

Histamine in Fish and Theanine in Tea – Two Cases for Employing Planar Chromatography

Prof. Dr. Karl Speer

S. Kretzschmar, H. Dunger, I. Kölling-Speer

Technische Universität Dresden
Institut für Lebensmittelchemie
Bergstr. 66, D-01062 Dresden

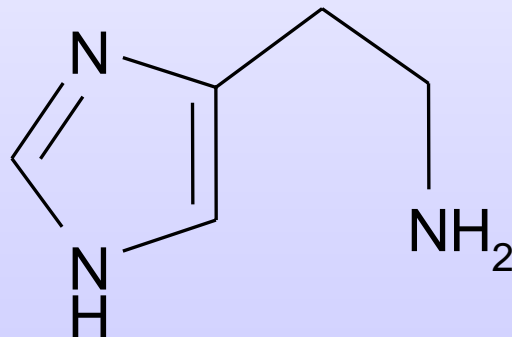


Biogenic amines



Why fish is a problem

- High water content and soft protein
- Enzymes show high activity at low temperatures
- Some fish species obtain high contents of free amino acids



Histamine

Histamine toxication

- Short incubation time
- Decrease of the blood pressure
- Dizziness
- Gastric spasms and headache
- Nausea and vomiting

Histamine in food

< 50 ppm are safe

50 - 200 ppm may be toxic

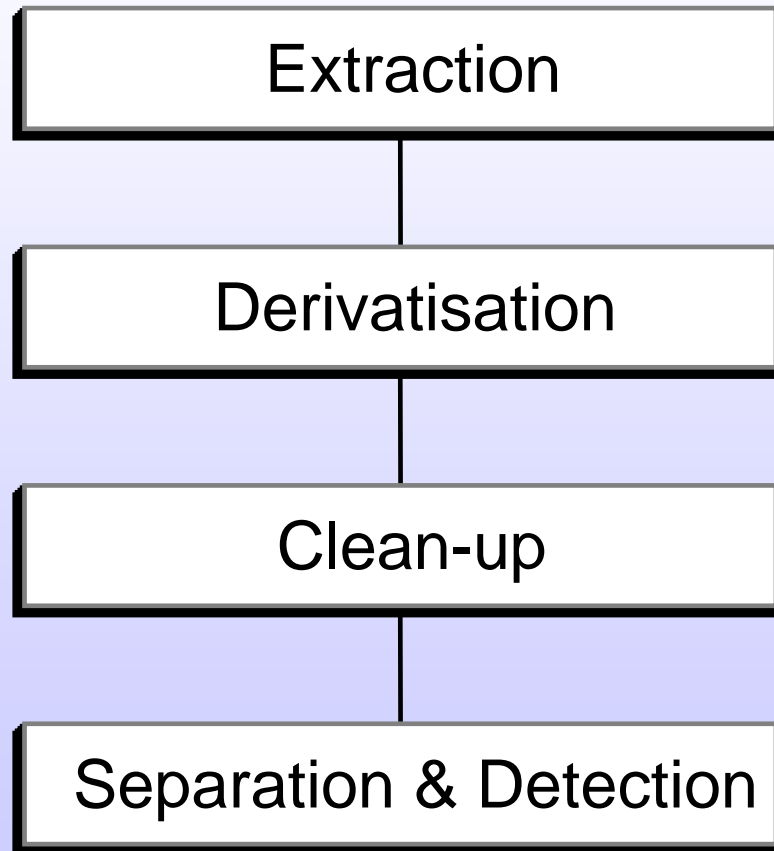
200 – 500 ppm are toxic

500 – 1000 ppm are toxic and dangerous

Legal regulations in Germany

- Paragraph 16 FischhygieneVO and guideline 95/71/EU
- Scombridae, Clupeidae, Engraulidae, Coryphaenidae and Istiophoridae
> 200 mg Histamine/kg » Marketing ban
- Engraulidae in salt brine
> 400 mg Histamine/kg » Marketing ban
- Appendix 3 of Fischhygiene-VO
with guideline 91/493/EU

Analysis of biogenic amines



Development of a TLC - screening-method



CAMAG Automatic TLC Sampler III



CAMAG TLC Scanner II

Extraction

10 g of the homogenized sample
+ 80 ml TCA (10%)
Ultra-Turrax 90 s
adjust to 100 ml with TCA and filter

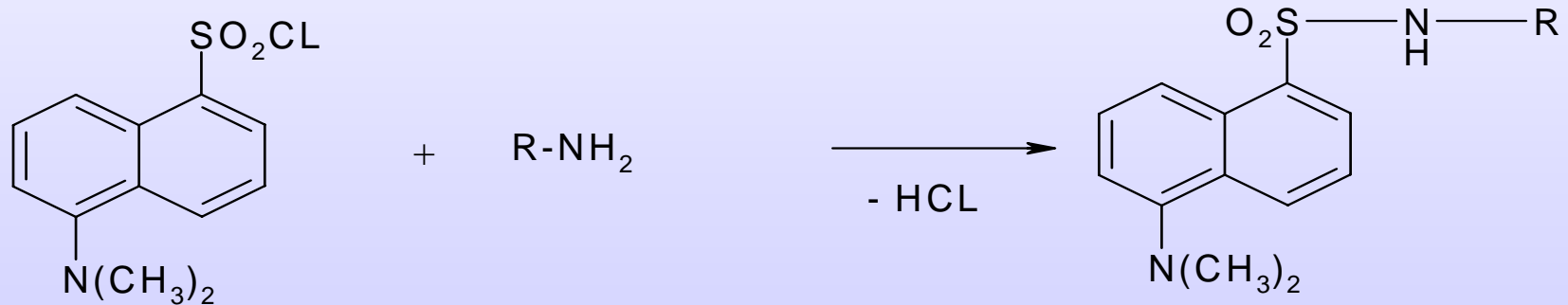
Derivatisation

1 ml of the filtrate is adjusted to pH 8

Add 1 ml borate-puffer and 2 ml dansyl chloride solution

Incubate for 1 h

Adjust to 10 ml with water



Clean-up

Add 5 ml of diethyl ether

Shake vigorously and centrifuge

Remove the organic layer and shake the water residue twice more

Reduce the combined extracts to dryness and dissolve the residue in
5 ml acetonitrile

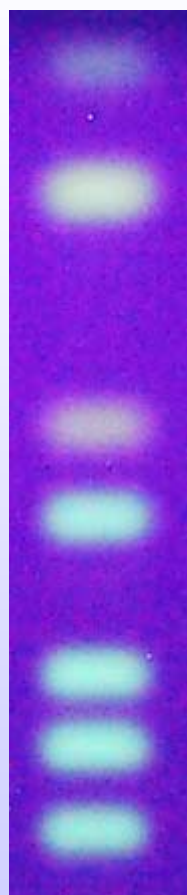
Separation

TLC plates silica gel 60 (Merck) (20 x 10 cm, layer thick-ness 0.25 mm,
pre-washed with developing solvent)



Separation was performed in a horizontal
development chamber

Densitogram of a standard solution (365/>400 nm)



PHE

TYR

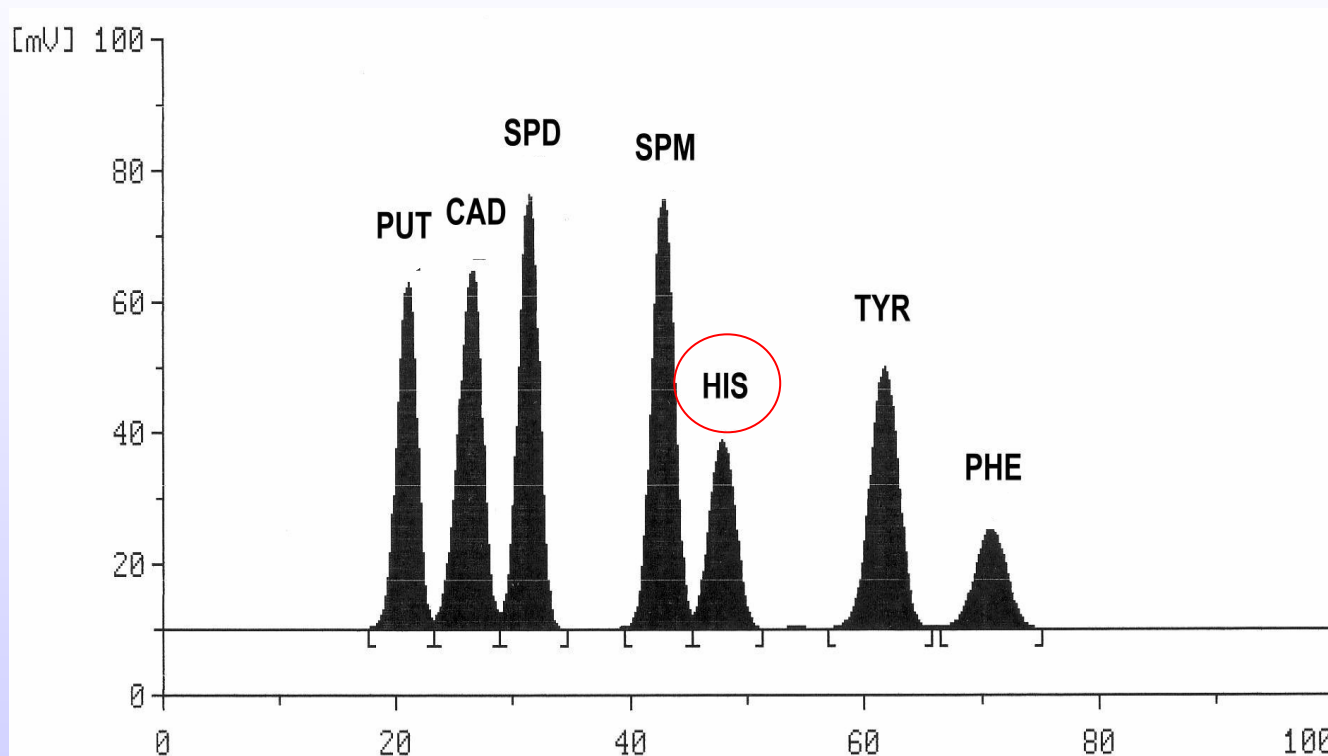
HIS

SPM

SPD

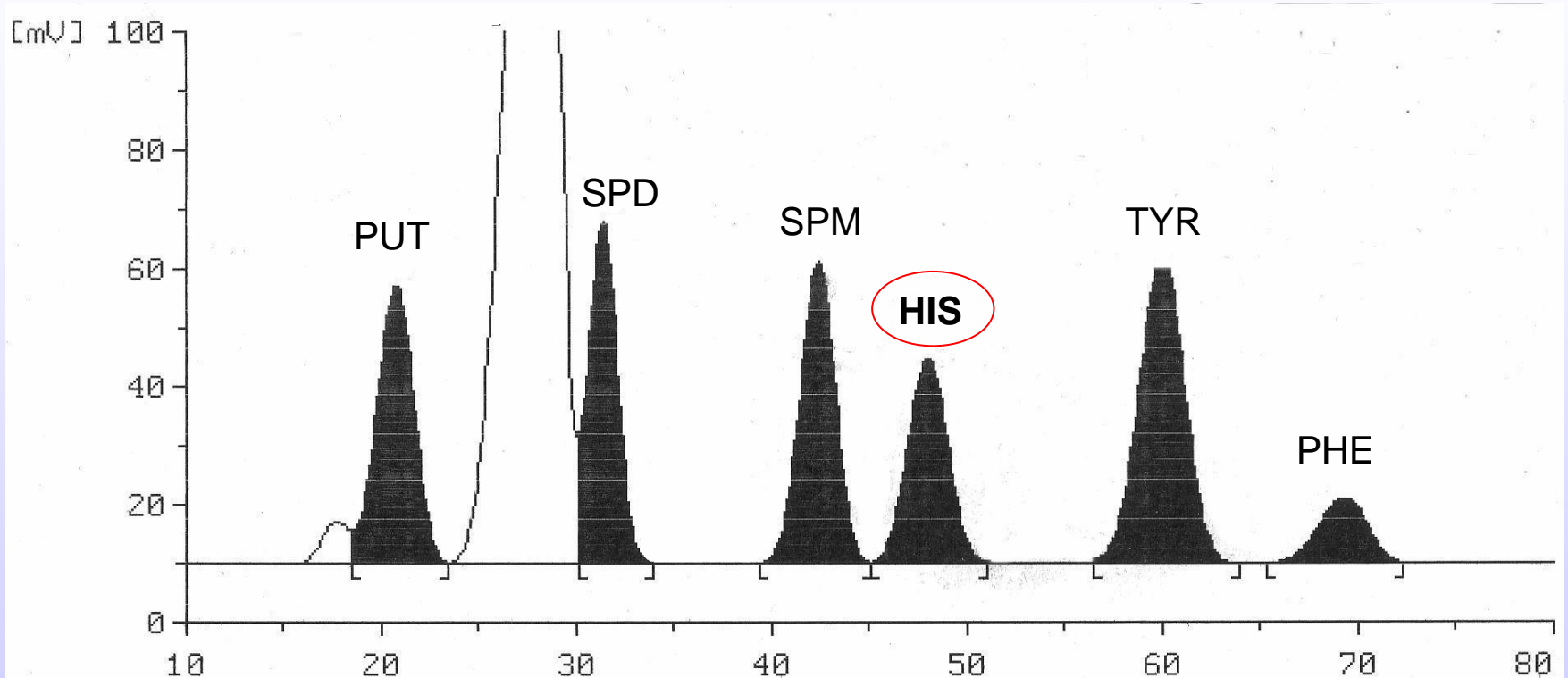
CAD

PUT



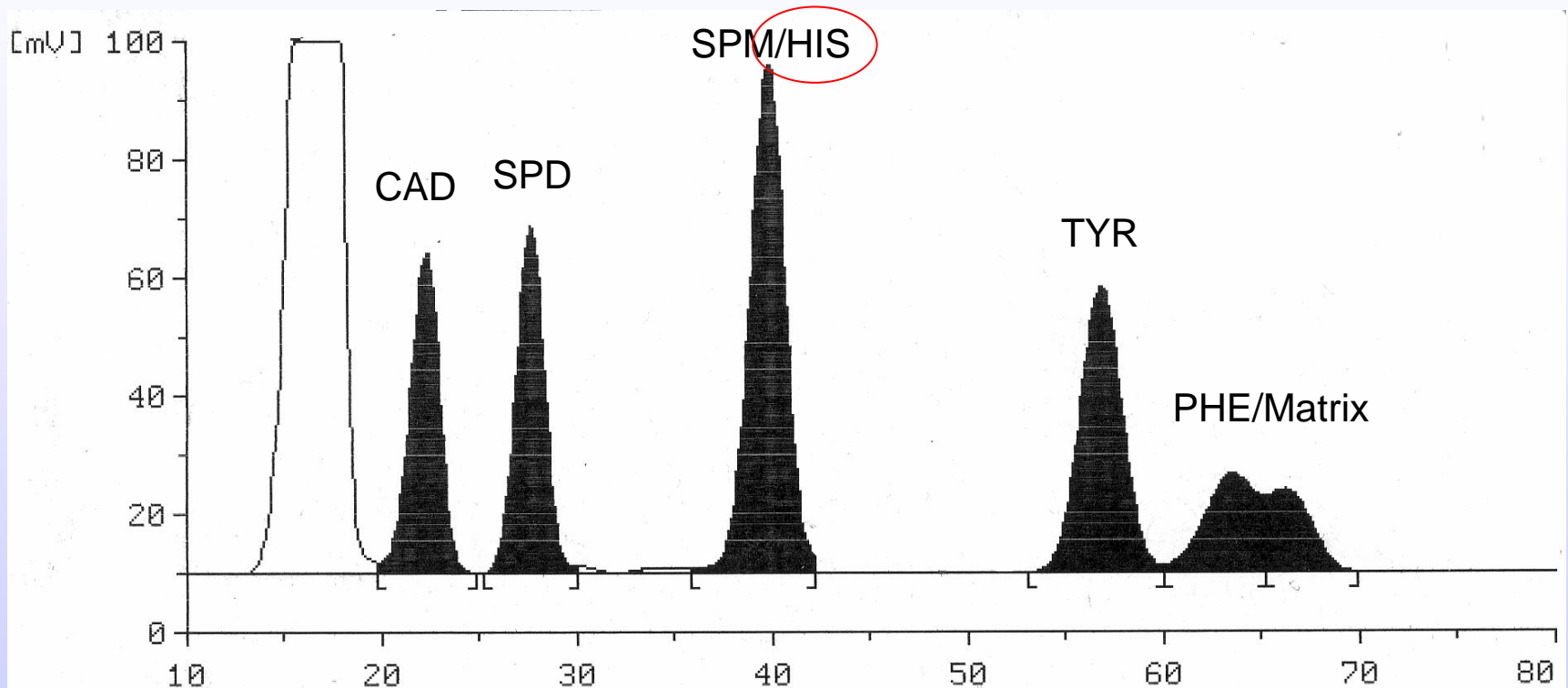
benzene : chloroform : triethylamine (10:6:7)

Densitogram of a spiked fish extract



benzene : chloroform : triethylamine (10:6:7)

Densitogram of a spiked fish extract

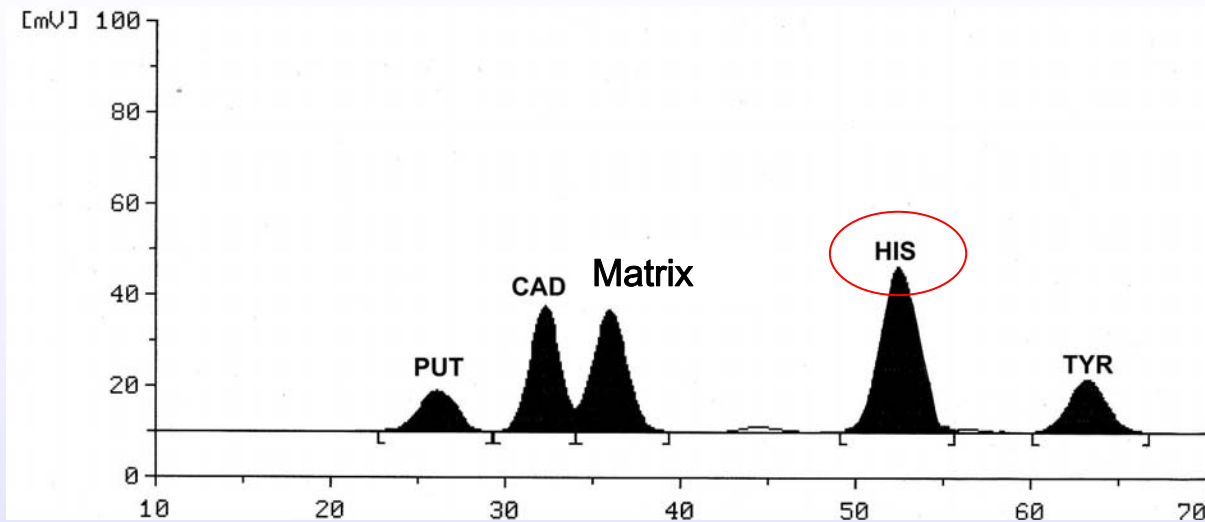


benzene : chloroform : triethylamine (10:6:2)

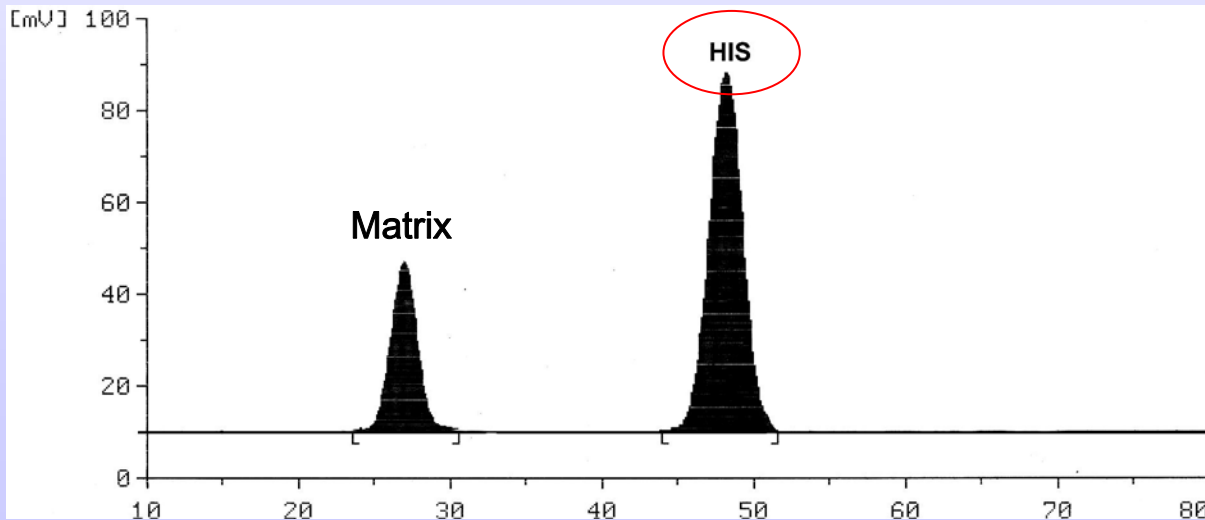
Validation data for histamine

Linearity	50 – 250 mg/kg
Recovery (200 mg/kg)	108 %
Confidence interval (n = 5, P = 0,95)	± 9.9 mg
LOD _{DIN 32645}	17.5 mg/kg
LOQ _{DIN 32645}	56 mg/kg

Densitograms of two fish samples



“Mackerel smoked”
178 mg/kg histamine



“Tuna in oil”
2804 mg/kg histamine

Histamine contents using different methods

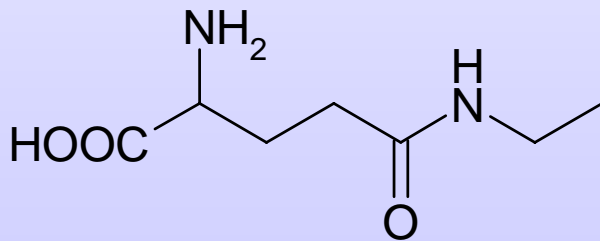
	TLC	HPLC	ELISA	Fluorimetry
Mackerel, smoked	178	174	251	-
Salmon, smoked	n.d.	<5	<1	1
Tuna in oil	2804	2541	2893	-
Tuna (Interlaboratory tests)	39	-	-	37

Contents of histamine in mg/kg

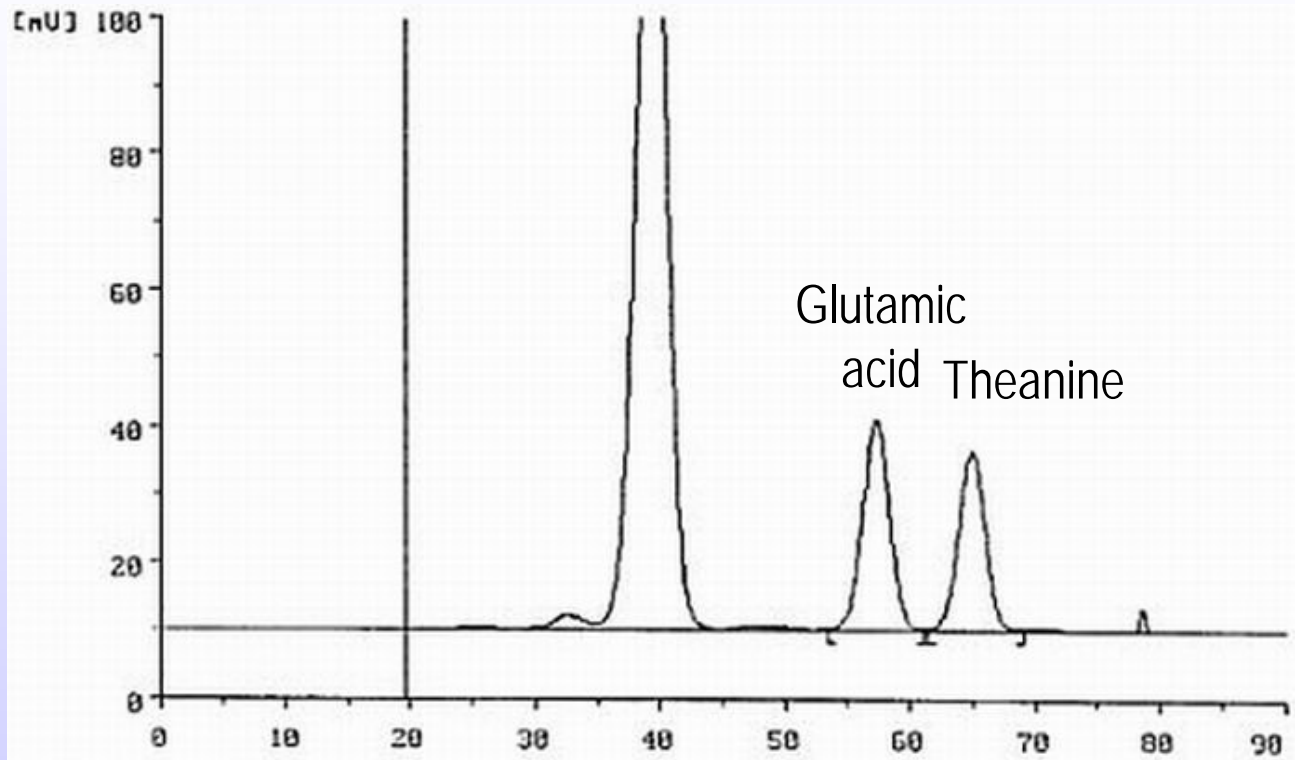
Theanine



- characteristic constituent of tea
- 50 % of the total free amino acids
- theanine content in tea leaves
c. 0.2 - 2 %
- sweeter taste
- pleasant tranquilising or gentle
freshening effect



Densitogram of a standard mixture



chloroform:methanol:glacial acetic acid (75:20:5)

Detection

- Excitation wavelength of dansylated amino acids

(Literature data): 250 - 370 nm;

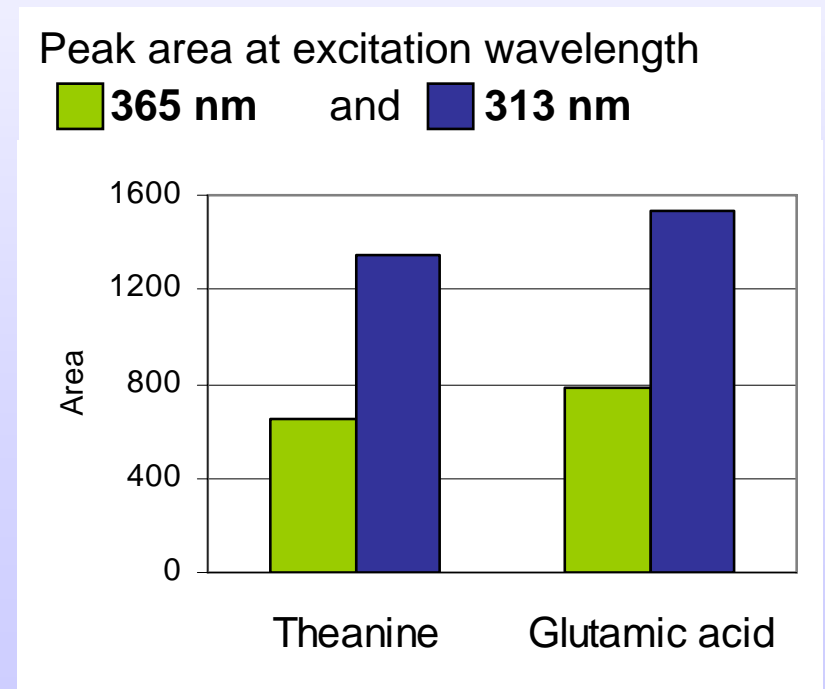
CAMAG: 313 nm

- Kretzschmar et al.:

365 nm for histamine

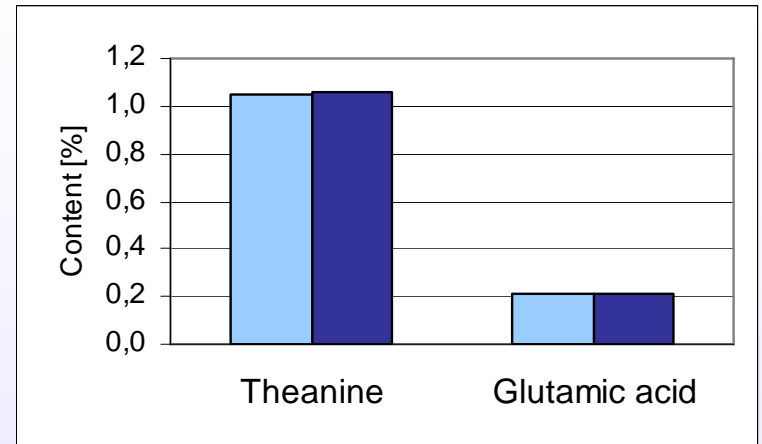
- **Detection**

> 400 nm

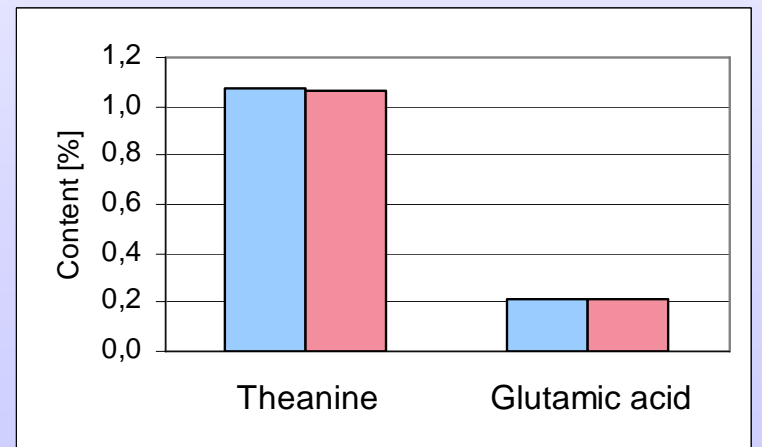


Sample preparation:

0.2 g of ground tea
are extracted with cold
water (15 ml)
(10 min in an ultrasonic bath)

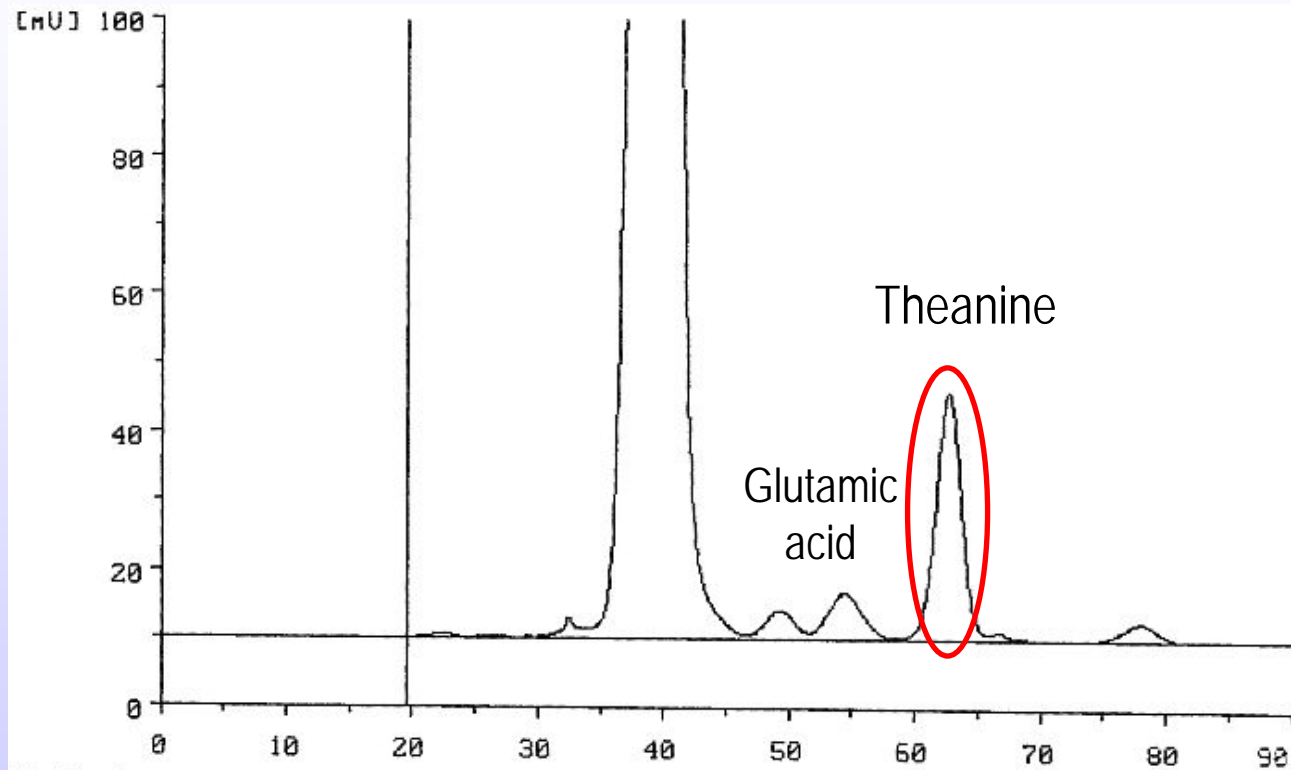


■ single extraction
■ twofold extraction



■ cold water extraction
■ hot extraction

Densitogram: black tea, Nepal

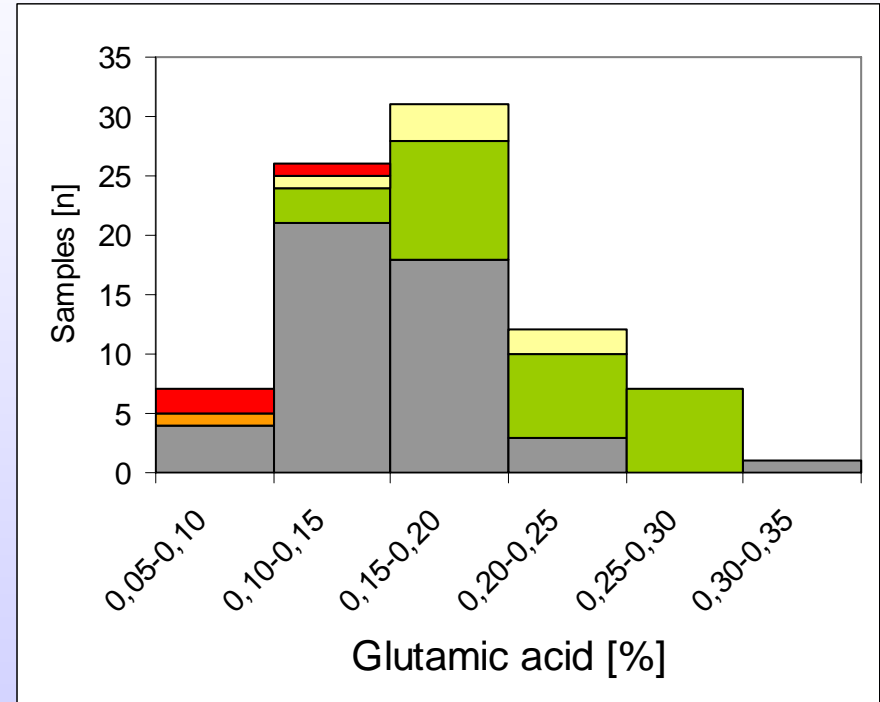
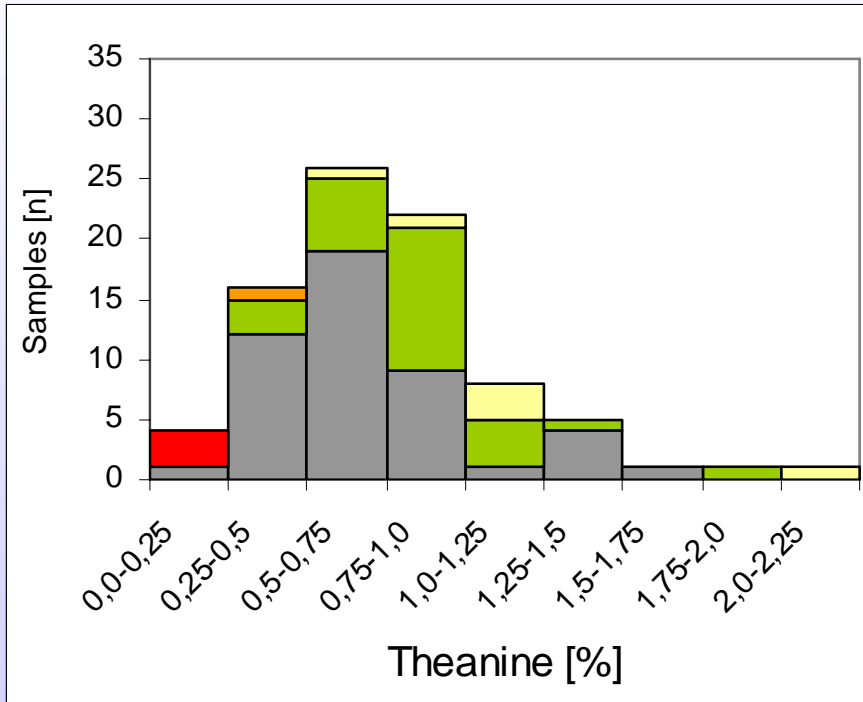


Validation data

	Theanine	Glutamic acid
Linearity	0.045 – 3.75 %	
Recovery	94 %	99 %
Confidence interval (n = 5, P = 0,95)	± 0.02 %	± 0.01 %
LOD*	0.10 %	0.08 %
LOQ*	0.35 %	0.28 %

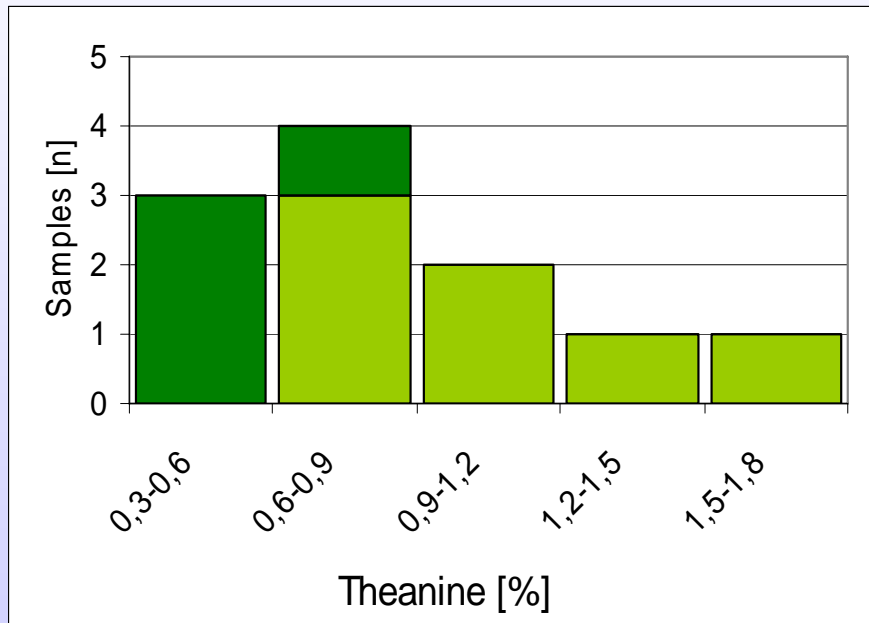
* DIN 32645

Contents of theanine and glutamic acid in different teas

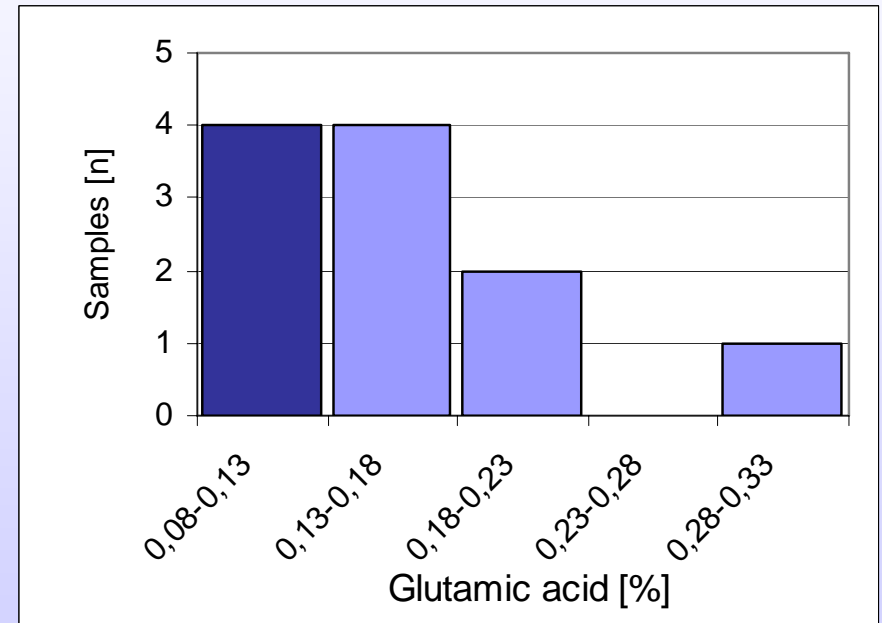


- Pu-Erh Tea (n = 3)
- Oolong Tea (n = 1)
- White Tea (n = 6)
- Green Tea (n = 27)
- Black Tea (n = 47)

Contents of theanine and glutamic acid in first flush and second flush teas

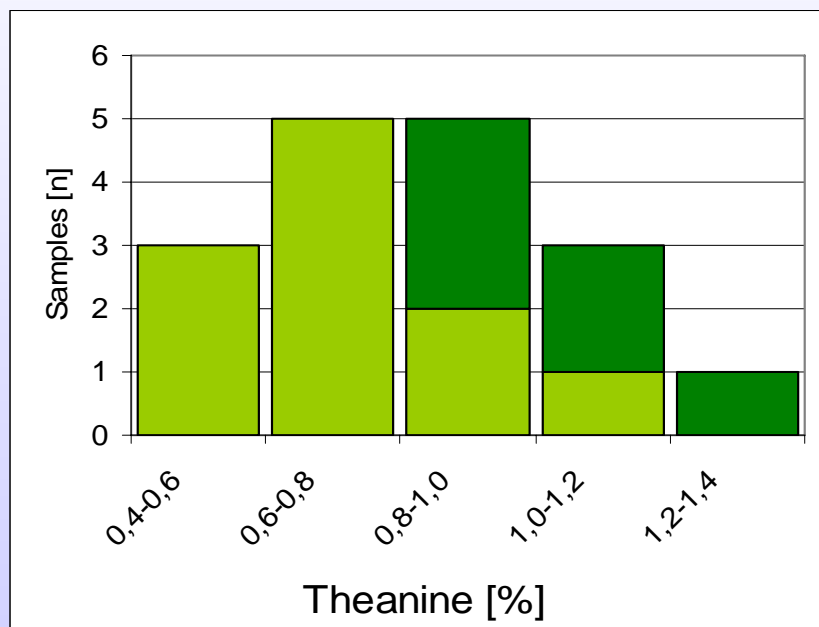


■ First Flush (n=7)
■ Second Flush (n=4)

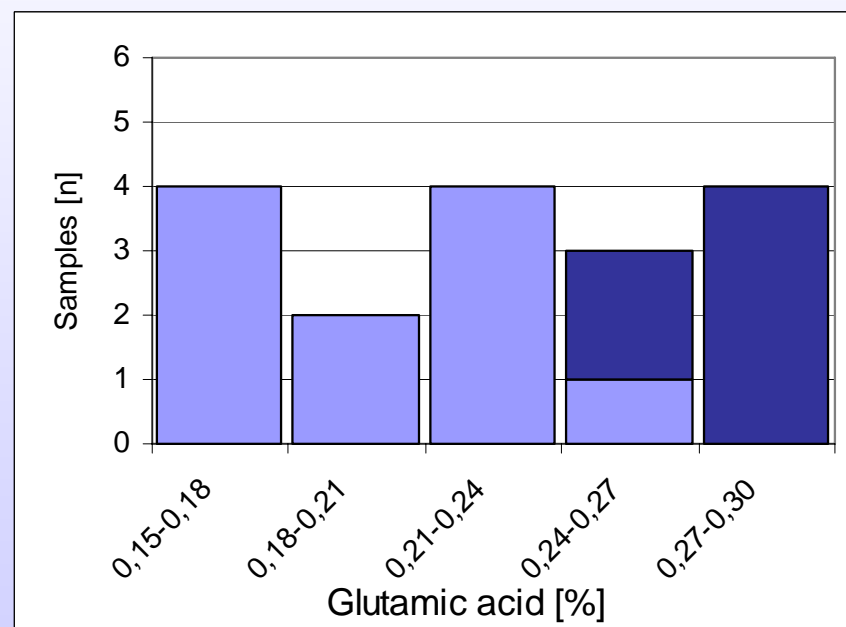


■ First Flush (n=7)
■ Second Flush (n=4)

Contents of theanine and glutamic acid in Japanese teas and Chinese teas

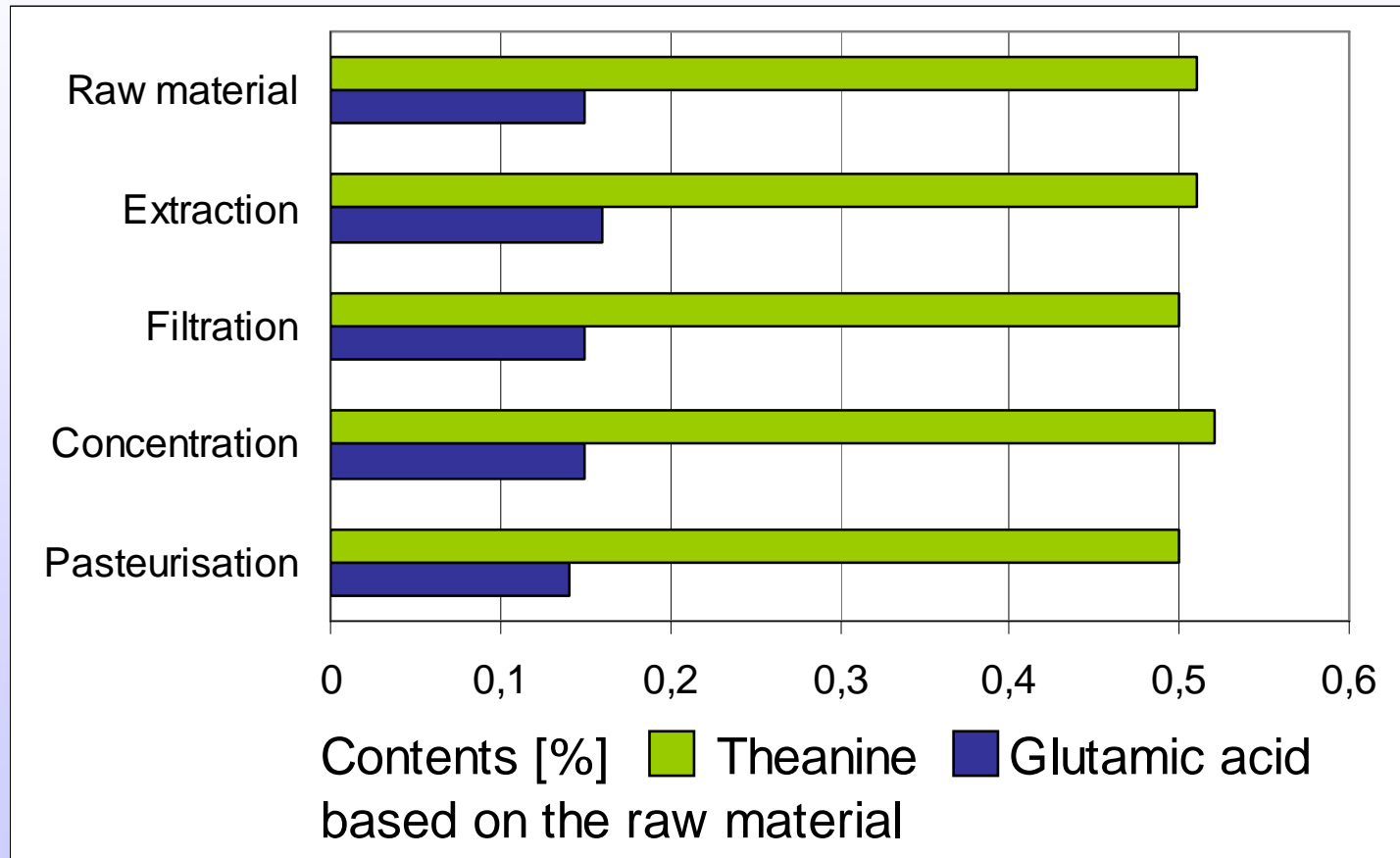


■ China (n = 11)
■ Japan (n = 6)



■ China (n = 11)
■ Japan (n = 6)

Behaviour of theanine and glutamic acid during the production of tea extract



Summary

Histamine in fish

12 samples can be analyzed
within 6 hours

Theanine in tea

24 samples can be analyzed
within 8 hours

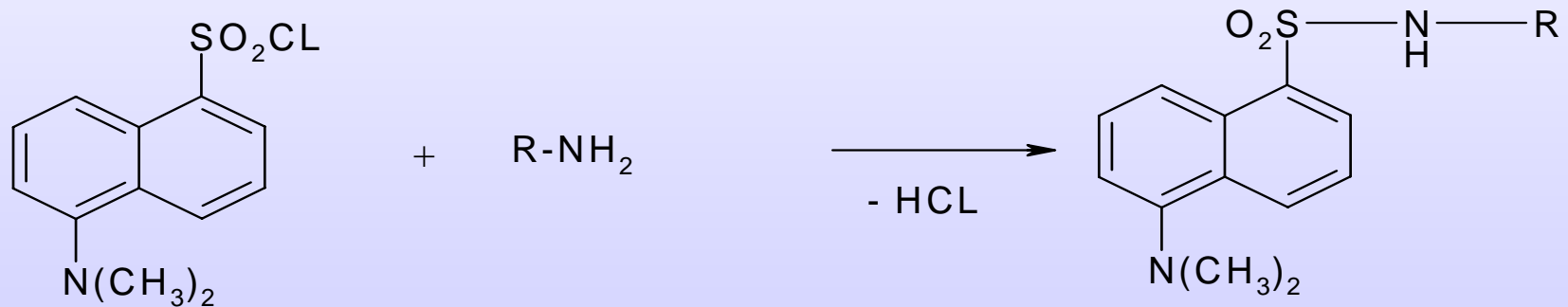
Derivatisation

1 ml of the filtrate is adjusted to pH 8

Add 1 ml borate-puffer and 2 ml dansyl chloride solution

Incubate for 1 h

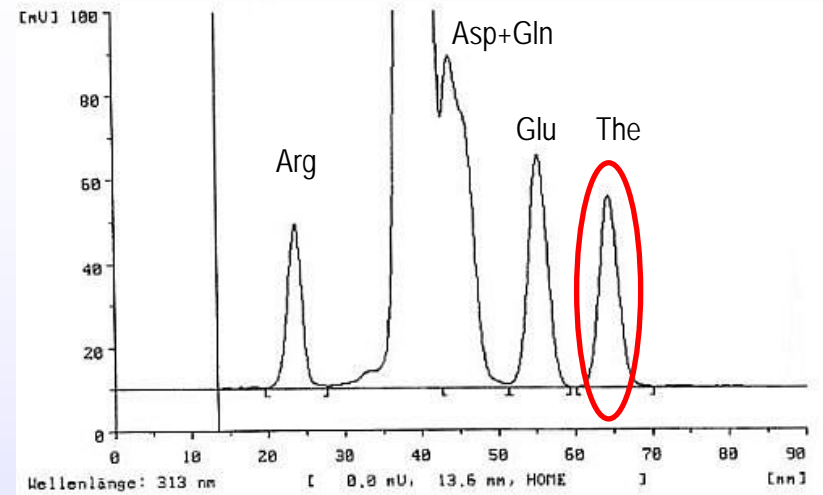
Adjust to 10 ml with water



Borate puffer

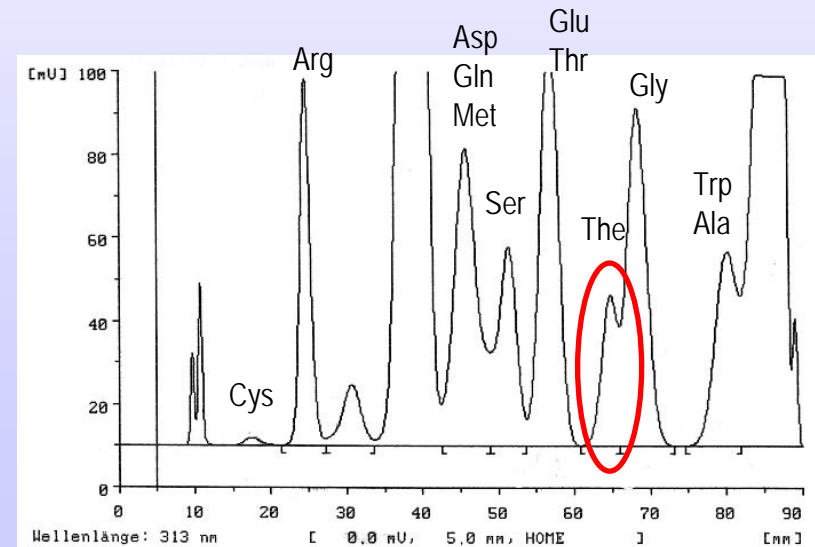
Mix 67.3 ml of a 0.25 M borax solution with 32.7 ml of a 1 M boric acid, for preparing 100 ml you need 6.225 g boric acid and 9.525 g sodium borate

- Trennung der wichtigen AS
The, Arg, Asp, Glu und Gln



Trennung der Aminosäuren Arg, Asp, Glu, Gln und The

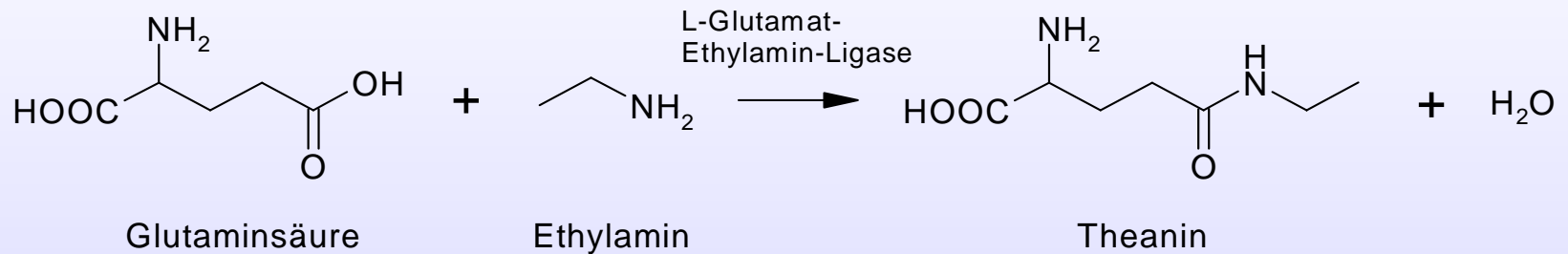
- weitere Aminosäuren:
 - Elution in ausreichendem Abstand zu Theanine (außer Gly)
 - Coelution von Glu und Thr



Trennung eines Aminosäuregemisches

$$BAI = \frac{c_{\text{Histamin}} + c_{\text{Putrescin}} + c_{\text{Cadaverin}}}{1 + c_{\text{Spermin}} + c_{\text{Spermidin}}}$$

Theanine biosynthesis



- in den jungen Wurzeln der Teepflanze
 - danach Transport in die jungen Blätter
- (Wickremasinghe et al. 1972)