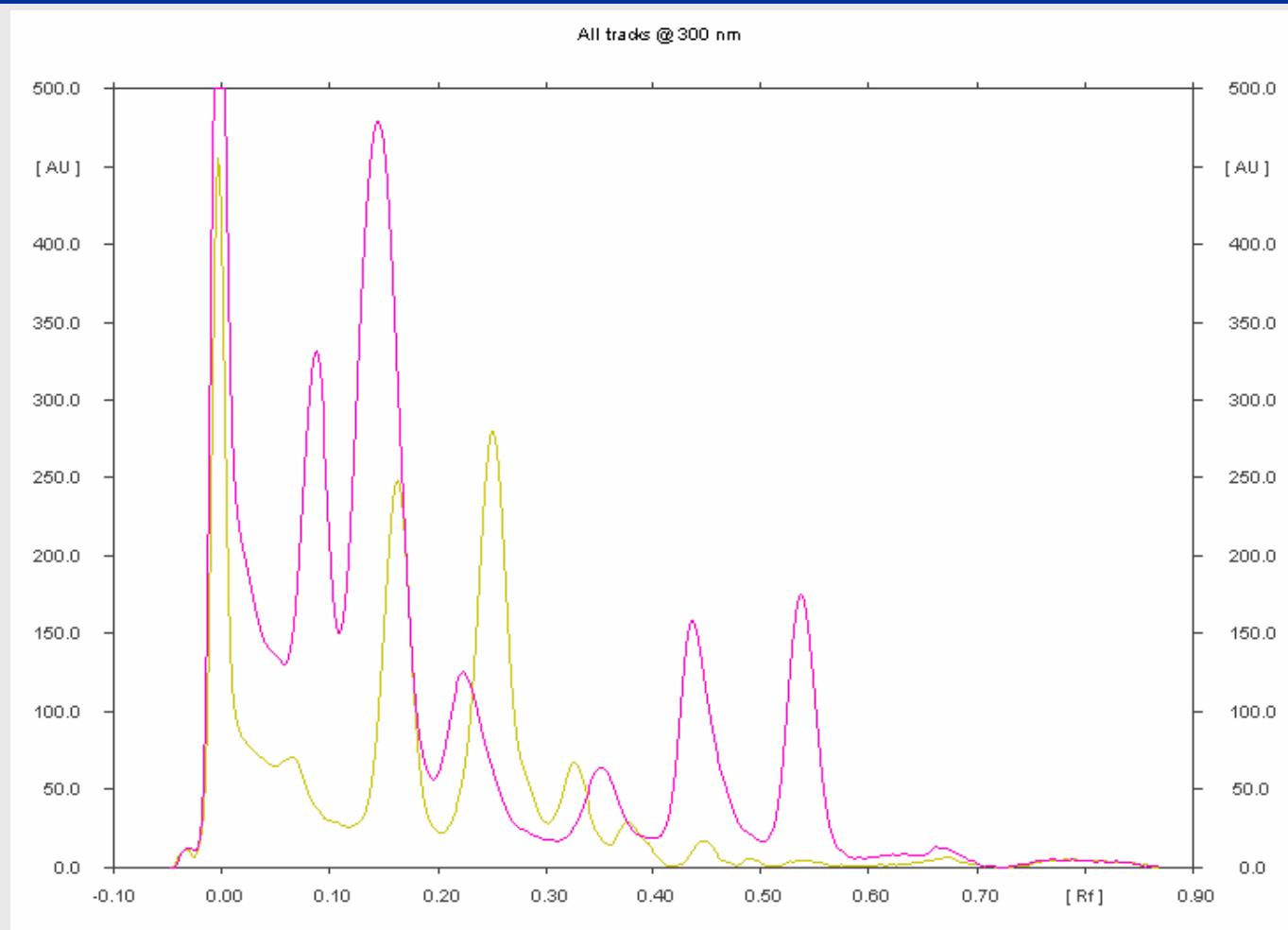
## *A. racemosa* vs *A. dahurica*



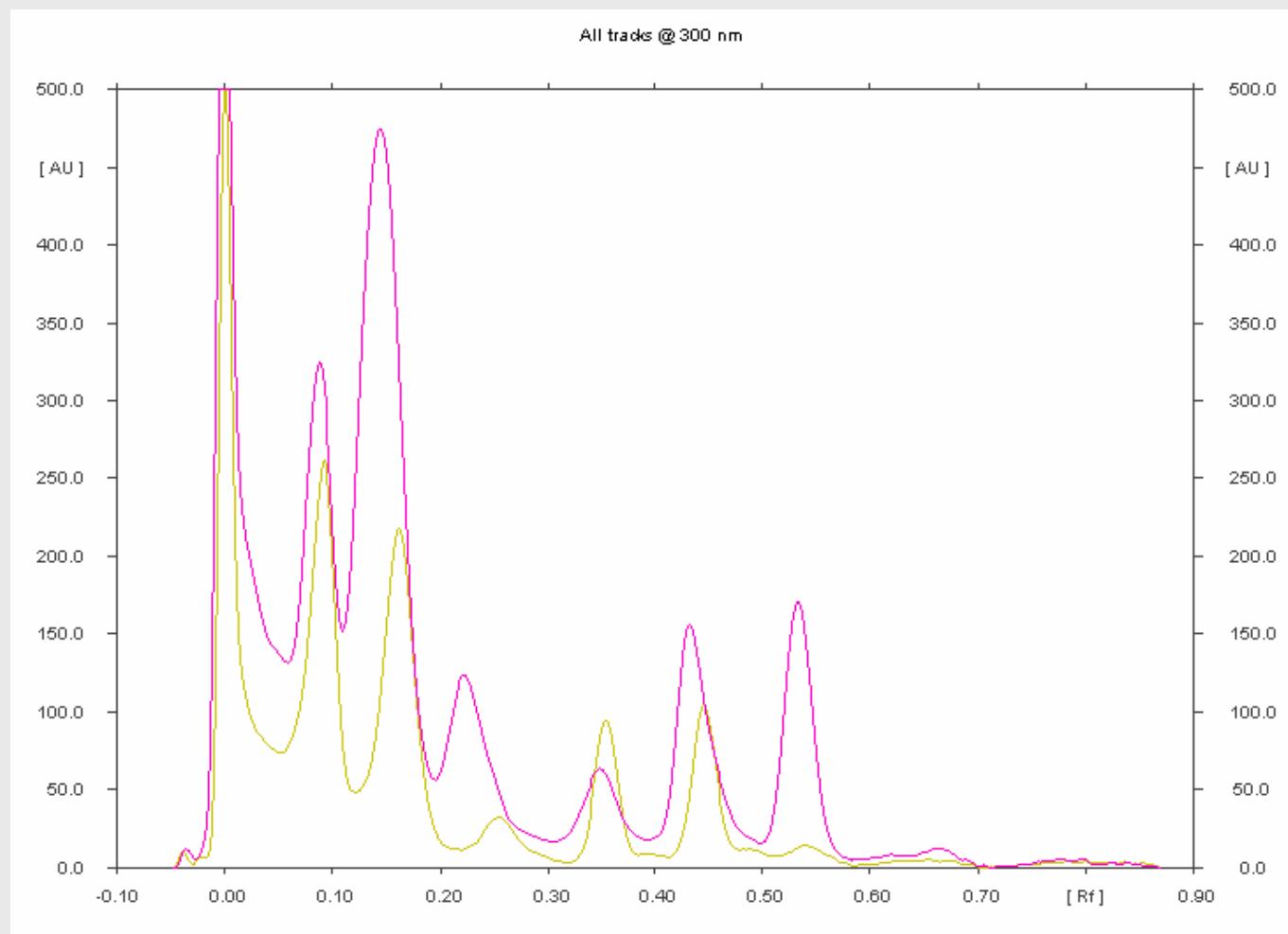
## *A. racemosa* vs *A. rubra*



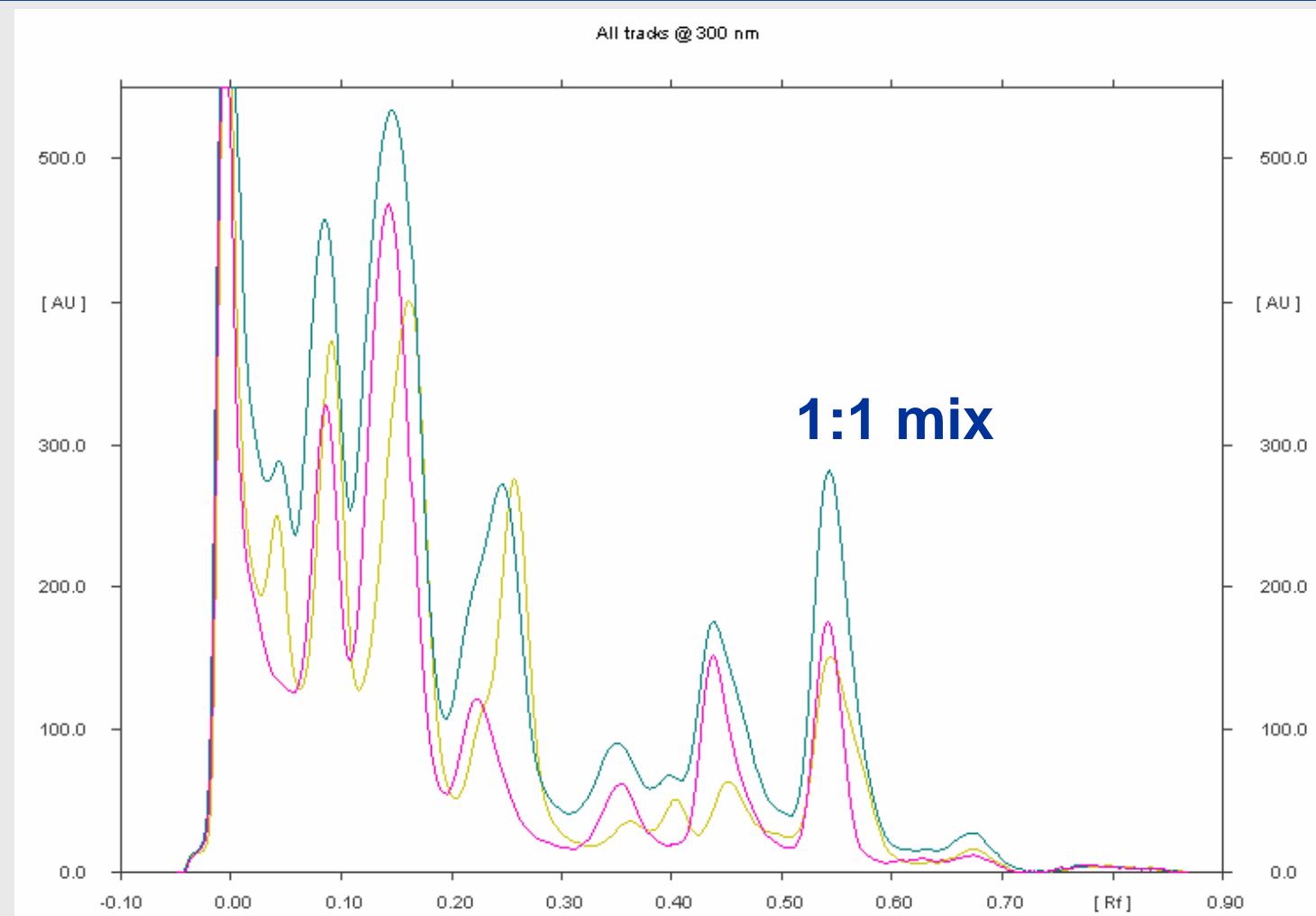
## *A. racemosa* vs *A. podocarpa*



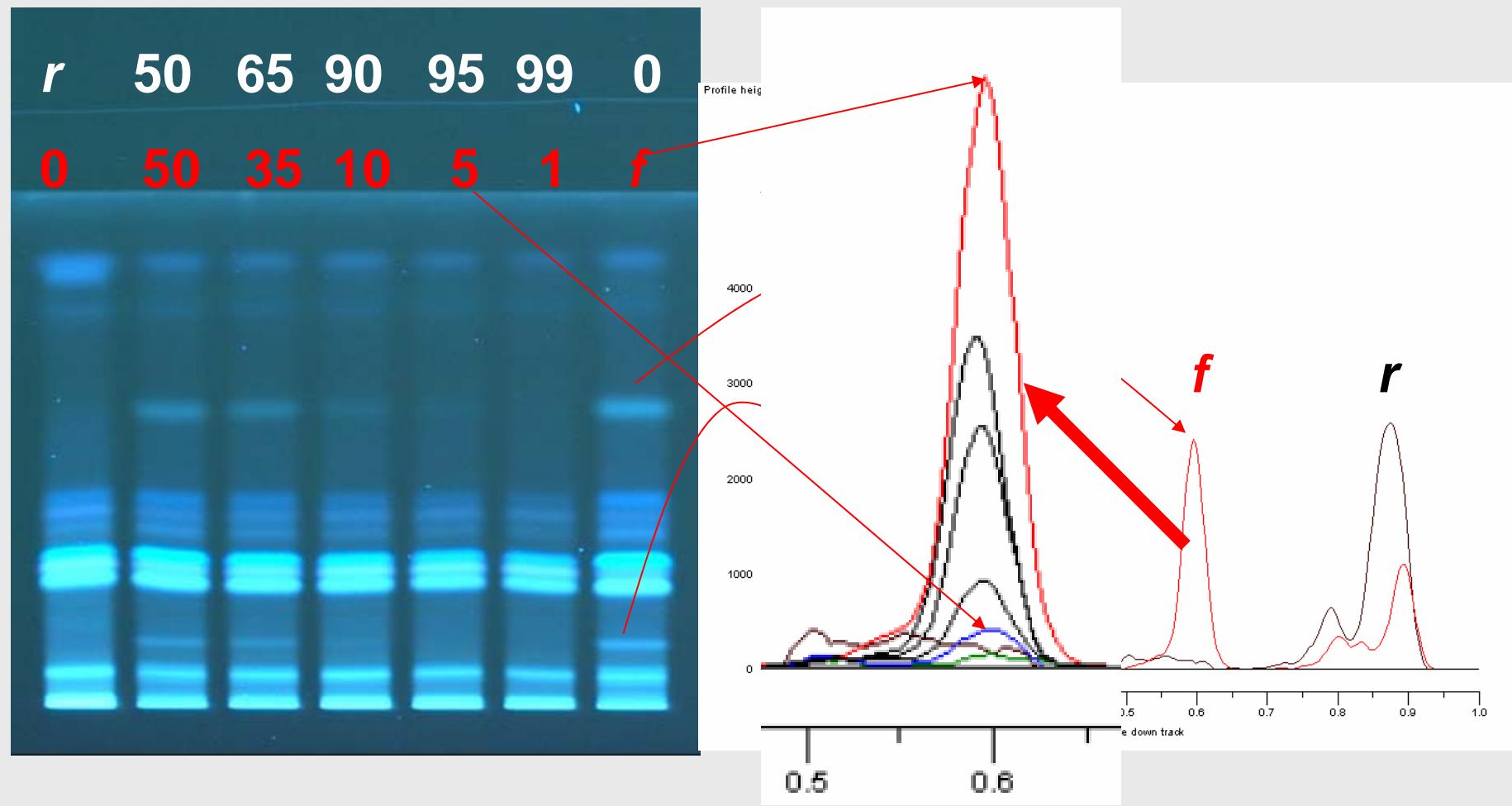
## *A. racemosa* vs *A. pachypoda*



## *A. racemosa* vs *A. foetida*



## *A. racemosa – A. foetida*

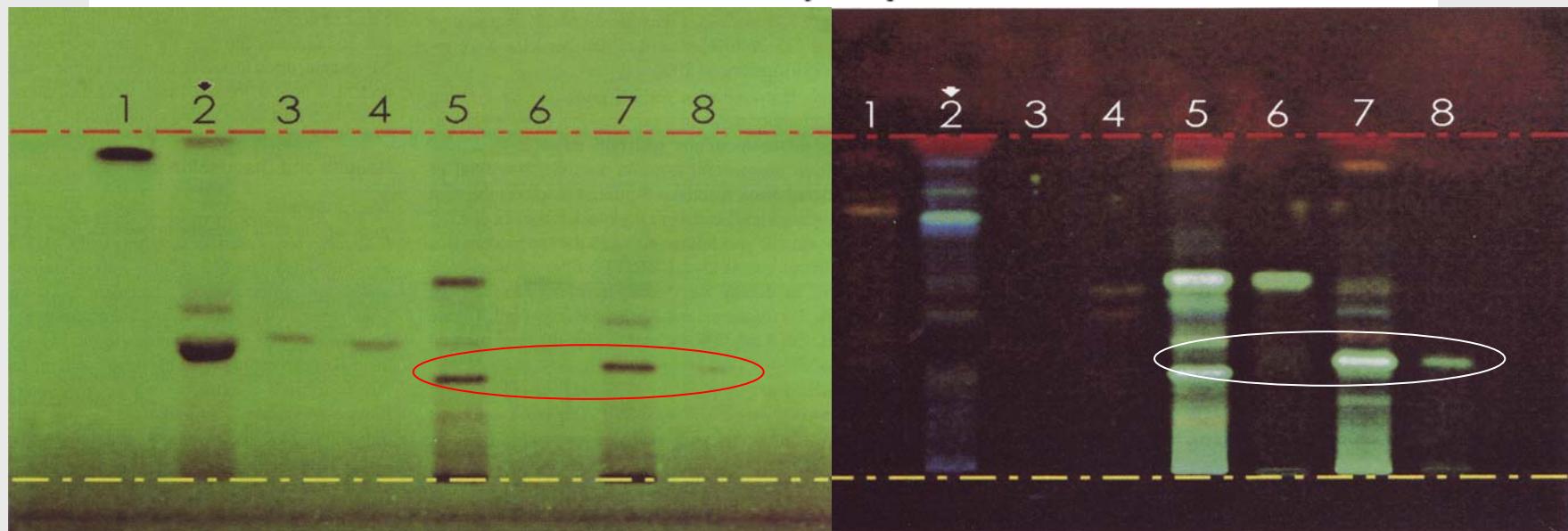


# Identification of Skullcap

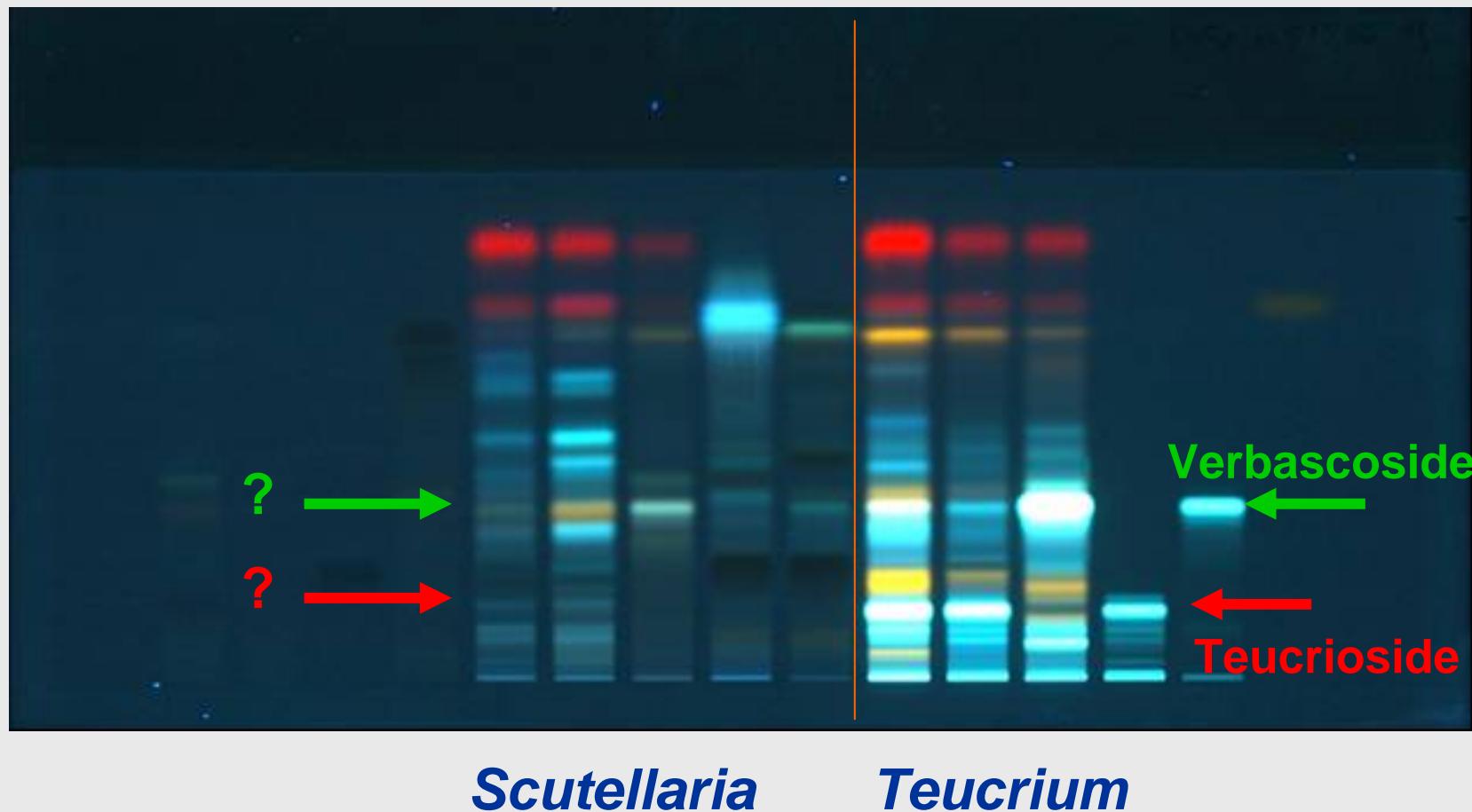
GAFNER ET AL.: JOURNAL OF AOAC INTERNATIONAL VOL. 86, No. 3, 2003 453

## DIETARY SUPPLEMENTS

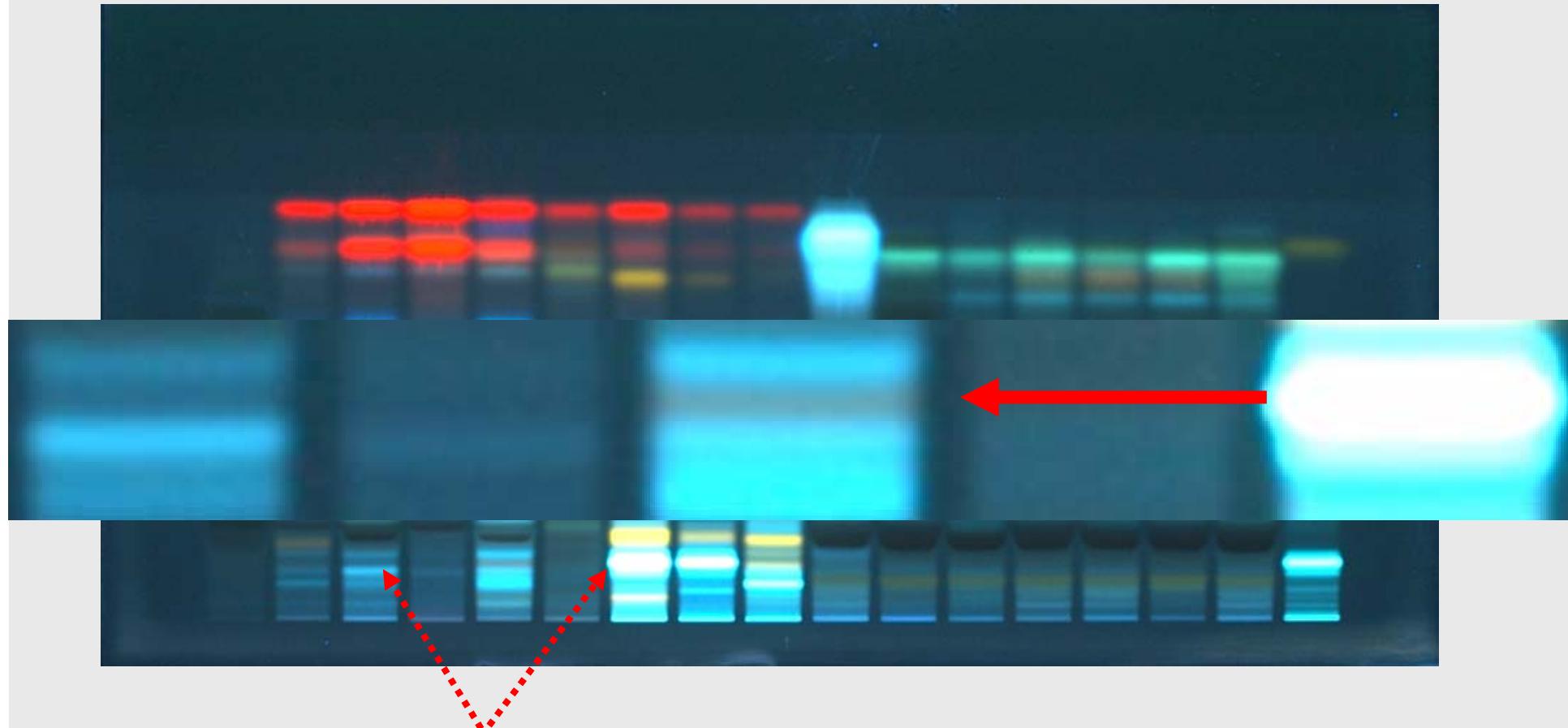
### Analysis of *Scutellaria lateriflora* and Its Adulterants *Teucrium canadense* and *Teucrium chamaedrys* by LC–UV/MS, TLC, and



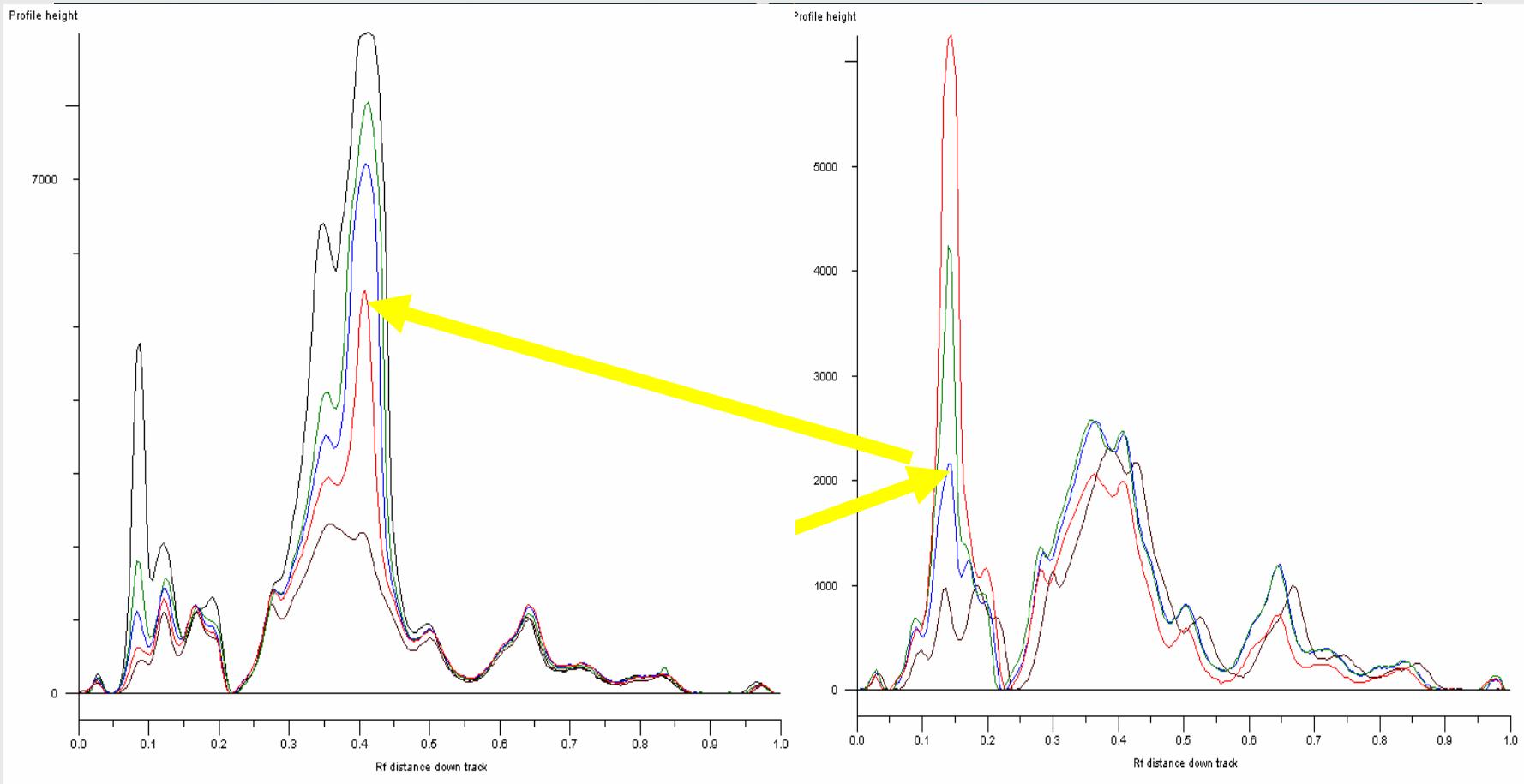
## Identification of Skullcap (Gaffner et.al.)



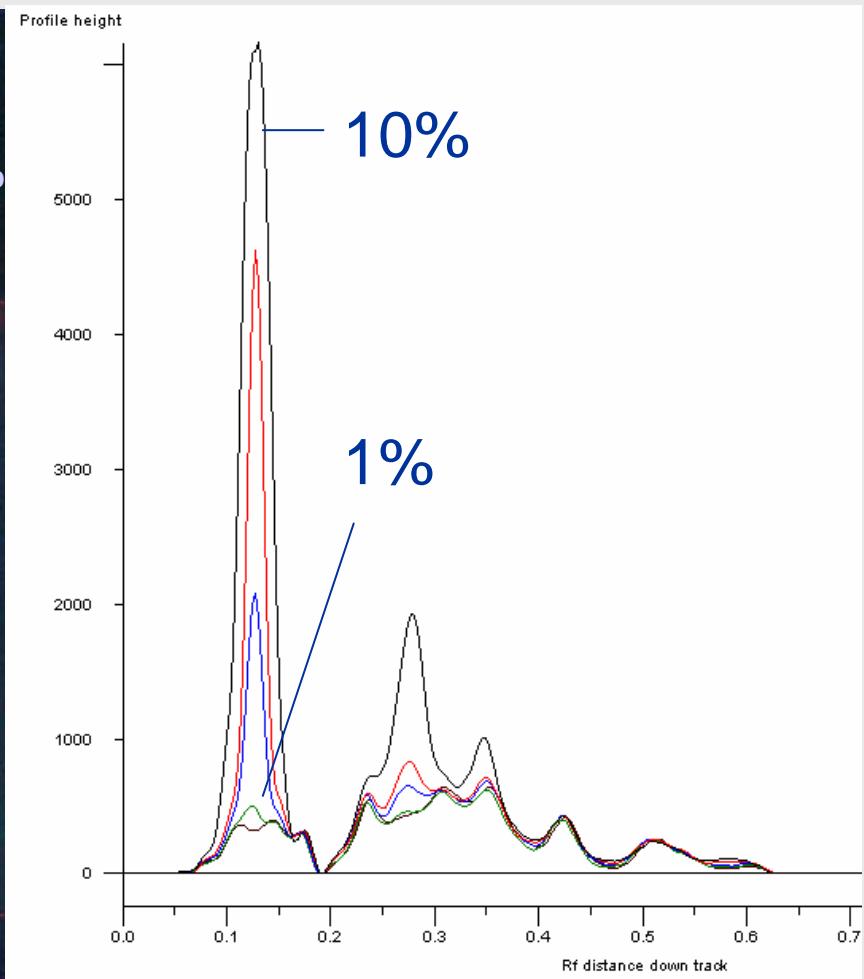
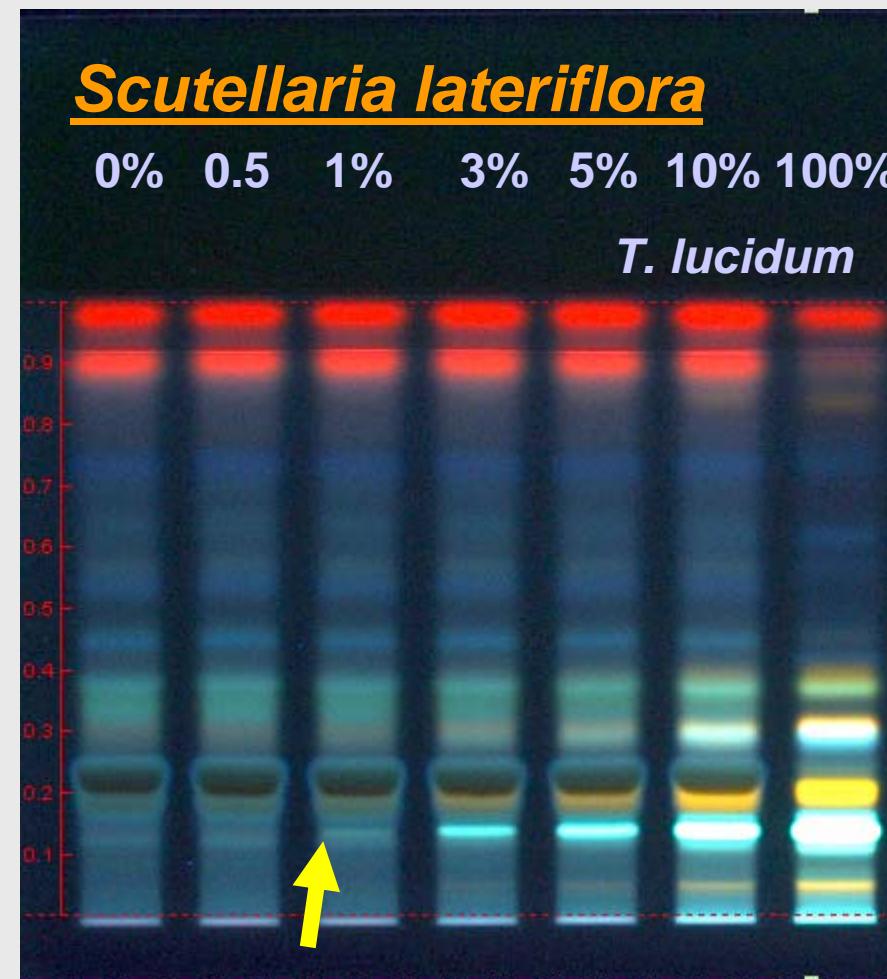
## Identification of Skullcap (AHP method)



## What about mixtures?



## What about mixtures?



## Summary

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- ▶ Standardization and validation are prerequisites for reliable results.
- ▶ Validated HPTLC methods are complementary to more sophisticated techniques.
- ▶ They allow identification of pure samples and mixtures with certainty.
- ▶ HPTLC is a very flexible and inexpensive analytical tool.

## Questions?

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- ▶ [www.camagusa.com](http://www.camagusa.com)